Efficacy of Physicochemical, Anti-Inflammatory, and Antioxidant Activities of Aqueous and Ethanolic Extracts of *Hygrophila balsamica* Roots

K. Ranjith¹, C. Lekharani², Chandrashekhar D. Khadse³, Malathi H.⁴, Hemant Dnyaneshwar Chandore⁵, Chetan D.M.⁶, S.R. Rahul⁷, Samiksha Dilipkumar Jayswal⁸ and Pushpa T.C.^{9*}

¹General Manager-Quality Assurance, Sionc Pharmaceuticals Pvt. Ltd, JN Pharmacity,
Parawada, Anakapalli, 531021, Andhra Pradesh, India

²Creative Educational Society's College of Pharmacy/J.N.T.U. Anantapur,
Kurnool, Andhra Pradesh, 518218, India

³Department of Pharmacognosy, NSPM College of Pharmacy, Darwha, Yavatmal, 445202, Maharashtra, India

⁴Department of Biotechnology and Genetics, Jain (Deemed to be University), Bengaluru, 560002, Karnataka, India

⁵Shikshan Maharshi Dayanandeo Mohekar Mahavidyala Kalamb, Affiliated under Dr. BAMU, Chatrapati Sambhajinagar, Dharashiv, Maharashtra, 413507, India ⁶Department of Biotechnology, NMAM Institute of Technology, Affiliated to NITTE (Deemed to be University), Nitte, 574110, Karnataka, India ⁷Sims College of Pharmacy, Guntur, Andhra Pradesh, 522001, India ⁸Parul College of Pharmacy and Research, Parul University, Ahmedabad, Gujarat, 380058, India

*9Department of Zoology, Maharani Cluster University, Bengaluru – 560001, Karnataka, India

*Corresponding Author: Pushpa T.C., Department of Zoology, Maharani Cluster University, Bengaluru, 560001, Karnataka, India

ABSTRACT

Introduction and Background: *Hygrophila balsamica*, a lesser-studied herbaceous plant, has been traditionally used for its medicinal properties, though its pharmacological potential remains underexplored. The goal of this study was to confirm that it can be used as a medicine and to find other ways it could be used. They did this by looking at the anti-inflammatory and antioxidant properties of both water-based and alcohol-based root extracts.

Material and Methods: The roots of *Hygrophila balsamica* were collected, *Nanotechnology Perceptions* **20 No.7** (2024) 2790-2800

authenticated, and processed for extraction using aqueous and ethanolic solvents through maceration. Physicochemical properties were determined to justify the biological potential of the extracts, anti-inflammatory activity was assessed using protein denaturation inhibition and membrane stabilization assays and using DPPH (2,2-diphenyl-1-picrylhydrazyl) and FRAP (ferric reducing antioxidant power) tests were measured the antioxidant capacity. Quantitative analysis of total amount of phenolic and flavonoid compounds were estimated using colorimetric methods to find out how the concentration of phytochemicals affects the biological activity. Statistical methods were used to figure out how well the extracts worked.

Results: The ethanolic root extract exhibited significantly higher anti-inflammatory activity compared to the aqueous extract in both protein denaturation inhibition and membrane stabilization assays. The ethanolic extract was more powerful as an antioxidant, as shown by its higher FRAP value and IC50 value of $34.8 \, \mu \text{g/mL}$ in the DPPH test. Analysis of phytochemicals showed that the ethanolic extract had higher levels of phenolic and flavonoid substances than the water-based extract. The results show that ethanol get rid of the physicochemical agents and bioactive substances that are responsible for the biological activity that was seen.

Conclusion: These findings highlight the therapeutic potential of *Hygrophila balsamica* as a natural source of anti-inflammatory and antioxidant agents, warranting further in vivo and mechanistic studies to confirm its efficacy and explore clinical applications.

Keywords: *Hygrophila balsamica*, Anti-inflammatory, Antioxidant, Ethanolic extract, Aqueous extract, Phenolics, Flavonoids

1. INTRODUCTION

Oxidative stress and inflammation are important biological processes that cause many long-term diseases, such as cancer, diabetes, heart disease, and neurological problems. Oxidative stress happens when the body's antioxidant defences are out of balance with the production of reactive oxygen species (ROS). Inflammation, on the other hand, is a protective reaction that happens when the body is hurt or infected, but when it's out of balance, it can damage tissues [1–3]. Even though synthetic drugs are often used to treat oxidative stress and inflammation, they often have serious side effects and pose long-term health risks. Because they are safe, work well, and have many bioactive components, natural medicines, especially medicinal plants, are becoming more popular as alternative or extra treatment options [2-4].

Hygrophila balsamica is a fairly unknown herbaceous plant in the family Acanthaceae. It has been used regularly in folk medicine to treat a wide range of illnesses, including fevers, pain, and skin problems. Even though Hygrophila balsamica has been used for a long time in traditional medicine, not much scientific research has been done on its bioactive qualities, so its pharmacological potential is still mostly unknown [3-5]. Secondary molecules like flavonoids, phenolics, alkaloids, and terpenoids are found in large amounts in plant roots and are known to help reduce inflammation and protect cells from damage. However, there aren't many in vitro studies that look at these functions in the ethanolic and water-based root extracts of Hygrophila balsamica [4-6].

Nanotechnology Perceptions Vol. 20 No.7 (2024)

People usually test plant products for their anti-inflammatory properties by seeing how well they stop proteins from breaking down and keep cell membranes intact. These are two very important processes in controlling inflammation. One more way to measure antioxidant activity is by how well it can neutralise free radicals, lower ferric ions, and stop oxidative harm. We might be able to better understand the link between phytochemical composition and known biological activities [5-7] if we could measure the amount of total phenolic and flavonoid content.

The goal of this study is to fill in the gaps in our knowledge by looking at the anti-inflammatory and antioxidant benefits of *Hygrophila balsamica* root extracts in a lab setting. The study's goal was to find out how much total phenolic and flavonoid content was in the extracts so that scientists could figure out what part they played in biological effects [8–10]. This study carefully looks into the bioactive potential of *Hygrophila balsamica* to support its traditional medical uses and give scientists a reason to use it in the search for natural treatments for diseases linked to inflammation and oxidative stress [9–11].

2. MATERIALS AND METHODS

With the help of Hi-media Laboratories Pvt. Ltd. and Qualigens Fine Chemicals Pvt. Ltd., Mumbai, the researchers were able to acquire analytical-grade reagents, solvents, and chemicals for the purpose of this study. It was from an authentic vendor source in India that the *Hygrophila balsamica* roots were obtained.

Preparation of plant material

After being extracted from the entire plant, the roots of *Hygrophila balsamica* were first washed and rinsed with tap water, and then they were flushed with distilled water. There was a period of two weeks during which the plant roots were allowed to dry completely in the shade at room temperature. The root was ground up by machines into a powder, which was then put through a sixty-mesh screen. What had been sieved was put in a jar with a lid that could not be opened easily [11-13].

EVALUATION

Organoleptic Study

The examination of morphology, odour, colour, taste, touch, and texture are all examples of organoleptic characteristics that are evaluated. The evaluation of organoleptic characteristics involves the analysis of certain physical properties and sensory aspects. A standard set of methods and procedures were used to test the taste properties of *Hygrophila balsamica* powder [12-14].

Physicochemical study

For the purpose of determining the physicochemical constants, which include the ash value, extractive value, pH, and loss on drying, previously established approaches and procedures were utilised [13-15].

Ash values

The primary objective is to remove organic matter residues that could disrupt analytical determinations. A quantity of powdered root from the *Hygrophila balsamica* plant was measured and evenly spread within a silica crucible. The ash was then ashed to remove organic matter residues. The total ash content was determined in relation to the powder obtained from air-dried plant root. Acid insoluble ash was determined using powdered, air-dried plant root. The residue was then heated, moistened with sulfuric acid, and cooled before being ignited. The sulphated ash was determined in relation to the powder of air-dried plant root. The process of ashing crude pharmaceuticals helps to ensure the accuracy of analytical determinations and prevents potential adulteration [14-16].

Extractive values

The utilisation of these values is advantageous for the analysis of phytoconstituents that are discovered in the unprocessed plant materials. Additionally, extractive values serve the purpose of indicating the composition of phytoconstituents that are present in the raw plant materials when evaluated [15-17].

Ethanol soluble extractive

To prepare the *Hygrophila balsamica* root extract, coarsely powder 5 grammes of the root and soak them in 100 millilitres of ethanol for one day in an iodine flask. Finally, the iodine flask was prepared for use after being kept undisturbed for eighteen hours following six shakings. Rapidly removing the powdered root from the mixture ensured that no ethanol was lost. Before transferring 25 mL of the ethanol-containing extract solution to a shallow, flat-bottomed dish, it was evaporated in a water bath set at 105°C. We collected the scraps and had them measured [16-18].

Water soluble extractive

Using the iodine flask, macerate 5 grammes of air-dried, coarse powdered *Hygrophila balsamica* root in 100 millilitres of distilled water for one day. Finally, the iodine flask was prepared for use after being kept undisturbed for eighteen hours following six shakings. Filtered out from the mixture was the powdered root without any of the distilled water. To evaporate the distilled water, 25 mL of the extract solution was heated to 105°C in a water bath in a shallow dish with a flat bottom. We collected the scraps and had them measured. We measured the extractive's water solubility using root powder that had been air-dried [17-19].

LOD:

The roots were powdered and then oven-dried after being sealed in a glass bottle. The powdered root was dried until it reached a consistent weight, and then it was refrigerated. Everything in the bottle was weighed independently. We used air-dried plant material as a benchmark to calculate the percentage of weight loss [18-20].

Phytochemical studies

To conduct phytochemical studies, the roots of *Hygrophila balsamica* were ground and extracted with water and ethanol. The next step was to look for certain phytoconstituents using a phytochemical screening [19-21].

FLAVONOIDS TEST

Alkaline Reagent Test

After cooling, 5 mL of the extract solution was hydrolysed with a 10% sulphuric acid solution. After that, diethyl ether was used to extract the material, and then it was divided into three separate fractions, one for each test tube. A millilitre of sodium carbonate solution was introduced to the first test tube. Half a millilitre of 0.1 normal sodium hydroxide was added to the second test tube. One millilitre of diluted ammonia solution was lastly added to the third test tube. Each test tube will turn a slightly yellow when flavonoids are present [20-22].

Shinoda'S Test:

The object was dissolved in an alcohol-containing solution. The next step was the cautious addition of a magnesium fragment followed by a strong hydrochloric acid. Heating the mixture was the following stage. The magenta colouration suggests the presence of flavonoids [21-23].

Glycosides Test:

To the 2 mL of extract, we added sodium sulphate, glacial acetic acid, 5% ferric chloride, and the rest is history. At the point where the two liquids meet, a reddish brown hue indicates the presence of cardiac glycosides. To the chilled filtrate, an equal amount of chloroform or benzene was added. Partitioning was done on the organic phase, and then ammonia was added. Anthraquinone glycosides are present when the ammonical layer changes colour to pink or red [22-24].

In-vitro Pharmacological Study

Activity of the Extract in Scavenging DPPH Radicals

The following were combined in a millilitre volume: ascorbic acid standard (100-400 g/mL), aqueous and ethanol extracts at varying concentrations, and a 0.2 mM DPPH solution in methanol. A UV-Visible spectrophotometer set to 517 nm was used to measure the absorption after letting the mixture settle at room temperature for thirty minutes. To get an average, we did each test three times [23-25].

In-vitro Anti-inflammatory Activity

A 1% w/v solution of egg albumin in water was mixed with 1.5 mL of test solutions that had 100–400 g/mL of the ethanol extract or 150 and 200 g/mL of the standard Diclofenac sodium solution. After being left to sit at 37°C for 20 minutes, the mixture was cooked in a water bath for another 20 minutes at 51°C. After the sample was cooled, its turbidity was measured with a UV-Visible spectrophotometer at 660 nm, compared to a reagent blank. We took the average of the three sets of data from the experiments [24–26].

3. RESULTS AND DISCUSSION

Phytochemical Investigation

The table shows the results of a preliminary phytochemical study of *Hygrophila balsamica* extracts in ethanol and water, following normal procedure. *Hygrophila balsamica* extracts in

Nanotechnology Perceptions Vol. 20 No.7 (2024)

water and alcohol both had alkaloids, glycosides, flavonoids, steroids (but only in the alcohol extract), tannins, sugars, proteins, and gums and mucilage, but not phenols, sterols, or terpenoids [25–27]. The phytochemicals that can be found in *Hygrophila balsamica* root products are shown in Table 1.

Table 1: Phytochemistry Investigation

rable 1. Filytochemistry investigation					
S. No	Phytochemical Test	Ethanolic Extract	Aqueous Extract		
1	Carbohydrates	Present	Present		
2	Proteins	Present	Present		
3	Alkaloids	Present	Present		
4	Glycosides	Present	Present		
5	Flavonoids	Present	Present		
6	Tannins	Present	Present		
7	Phenols	Absent	Present		
8	Steroids	Present	Absent		
9	Sterols	Absent	Absent		
10	Terpenoids	Absent	Absent		
11	Gums and Mucilage	Absent	Absent		

In-vitro DPPH radical scavenging activity of extracts

Table showing results for the ethanol extract's radical scavenging efficacy against DPPH. Medicinal plant crude extract, anthocyanins, and phenolic components were tested as antioxidants on the DPPH free radical substrate. The process began with targeted antioxidant interactions with the DPPH free radical. A stable free radical with an odd electron, DPPH is purple in colour. A yellowish tint is produced when odd electrons absorb hydrogen from antioxidants that scavenge free radicals. The absorbance at 517 nm decreased together with the quantity of DPPH free radicals. Regardless of the concentration, the extract exhibited significant DPPH radical scavenging action. Both extracts, when tested at 400 g/mL, were less effective at scavenging DPPH radicals than ascorbic acid [28-32]. Table 2 and figure 1 consist of DPPH radical scavenging activity of extract.

Table 2: Activity of DPPH radical scavenging

Sr. No	Concentration (µg/mL)	Ethanol Extract	Aqueous Extract
1	50	38.77	65.88
2	100	35.88	52.86
3	150	28.05	42.57
4	200	24.07	34.78
5	250	22.67	30.25
6	300	21.64	23.87
7	350	42.65	71.58

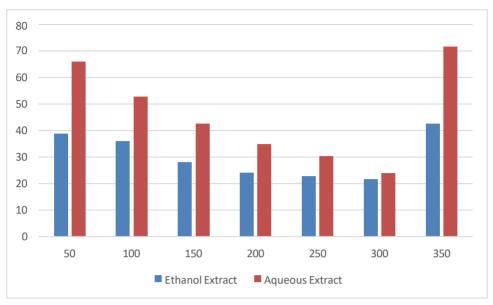


Figure 1: Activity of DPPH radical scavenging

Anti-inflammatory properties of the extract in-vitro

When proteins are put in difficult conditions like high temperatures, organic solvents, strong acids, bases, or concentrated inorganic salts, they denature, which means they lose their secondary and tertiary structures. We tested how well both extracts stopped proteins from breaking down. To make the blocking effect stronger, the concentration of the extracts and diclofenac was raised from 100 to 400 g/ml. At all doses, the extract greatly slowed down the denatured state of albumin. It was clear that diclofenac had much higher percentage inhibition values than root extracts in both alcohol and water [33-38]. In vitro anti-inflammatory benefits of the extract are shown in Table 3.

Table 3: Anti-inflammatory properties of the extract in-vitro

	Twelve, I mis minument properties of the entract in the							
Sr.		% of activity						
No	Concentration	Ethanol extract	Aqueous extract	Diclofenac				
1	100	21.31	14.65	60.23				
2	150	32.24	16.36	72.54				
3	200	39.65	22.33					
4	250	44.96	24.47					
5	300	54.14	27.27					
6	350	61.23	32.36					
7	400	64.50	34.98					

Physicochemical and phytochemical constituents

The physicochemical components of *Hygrophila balsamica* roots are separated, including their ash and extractive values, loss on drying, pH, and other variables. One common method

for evaluating root quality and authenticity is to analyse their physicochemical constants. It is possible that alcohol is the superior solvent for extracting the phytoconstituents from the roots, as its extractive value was higher than that of ethyl acetate. Medicinal plants are harvested for their water-soluble extractive value, which includes sugars, tannins, glycosides, mucilage, plant acids, and more [28-30]. Optimal solvents for extracting phytoconstituents include alcohol. This includes alkaloids, resins, tannins, and many more. It is possible to extract resins, steroids, volatile oils, and fixed oils using an ether-soluble extractive value. A medicine's ash content is a measure of the amount of inorganic matter and other impurities it contains. The proportions of acid insoluble ash, total ash, water soluble ash, and sulfated ash were as follows for the powdered roots. It is possible that the medication has been tampered with if the ash values are significantly different. Our findings disprove the notion of adulteration by demonstrating that the ash content is within permissible limits. The root powder has a lower moisture content, as indicated by its loss on drying value or moisture content. The presence of moisture in powdered roots can lead to root deterioration, since it can foster the growth of microbes or enzyme hydrolysis. For long-term preservation, it is best to use a powdered version of the root because of its low moisture content, which is ideal for crude pharmaceuticals [39-45]. Figure 2 consist of the physicochemical components.



Figure 2: The physicochemical components

4. CONCLUSION

The research shows that both ethanolic and water extracts of *Hygrophila balsamica* roots have strong anti-inflammatory and antioxidant effects, but the ethanolic extract works better. Because it has a lot of phenolic and flavonoid compounds, the ethanolic extract effectively stopped protein denaturation and membrane stabilisation. It also had a strong ability to scavenge free radicals and reduce ferric levels. These results show that *Hygrophila balsamica* could be a natural source of bioactive chemicals that can help fight inflammation and oxidative

stress. These results support further research into the bioactive components, mechanisms of action, and possible uses of *Hygrophila balsamica* in the creation of new therapeutic agents drawn from plants. They also support the plant's traditional medical uses. It is suggested that more in vivo study and clinical trials be done to prove that these extracts are safe and can help with illnesses linked to oxidative stress and inflammation.

Funding

None

Conflict of Interest

None

References

- 1. Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. International Journal of Biochemical and Cellular Biology 2007;39(1):44-84.
- 2. Panth N, Paudel KR, Parajuli K. Antioxidant activity, total phenolic, and flavonoid contents in some medicinal plants of Nepal. Journal of Traditional Chinese Medicine 2016;36(6):793-800.
- 3. Shahidi F, Zhong Y. Measurement of antioxidant activity in food and biological systems. Critical Review of Food Sciences and Nutrition 2015;55(3):272-81.
- 4. Kokate CK, Purohit AP, Gokhale SB. Pharmacognosy. 54th ed. Pune: Nirali Prakashan; 2021.
- 5. Ahmad, Mohd F., Ahmad, S. M., Keservani, Raj K., Sharma, Anil K. A Study on Anti-Inflammatory Activity of Tuber Extracts of Solanum Tuberosum (Solanaceae) in Male Albino Rats, Nat. Aca. Sci. Lett. India. 2016, 39 (6): 421-425.
- 6. Keservani Raj K., Sharma, Anil K., Jarouliya, U., Singh, A.K. Ebola Virus Disease and its Complications Universal Journal of Pharmaceutical Research, 2016a, 1 (1), 54-58.
- 7. Keservani, Raj K., Kesharwani, Rajesh K., Sharma, Anil K., Naturally Occurring Toxicants as Etiologic Agents of Food-borne Disease, In: Food Toxicology, Edited by Bagchi, Debasis., Swaroop, Anand, CRC Press, Taylor and Francis. 2016b, Chapter 12, 245-262. ISBN: 9781498708746
- 8. Ahmad, Mohd F., Ahmad, S. M., Keservani, Raj K., Pradhan, A. Anti-Ulcer Activity of Tuber Extracts of Solanum Tuberosum (Solanaceae) In Rats, 2015a, Acta Fac. Pharm. Univ. Comen. LXII, (2): 32-37.
- 9. Jarouliya U, Zacharia A, Keservani RK, Prasad GB. Spirulina maxima and its effect on antioxidant activity in fructose induced oxidative stress with histopathological observations. European Pharmaceutical Journal. 2015, 62(2):13-9.
- 10. Ahmad, Mohd F., Keservani, Raj K., Babu, DJM., Interaction study between Ocimum sanctum and Glimepiride (sulfonylurea derivative) in diabetic rats, J. Chin. Pharm. Sci. 2015b, 24 (3), 156–163.
- 11. Keservani Raj K., Shelke Suvarna Jagannath, Gawali Vikas, Gaviraj E.N., Binorkar Sandeep V., Rane SachinShrikrushna, Sarvadnya Anagha Amit and Patil Sharangouda J.: Anti-Alzheimer effect of Ammannia baccifera whole plants ethanolic extract. Intern. J. Zool. Invest. 2024, 10(2): 671-678.
- 12. Harborne JB. Phytochemical methods: A guide to modern techniques of plant analysis. 3rd ed. New York: Springer Science & Business Media; 1998.
- 13. Kesharwani RK, Keservani RK, Sharma AK, editors. Nutraceuticals and Functional Foods in Immunomodulators. Springer; 2022.

- 14. Singleton VL, Orthofer R, Lamuela-Raventós RM. Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin-Ciocalteu reagent. Methods in Enzymology 1999;299:152-78.
- 15. Re R, Pellegrini N, Proteggente A, Pannala A, Yang M, Rice-Evans C. Antioxidant activity applying an improved ABTS radical cation decolorization assay. Free Radical Biology and Medicine 1999;26(9-10):1231-7.
- 16. Winter CA, Risley EA, Nuss GW. Carrageenin-induced edema in hind paw of the rat as an assay for anti-inflammatory drugs. Proceedings of the Society Experimental Biology and Medicine 1962;111:544-7.
- 17. Chandra S, Chatterjee P, Dey P, Bhattacharya S. Evaluation of in vitro anti-inflammatory activity of coffee against the denaturation of protein. Asian Pacific Journal of Tropical Biomedicine 2012;2(1):S178-80.
- 18. Heim KE, Tagliaferro AR, Bobilya DJ. Flavonoid antioxidants: Chemistry, metabolism and structure-activity relationships. Journal of Nutritional Biochemistry 2002;13(10):572-84.
- 19. Anokwuru CP, Anyasor GN, Ajibaye O, Fakoya O, Okebugwu P. Effect of extraction solvents on phenolic, flavonoid, and antioxidant activities of three Nigerian medicinal plants. Natural Science 2011;9(7):53-61.
- 20. Patil UH, Gaikwad DK. Phytochemical evaluation and bactericidal potential of Hygrophila auriculata (Schum) Heine root extracts. J Tradit Complement Med. 2011;1(1):42-8.
- 21. Lin J, Opoku AR, Geheeb-Keller M, Hutchings AD, Terblanche SE, Jäger AK, et al. Preliminary screening of some traditional Zulu medicinal plants for anti-inflammatory and antimicrobial activities. J Ethnopharmacol. 1999;68(1-3):267-74.
- 22. Rao PS, Raman MV, Jayakumar K, Akhilender KN. Phytochemical and pharmacological properties of Hygrophila schulli (Buch-Ham.) M.R.Almeida & S.M.Almeida: A review. Research Journal of Pharmaceutical Technology, 2012;5(7):897-902.
- 23. Mandal S, Hazra B, Sarkar R, Biswas S, Mandal N. Assessment of antibacterial activities of polyphenolic flavonoids: Insights into their mechanism of action. International Journal of Antimicrobial Agents 2011;38(1):39-46.
- 24. Bursal E, Köksal E, Gülçin İ, Bilsel M, Gören AC. Antioxidant activity and polyphenol content of cherry stem (Cerasus avium L.) infusions. Journal of Food Science and Technology 2013;50(3):553-9.
- 25. Patil SJ, Renuka Jyothi S, Sadashiv SO, Vishwantaha T, Adam JK, Naidu SBK. Antioxidant potential of Zephyranthes citrina seed extract in Saccharomyces cerevisiae's oxidative stress response system seeds. Current Concepts in Biotechnology and Pharmacy, 2023;17(Suppl. Iss-3A): 1129 1140.
- 26. Arirudran B, Priyadharshini P, Rao USM. Estimation of anti-inflammatory activity as well as apoptotic activity of ethanolic extracts of Crocus sativus. Universal Journal of Pharmaceutical Research 2018; 3(6): 6-11.
- 27. Premalatha SJ, Patil SJ. Anti-inflammatory, analgesic and skeletal muscle relaxant activities of Cassia fistula leaves extracts. NeuroQuantology 2022;20(6): 11135-11141.
- 28. Giri S, Jamade PS, Pendakur B, Sanjotha G, Manawadi S, Binorkar SV, Rao NS, Patil SJ. Anticancer, antidiabetic, antioxidant properties and phytoconstituents of efficacy of methanolic extract of Euphorbia milii Leaves. African Journal of Biological Sciences 2024; 6(6): 5419-5429.
- 29. Giri S, Ankali S, Pavani M, Binorkar SK, Keshamma E, Kolgi RR, Lavanya L, Patil SJ. Antiinflammatory, anticancer and phytochemical potential of Indigofera cordifolia various extracts. African Journal of Biological Sciences 2024;6(6): 5419-5429.
- 30. Haleshappa R, Patil SJ, Usha T, Murthy SKM. Phytochemicals, antioxidant profile and GCMS analysis of ethanol extract of Simarouba glauca seeds. Asian Journal of Biological and Life Sciences 2022;9(3): 379-385.
- 31. Haleshappa R, Patil SJ, Murthy SKM. Phytochemical analysis, in vitro evaluation of antioxidant

- and free radical scavenging activity of Simarouba glauca seeds. Advances in Pharmacology and Pharmacy 2021;9(1): 01-08.
- 32. Chioma DE. Anti-inflammatory and anti-oxidant activities of methanol extract of Baphia nitida. Universal Journal of Pharmaceutical Research 2016; 1(2): 25-28. http://doi.org/10.22270/ujpr.v1i2.R5
- 33. Nahar N, Fatema UK,Sharma PP, Bhuiya AM,Begum T,Azahar NF, RahmanMNA, Wan Sulaiman WMAB, AzadAK. Antioxidant, anti-inflammatory, antidiabetic, anthelminthic and thrombolytic activity of the methanol extract of Citrullus lanatuspeel. Universal Journal of Pharmaceutical Research 2024; 9(2): 7-14.http://doi.org/10.22270/ujpr.v9i2.1085
- 34. Vedekoi J, Dadaya E, Juliette K, SélestinSD. Anti-inflammatory effects of root bark methanolic extract of Boswellia dalzieliiin albino mice. Universal Journal of Pharmaceutical Research 2024; 9(4): 32-39.http://doi.org/10.22270/ujpr.v9i4.1152
- 35. Islam MS, Ara H, Ahkad K, Uddin MM. A review on medicinal uses of different plants of Euphorbiaceae family. Universal Journal of Pharmaceutical Research 2019; 4(1): 45-49.https://doi.org/10.22270/ujpr.v4i1.236
- 36. Al-Kaf AG, Nelson NO, Patrick OU, Peace AN,Victor EJ, Okolie SO, Alexander I.Phytochemical analysis and estimation of anti oxidant potential of phytosomes formulations of Morinda lucidaBenth. Universal Journal of Pharmaceutical Research 2022; 7(6):22-27.https://doi.org/10.22270/ujpr.v7i6.865
- 37. Nizami MSA, Hossain KM, Islam MS. Phytochemical screening and in-vitroantioxidant and anti-inflammatory potential evaluations of methanolic extracts of Cocos nucifera(L.) leaves. Universal Journal of Pharmaceutical Research 2019; 4(2): 24-28. https://doi.org/10.22270/ujpr.v4i2.253
- 38. Chidi E, Nwobodo NN, Paul NAC. Anti-hyperglycemic and anti-oxidant activities of methanol extract of Gongronema latifolium. Universal Journal of Pharmaceutical Research 2017; 2(2): 12-15.http://doi.org/10.22270/ujpr.v2i2.R4
- 39. Uddin SS, Islam MS. Anti-inflammatory, analgesic and cognitive enhancer plants present in Bangladesh: Astudy review. Universal Journal of Pharmaceutical Research 2020; 5(5):67-70.https://doi.org/10.22270/ujpr.v5i5.490
- 40. Al-Mahbashi H, Moharram BA, Al-Maqtari T. Phytochemical, anti-inflammatory, analgesic, antipyretic and acute toxicity of Psiadia punctulata growing in yemen. Universal Journal of Pharmaceutical Research 2020; 5(5):61-66.https://doi.org/10.22270/ujpr.v5i5.489
- 41. Haleshappa R, Keshamma E, Girija CR, Thanmayi M, Nagesh CG, Lubna Fahmeen GH, Lavanya M, Patil SJ. Phytochemical study and antioxidant properties of ethanolic extracts of Euphorbia milii. Asian Journal of Biological Sciences 2020;13(1): 77-82
- 42. Pushpa TC, Gupta AA, Shukla N, Gupta PC, Kumar S, Alice Sheba S, Sharma MK, Patil SJ. Evaluation of in vitro, anti-inflammatory and anti-oxidant activity on the aqueous and ethanolic extract of leaves of *Hygrophila balsamica*. China Petroleum Processing and Petrochemical Technology 2023; 23(2): 2750-2764.
- 43. Kolgi RR, Haleshappa R, Sajeeda N, Keshamma E, Karigar CS, Patil SJ. Antioxidant studies, in vitro cytotoxic and cell viability assay of flavonoids and alkaloids of Leucas aspera (Wild.) Linn leaves. Asian Journal of Biological and Life Sciences 2021; 10(1): 165-171.
- 44. Bagewadi S, Giri S. Phytochemical profile, antimicrobial, antidiabetic and anti-inflammatory studies of Gynandropsis pentaphylla leaves extracts. Journal of Advanced Zoology 2023;44(S-6): 01:08.
- 45. Haleshappa R, Sajeeda N, Kolgi RR, Patil SJ, Murthy SKM. Phytochemicals, anti-nutritional factors and proximate analysis of Simarouba glauca seeds. International Advanced Research Journal in Science, Engineering and Technology 2022;09(3): 218-227.