Development And Validation Of Stability Indicating RP- HPLCMethod For Simultaneous Estimation Of Lobeglitazone And Metformin In Tablet Dosage Form

Bharatbhusan Sahu

Centurion University of Technology and Management, Odisha Email: - sahubharat378@gmail.com

Background: A simple, rapid, sensitive and selective stability-indicating (RP-HPLC) method is suggested for the determination of Lobeglitazone and Metformin in pharmaceutical formulation. Lobeglitazone and Metformin was eluted from a Cosmosphere C18 (250×4.6 mm, 5 μ m) column with mobile phase consisting of methanol, Acetonitrile and Potassium Dihydrogen Phosphate Buffer 2.5 pH (70:5:25 %v/v/v) pH adjusted to 2.5 with Orthophosphoric acid. The gradient was optimized with a flow rate of 0.8 mL/min and a wavelength of 250 nm.

Result: The complete analytical method validation was successfully carried out as per ICH guidelines. The retrieval study was carried out at 50% to 150% level of working concentration, and results were in the range of 98 to 102%. The linearity was proven in range of 0.5-0.25 $\mu g/ml$ $\mu g/mL$ of working concentration of Lobeglitazone and 50-250 $\mu g/ml$ for working concentration of Metformin with linear regression curve ($R^2=0.999$) with limits of detection (LOD) and quantitation (LOQ) being 0.0010 and 0.0031 $\mu g/mL$ for Lobeglitazone and 0.30 and 0.91 $\mu g/mL$ for Metforminrespectively. The retention time for Lobeglitazone was 8.37 min and for Metformin was 2.83 min. Themethod shows good recoveries and intra-day and inter-day relative standard deviations were less than 2%. Validation parameters as ruggedness and robustness were also determined as per ICH guidelines and were found to be satisfactory. For stability study, the drug was exposed to variousstress conditions such as acid, base, oxidation, Thermal and sunlight as per recommendations of ICH guidelines.

Conclusion: The developed HPLC method couldbe successfully used for the estimation of Lobeglitazone and Metformin in pharmaceutical formulation. The high recovery and low relative standard deviation confirm the suitability of proposed method that can be employed for the routine analysis in bulk and pharmaceutical formulation.

Key Words: Lobeglitazone, Metformin, RP- HPLC, Stability, Validation

Introduction

1. BACKGROUND OF LOBEGLITAZONE AND METFORMIN

Lobeglitazone IUPAC name 5-[(4-[2-([6- (4-Methoxyphenoxy) pyrimidin-4-yl]-methylamino)ethoxy]phenyl)methyl]-1,3- thiazolidine-2,4-dione.

Chemical formula C24H24N4O5S (Fig. 1). It is an anti-diabetic drugin the thiazolidinedione class of drugs. It primarily function as an insulin sensitizer by binding and activating

Peroxisome Proliferator-ActivatedReceptors (PPAR) gamma within fat cells. PPAR is a transcription factor that plays a role in regulatingmetabolism. By promoting the binding of insulin atfat cells, Lobeglitazone has been shown to reduceblood sugar levels, lower hemoglobin A1C levels, and improve lipid and liver profiles. MetforminIUPAC name N, N-Dimethylimidodicarbonimidic diamide. Chemical formula $C_4H_{12}ClN_5$ (Fig.2). It reduces glucose absorption from the intestines, lowers liver glucose production, and improves insulin sensitivity. Metformin is recommended with dietary changes and exercise for better results. Managing blood sugar levels with medications like metformin can prevent complications such askidney damage, nerve issues, blindness, and amputations. This combination approved by CDSCO in the year 2022 for the treatment of Type-2 Diabetes Mellitus and available as LOBG-M in the market.

Fig.1 Chemical structure of Lobeglitazone

Fig.2 Chemical structure of Metformin

FORCED DEGRADATION:

Forced degradation experiments are used to relieve the development of analytical methodology, to achieve better insightful of the stability of the active pharmaceutical ingredient (API) and the drug product, and to provide information about degradation pathways and degradation products. However, no literature is available for which deals with the stress degradation profile of Lobeglitazone and Metformin in accordance with ICH guidelines using any of the above analytical techniques. High performance liquid chromatography (RP-HPLC) for analysis of Lobeglitazone and Metformin in pharmaceutical formulation. This paper

describes an accurate, specific, repeatable, and stability- indicating method for analysis of Lobeglitazone and Metformin in the presence of its degradation products. The method was validated in accordance with the guidelines of International Conference on Harmonization (ICH).

Necessity and importance of stability-indicating method

The goal of the stabilization studies is to track potential improvements to a substance or material over time and under various storage conditions. The factors and parameters that affect the stability are production timeframe, batch factors along with process parameters, excipients efficiency, and environmental conditions like temperature and humidity.

The stability indicating method can be defined as Validated Quantitative analytical method that can detect the change with time in the chemical, physical or microbiological properties of the drug substance and drug product, and that are specific so that the content of active ingredient, degradation can be accurately measured without interference.

The precision of the stability methods showing potential impurities of the drug material and of drug components is demonstrated by forced degradation (FD). Stress experiments help to generate impurities in a much shorter period. The formulations scientist will then generate consistent formulations in less time. FD studies now include the completion of the file and the comprehension of the drug production mechanism for global controlled markets.

GMP includes a structured written monitoring program for stability, the results of which can be used to specify the storage requirements, the expiry dates and the use of accurate, meaningful and precise test procedures. If there is an effort to document drug product stability, the use of such approaches is acceptable. These data are being used to assess, conform or expand retest cycles or expiration date for the drug substance.

The rationale for the stability studies research is to provide data as to how the consistency of the substance varies over the time under the control of amultiplicity of ecological variables, such as humidity, temperature and light, allows the proposed storage conditions, re-analysis periods and shelf life.

METHOD DEVELOPMENT

Reagents and chemicals

Lobeglitazone was supplied as a giftsample by a Allastir Pvt. Ltd. Chennai andMetformin was by a Endoc Lifecare Pvt. Ltd. Gujarat. All the Chemicals used of (RANKEM, INDIA). Solvents and solutions were filtered through a membrane filter (0.45 μ m pore size) and degassed by sonication before use.

Instrumentation

The chromatographic analysis was performed on Waters Alliance HPLC system equipped with PDA detector. The output signals were monitored and processed using LC Solution software. The analytical column was Cosmosphere C18 (4.6 mm \times 250 mm, 5 μ) and the samples were introduced through an injection valve with 10 μ L sample loop.

Wavelength detection

25 mg of Lobeglitazone & 25 mg of Metformin take into 25 ml of volumetric flask separately and dissolved with diluent (Stock-1 Solution) (Lobeglitazone $1000\mu g/ml$ and Metformin $1000\mu g/ml$). From that 1ml in 10ml volumetric flask separately (Stock-2 Solution) (Lobeglitazone $100\mu g/ml$ and Metformin $100\mu g/ml$). From that 1ml in 10ml volumetric flask separately (Working standard Solution) (Lobeglitazone $10\mu g/ml$ and Metformin $10\mu g/ml$). UV Spectra was taken between range of 200- 400nm using UV- Visible Double beam spectrometer. Absorbance of both Lobeglitazone and Metformin was observed at 250nm and 234nm respectively.

Chromatographic conditions

Mobile phase selection involved selection of solvent, selection of buffer, pH of buffer and ratio of buffer and solvent. The standard solutions of Lobeglitazone and Metformin were injected into the HPLC system and run in different solvent system. Various ratios of mobile phase containing Methanol: Water, ACN: Water, Phosphate Buffer pH 4.0: Methanol, Phosphate Buffer pH 6.0: Methanol were tried in order to find the best conditions for the separation of both drugs. It was found that Methanol, ACN and Phosphate buffer pH 2.5 gives satisfactory result. Finally, Methanol: ACN: Potassium Dihydrogen Phosphate buffer pH 2.5 (70:5:25% v/v/v) ratio was optimized as the mobile phase for the determination. pH was set by using 1% orthophosphoric acid. Injection volume was $10\mu L$, flow rate was 0.8mL/min and the eluent were detected at 250 nm at column temperature 25°C. These conditions showed sharp peak of Lobeglitazone and Metformin with retention time of 8.37 min and 2.83 min respectively.

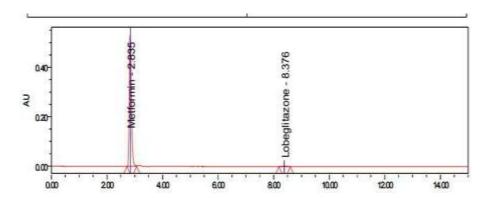
Preparation of stock standard solution and sample

Stock solution:

Weigh 5mg of Lobeglitazone and transferred it to 50ml of volumetric flask and makeup to the given markwith methanol (stock solution-1; $100\mu g/ml$) Furtherfrom stock-1, take 0.5 ml in 50ml flask and makeupwith methanol (Standard stock solution-2; $1\mu g/ml$). Weigh 50mg of Metformin. Transferred it to 50ml of volumetric flask and makeup to the given mark with methanol (Standard stock solution $1000\mu g/ml$). Take 1ml from Lobeglitazone standard stock-2 solution and 1 ml from Metformin stock solution into 10ml volumetric flask and makeup to the given mark with diluent. Lobeglitazone ($0.1\mu g/ml$) Metformin ($100\mu g/ml$).(Fig. 3)

Sample solution: (Label claim: Lobeglitazone- 0.5mg; Metformin-500mg)

Twenty tablets were weighed; average weight was calculated and tablets were powdered finely. Tablet Powder equivalent to 0.5mg of Lobeglitazone and 500mg of Metformin were added into 100ml of volumetric flask Lobeglitazone ($5\mu g/ml$) and Metformin ($5000\mu g/ml$). Volume was made up to the mark with Methanol. The solution is then sonicated for 20mins and further the solution is filtrated. 1ml of each above solution of Lobeglitazone and Metformin was transferred to 10ml volumetric flask. Volume was made up to the mark with diluent, which gives Lobeglitazone and Metformin.



3 Lobeglitazone (0.1μg/ml) and Metformin (100μg/ml) by using Methanol: ACN: Potassium Dihydrogen Phosphate Buffer pH 2.5 (70:5:25 % v/v/v) mobile phase

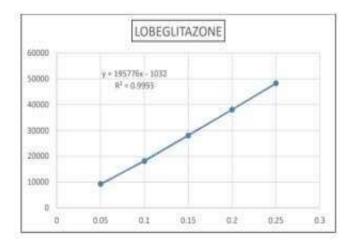
ANALYTICAL METHOD VALIDATION

1. Specificity:

Demonstration of specificity is required to show that the procedure is unaffected by the presence of impurities or excipients. Specificity of an analytical method indicates that the analytical method is its able to measure accurately and specifically the analyte of interest without any interference from blank. So here, the specificity was determined by the comparison of the chromatograms of

- Blank (mobile phase).
- Standard solutions Lobeglitazone and Metformin.
- Sample solution of Lobeglitazone and Metformin.

2. Linearity:



The linearity for Lobeglitazone and Metformin was assessed by analysis of standardsolution in range of 0.05- $0.25\mu g/mL$ for Lobeglitazone and $50-250\mu g/mL$ for Metformin. To obtain Lobeglitazone 0.05, 0.1, 0.15, 0.2, $0.25\mu g/mL$; 1, 0.5, 1.5, 2, 2.5ml is pipetted outfrom standard stock solution($1\mu g/mL$) into 10ml volumetric flask and further volume was adjusted with diluent to the mark. Similarly, to obtain Metformin 50, 100, 150, 200, $250\mu g/mL$; 0.5, 1, 2, 1.5, 2.5ml is pipetted out from standard stock solution($100\mu g/ml$) into 10ml volumetric flask and further volume was adjusted with diluent to themark. In term of slope, intercept and correlation co-efficient value, the graph of peak area obtained verses respective concentration was plotted. (Fig.4)

Acceptance criteria: value of r^2 should be nearer to 1 or 0.999.

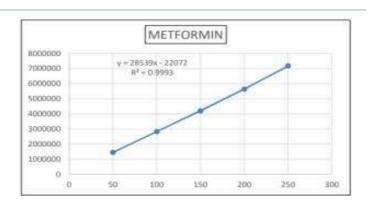


Fig.4 Calibration Curve of Lobeglitazone (0.05-0.25µg/ml) and Metformin (50-250µg/ml)

3. Precision:

Precision can be performed at two different levels: repeatability and intermediate precision. Repeatability refers to the use of the analytical procedure within the laboratory over the shorter

Nanotechnology Perceptions 20 No. S13 (2024)

period of the time that was evaluated by assaying the samples during the same day. Repeatability was carried out using six replicates of the sample injection. Intra-day precision was determined by analyzing, the three different concentrations for three times in the same day. Dayto day variability was assessed using above mentioned three concentrations analyzed on three consecutive days for inter-day precision. Results should be expressed as Relative standard deviation (RSD) or co-efficient of variance.

A. Repeatability:

Standard solution containing Lobeglitazone and Metformin (0.1 and $100\mu g/ml$ respectively) was injected six times and areas of peaks were measured and RSD was calculated.

B. Interday Precision:

Standard solution containing Lobeglitazone and Metformin (0.05, 0.1, 0.15 μ g/ml) and 50, 100, 150 μ g/ml respectively) were injected three times in same day and areas of peaks were measured and RSD was calculated

C. Intraday Precision:

Standard solution containing Lobeglitazone and Metformin (0.05, 0.1, 0.15 μ g/ml) and 50, 100, 150 μ g/ml respectively) were injected three times in different days and areas of peaks were measured and RSD was calculated.

Acceptance criteria: RSD of area should not be more than 2.0%.

4. Accuracy:

Preparation of Standard Stock Solution of Lobeglitazone and Metformin:

• Accurately weighed Lobeglitazone (5mg) was transferred into 50ml of volumetric flask and make up to the mark with diluent (Lobeglitazone $100\mu g/ml$). From this, transfer 1ml into 10 ml volumetric flask and make up to mark with diluent (Lobeglitazone $1\mu g/ml$). Accurately weighed Metformin (50mg) wastransferred into 50ml of volumetric flask and make up to the mark with diluent. (Metformin $1000\mu g/ml$)

• Preparation of Working Standard of Lobeglitazone and Metformin:

From the above prepared solutions; take 1ml of Lobeglitazone stock solution and 1ml of Metformin stock solution in 10ml of volumetric flask and make up to the mark with diluent. (Lobeglitazone $0.1\mu g/ml$ and Metformin $100\mu g/ml$).

• Preparation of Sample for Recovery:

Lobeglitazone and Metformin $(0.1\mu g/ml$ and $100\mu g/ml$ respectively) drug solution was taken in three different flask labeled A, B and C. Spiked 50%, 100%, 150% of working standard solution in it and diluted up to 10ml. The area of each solution peak was measured. The amount of Lobeglitazone and Metformin was calculated at each level and % recoveries were calculated.

5. Limit of detection (LOD) and limit of quantitation (LOQ) Sensitivity of the

Nanotechnology Perceptions 20 No. S13 (2024)

proposed method was estimated in terms of LOD and LOQ. LOD is the lowest concentration in a sample that can be detected, but not necessarily quantified;

The LOD was estimated from the set of 3calibration curves used to determination linearity. The LOD may be calculated as,

$LOD = 3.3 \times (SD/Slope)$

Where, SD= Standard deviation of Y-intercepts of 3 calibration curves. Slope = Mean slope of the 3 calibration curves.

• The LOQ was estimated from the set of 3 calibration curves used to determine linearity.

The LOQ may be calculated as,

$LOQ = 10 \times (SD/Slope)$

Where, SD = Standard deviation of Y-intercepts of 3 calibration curves.

6. Robustness:

Robustness of the method was studied by making small deliberate changes in few parameters.

- Lobeglitazone and Metformin (0.1 and $100\mu g/ml$ respectively) drug solution was taken andinjected by applying little deliberate changes of the following method conditions and evaluated by RSD.
- Column Temperature: ±1 °C
- Flow rate: ±0.1 ml/min
- Mobile phase pH: ±0.1

Acceptance criteria:

- Number of theoretical plates for the analytepeak should not be less than 2000.
- Asymmetry value for the analyte peak shouldnot be more than 2.0.
- RSD for the analyte peak should not be morethan 2.0%.

7. Application of Method on MarketedProduct:

(Label claim: Lobeglitazone –0.5mg;Metformin - 500mg)

Twenty tablets were weighed; average weight was calculated and tablets were powdered finely. Tablet Powder equivalent to 0.5mg of Lobeglitazone and 500mg of Metformin were added into 100ml of volumetric flask Lobeglitazone (5µg/ml) and Metformin (5000µg/ml). Volume was made up to the mark with Methanol. The solution is then sonicated for 20mins and further the solution is filtrated. 1ml of each above solution of Lobeglitazone and Metformin was transferred to 10ml volumetric flask. Volume was made up to the mark with diluent, which gives Lobeglitazone (0.1µg/ml) and Metformin (100µg/ml). The quantification was carried out by keeping these values to be straight line equation of calibration curve

8. System suitability test

System suitability testing is essential for the assurance of the quality performance of

chromatographic system. Earlier prepared solutions for chromatographic conditions were tested for system suitability testing.

FORCED DEGRADATION STUDIES

Degradation conditions

- Hydrolysis- (a) Acid Hydrolysis
 (b) Base Hydrolysis
- 2. Oxidative
- 3. Photolytic
- 4. Thermal

Preparation of Reagent:

- **0.1 N HCl Solution:** 0.85ml conc. Hydrochloric acid was taken in 100ml volumetric flask and volume was made up to the mark with water and mixed well.
- **0.1 N NaOH Solution:** 0.4gm of NaOH pelletswere taken in 100ml volumetric flask and volume was made up to the mark with water and mixed well.
- **3% H2O2 Solution:** 3ml of the 30% H2O2 solution was taken in 100ml volumetric flask and volume was made up to the mark withwater and mixed well.

Acid Degradation:

- Transferring 1ml of stock solution of Lobeglitazone and Metformin into 10ml of volumetric flask.
- Add 2ml of 0.1N HCl solution
- Mixed well and kept for 1 hour at RT (25°C).
- The solution was neutralized with 2ml of 0.1N NaOH solution.'
- Then the volume was adjusted with the diluent to get sample solution concentration (Lobeglitazone 0.1µg/ml and Metformin 100µg/ml).

Base Degradation:

- Transferring 1ml of stock solution of Lobeglitazone and Metformin into 10ml of volumetric flask.
- Add 2ml of 0.1N NaOH solution
- Mixed well and kept for 1 hour at RT (25°C).
- The solution was neutralized with 2ml of 0.1N HCL solution.
- Then the volume was adjusted with the diluent to get sample solution concentration (Lobeglitazone 0.1µg/ml and Metformin 100µg/ml).

Oxidative Degradation:

- Transferring 1ml of stock solution of Lobeglitazone and Metformin into 10ml of volumetric flask.
- Add 2ml of 3% H2O2 solution
- Mixed well and kept for 1 hour at RT (25°C).
- Then the volume was adjusted with the diluent to get sample solution concentration

Nanotechnology Perceptions 20 No. S13 (2024)

(Lobeglitazone 0.1µg/ml and Metformin 100µg/ml).

Photo Degradation:

- Transferring 1ml of stock solution of Lobeglitazone and Metformin into petri dish.
- Then it was kept in UV chamber for 3 Days under 1.2 million lux h for visible light.
- Then the volume was adjusted and then diluted with the diluent to get working solution concentration (Lobeglitazone 0.1µg/ml and Metformin 100µg/ml).

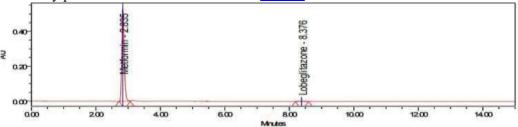
Thermal Degradation:

Lobeglitazone (25 mg) and Metformin (50mg) were taken in 50ml of volumetric flask and was kept in oven for 2 hours at 105°C temperature.

- Then after volumetric flask was removed and cooled down to room temp.
- Mobile phase was added to dissolve the drug and volume was made up with the diluent up tomark.
- 1ml of this solution was transferred in 10ml volumetric flask.
- Volume was made up with mobile phase to get working solution concentration (Lobeglitazone 0.1 µg/ml and Metformin 100 µg/ml).

RESULTS

To develop an accurate, precise and specific stability indicating RP-HPLC method for estimation of Lobeglitazone and Metformin using stressed samples, various mobile phases with different composition and flow rate were tried. After several compositions and permutations, chromatographic conditions were optimized and established. Satisfactory estimation of MUP with good peak symmetry and steady baseline was obtained with the mobile phase Methanol: ACN: Potassium Dihydrogen Phosphate buffer pH 2.5 (70:5:25 %v/v/v) at a flow rate of 0.8 mL/min.These conditions showed sharp peak ofLobeglitazone and Metformin with retention time 8.37 min and 2.83 min respectively and all the system suitability parameters meet with the criteria table-1.



	Peak Results									
	Name	RT	Aroa	Height	USP Tailing	USP Plate Count	Symmetry Factor	Purity1 Threshold	Purity1 Angle	Resolution
1	Metformin	2.835	2841062	536083	1.33	6679	1.33	0.637	0.219	
2	Lobegitasone	8.376	16784	1370	1.08	10154	1.06	3.014	2.397	23.33

Table 1 System Suitability Parameters METHOD VALIDATION SUMMARY

Parameters		Lobeglitazone	Metformin			
Specificity		Specific				
Linearity		0.05-0.25 μg/ml	50 – 250 μg/ml			
Intraday		1.35	0.73			
		0.56-1.29	0.51-0.94			
		0.51-1.29	0.53-0.92			
Accuracy	50%	100-101.06	98.37-99.55			
	100%	98.44-100.25	98.93-99.18			
	150%	100.46-101.65	99.80-101.03			
Robustness		The system suitability parameters were found well within the Acceptance criteria as per system suitability.				
Limit of Detecti	ion	0.0010μg/ml	0.3019μg/ml			
Limit of Quanti	tation	0.0031µg/ml	0.9151μg/ml			
% Assay		99.41 %	100.40 %			

Degradation Studies

The chromatograms obtained from samples exposed to acidic, alkaline, oxidative and photo degradation depicted well-separated peaks of pure Lobeglitazone and Metformin having tR 8.21 min and 2.89 min respectively also some additional peaks at different values. The % of degradation products with their tR values is listed in Table 12 and Figure 5, 6,7,8,9.

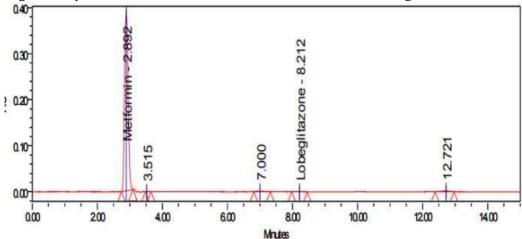


Fig.5. Chromatogram of Standard Lobeglitazone (0.1 μ g/ml) and Metformin (100 μ g/ml) for AcidDegradation

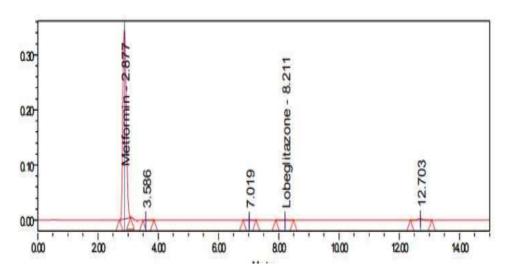


Fig. 6. Chromatogram of Standard Lobeglitazone (0.1 μ g/ml) and Metformin (100 μ g/ml) for BaseDegradation

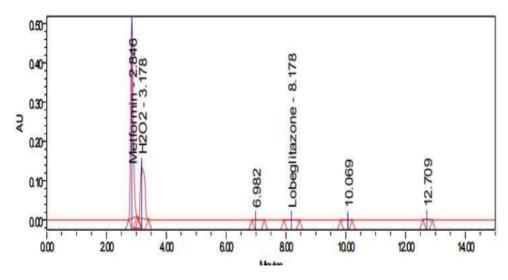


Fig. 7. Chromatogram of Standard Lobeglitazone (0.1 μ g/ml) and Metformin (100 μ g/ml) for OxidativeDegradation

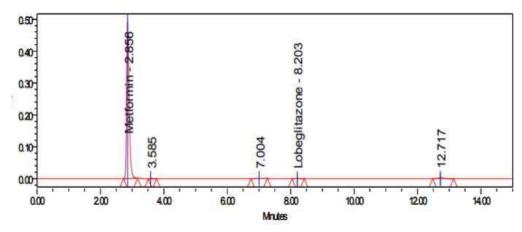


Fig. 8. Chromatogram of Standard Lobeglitazone (0.1 μ g/ml) and Metformin (100 μ g/ml) for PhotoDegradation

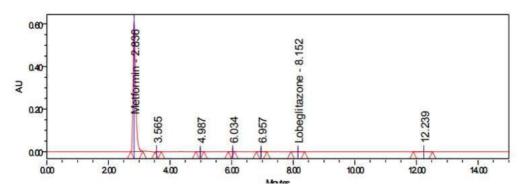


Fig. 9. Chromatogram of standard Lobeglitazone (0.1 μ g/ml) and Metformin (100 μ g/ml) for ThermalDegradation

SUMMARY OF FORCED DEGRADATION STUDIES

Sr.	Types	Condition	Duratio	Solution		%Degradati
No.	of Degradation		n			on
1	Acid Degradation	0.1 N HCL	1 Hour	Lobeglitazo ne	5882	18.03
				Metformin	301016 6	22.51
2	Base Degradation	0.1 N NaOH	1 Hour	Lobeglitazo ne	5871	17.19
				Metformin	274745	29.97

					7	
3	Oxidative Degradation	3% H ₂ O ₂	1 Hour	Lobeglitazo ne	5191	26.78
				Metformin	299579 4	22.88
4	Photo Degradation	-	18 Hours	Lobeglitazo ne	6117	13.72
				Metformin	276753 1	28.75
5	Thermal Degradation	-	2 Hour	Lobeglitazo ne	5800	18.19
				Metformin	342193 7	11.91

SUMMARY:

The combination of Lobeglitazone and Metformin has been approved by CDSCO on 30 December 2022. Glenmark Pharmaceuticals has launchedtablet formulation with combination of two drugs Lobeglitazone and Metformin under the brand name "LOBG-M" for treatment of Type-2 Diabetes Mellitus. Lobeglitazone is not official in any Pharmacopoeia and Metformin is official in Indian, United States, British and European Pharmacopoeia. An approach of forced degradationstudy was successfully applied for the development of stability indicating assay method for simultaneous estimation of Lobeglitazone and Metformin combined dosage form in presence of its degradation products. The method has shown adequateseparation of main peaks from their associated degradation products. Separation was achieved on Cosmosil C18 RP column, 250 mm \times 4.6 mm, 5 μ m, using a mobile phase Methanol: ACN: Phosphate Buffer pH2.5 (70:5:25 v/v/v), Adjust pH

In the present study, comprehensive stress testing of both drug in combined dosage form was carried out according to ICH guideline Q1A (R2). The specificity of the method was determined by assessing interference from blank and by Forced Degradation.

2.5 using 1% OPA at a flow rate of 0.8 ml/min and UV detection was carried out at 250 nm.

Specificity of the method was establishedby determining that peaks are separated well so there is no co-elution of any degradation products with main peaks and the results obtained werefound within the acceptance criteria. Hence, themethod can be termed as specific. For the linearity, correlation coefficient value should not be less than 0.995 for given range. Correlation coefficient value was found to be 0.999 and 0.999 for Lobeglitazone and Metformin respectively, which is greater than 0.995. Hence, the method is linear within the range. Accuracy was determined over the range from lowest sample concentration to highest concentration (i.e. at 50%, 100% and 150%). According to acceptance criteria individual % recovery should be in the range of 98-102%. The results show that the % recoveries for Lobeglitazone and Metformin were found to be 99.54-101.25 % and 100.82-101.096% respectively which is well within the acceptance criteria. Hence, the method can be termed as accurate.

In order to show the precision of the method, repeatability and intermediate precision were carried out. For the repeatability, RSD of the assay of six sample preparations should not be more than 2%. The obtained RSD was found to be

1.35 And 0.73 for Lobeglitazone and Metforminrespectively which are well within the limit of acceptance criteria. While for the intermediate precision of the method, the same procedure was followed on a same day at specific interval and on different day. RSD for intraday precision were found to be in the range of 0.56-1.29 and 0.51-0.94 for Lobeglitazone and Metformin respectively. RSD for Interday precision were found to be in the range of 0.51-1.29 and 0.53-0.92 for Lobeglitazone and Metformin respectively which also well within the limit of acceptance criteria and absolute difference between mean assay value of method precision and intermediate precision was found to be less than 2.0 % which is within the limit of acceptance criteria. Hence, the method can be termed as precise.

The LOD for Lobeglitazone and Metformin was found to be $0.076\mu g/ml$ and $0.021\mu g/ml$ respectively. Similarly, LOQ for Lobeglitazone and Metformin was found to be $0.20\mu g/ml$ and $0.63\mu g/ml$ respectively. The % assay results of 99.41 % for Lobeglitazone and 100.40% for Metformin indicate that the developedmethod was successfully utilized for the estimation of Lobeglitazone and Metformin in their Tablet Formulation.

The Robustness study is used to demonstrate the method's efficiency in the face of purposeful changes in conventional method factors, such as flow rate, pH, and so on. The assay obtained following the changes suggested was compared to the assay obtained under normal conditions. The test difference should not be greater than 2%, according to the approval requirements. The gained outcomes are well within the acceptable ranges. As a result, the approachmay be described as robust.

As its results for all validation parameters are well within the limit of acceptance criteria, the technique may be regarded validated as suitable for intended purpose.

So, during stability studies on Lobeglitazone and Metformin, the suggested stability indicating RP-HPLC method was effectively employed for the simultaneous assessment of both drugs in combination dosage form in the presence of degradation products

CONCLUSION

From above observations, it can be concluded that developed Stability Indicating Method and validation of Lobeglitazone and Metformin in tablets by RP-HPLC is, Specific, Linear, Accurate, Precise and Robust. Thus, above developed RP-HPLC method can be applied for routine analysis.

REFERENCE

- 1. Kazakevich, Yuri; LoBrutto, Rosario, eds. (2007). HPLC for pharmaceutical scientists. Hoboken, NJ: Wiley-Interscience. ISBN 978-0-471-68162-5.
- 2. ^ Levin, Shulamit (January 2004). "Reversed Phase Stationary Phases in Pharmaceutical Sciences". Journal of Liquid Chromatography & Related Technologies. **27** (7–9): 1353–1376. doi:10.1081/JLC-120030606. ISSN 1082-6076. S2CID 97490509.
- 3. ^ Gerber, F.; Krummen, M.; Potgeter, H.; Roth, A.; Siffrin, C.; Spoendlin, C. (2004). "Practical aspects of fast reversed-phase high-performance liquid chromatography using 3µm particle packed

- columns and monolithic columns in pharmaceutical development and production working under current good manufacturing practice". Journal of Chromatography A. **1036** (2): 127–133. doi:10.1016/j.chroma.2004.02.056. PMID 15146913.
- 4. ^ Bayne, Shirley; Carlin, Michelle (2017). Forensic Applications of High Performance Liquid Chromatography (1st ed.). CRC Press. ISBN 9780429251962.
- 5. ^ Seger, Christoph; Salzmann, Linda (2020-08-01). "After another decade: LC–MS/MS became routine in clinical diagnostics". Clinical Biochemistry. Advancement and Applications of Mass Spectrometry in Laboratory Medicine. **82**: 2–11. doi:10.1016/j.clinbiochem.2020.03.004. ISSN 0009-
 - 9120. PMID 32188572. S2CID 213186669.
- 6. ^ Dong, Michael (2018). "Ten Common-Sense Corollaries in Pharmaceutical Analysis by High Performance Liquid Chromatography". LCGC Europe. LCGC Europe-08-01-2018. **31** (8): 432–436.
- 7. ^ Snyder, Lloyd R.; Kirkland, Joseph J.; Glajch, Joseph L. (2012). Practical HPLC Method Development (2nd ed.). John Wiley & Sons.
- 8. ^ McMaster, Marvin C. (2007). HPLC: a practical user's guide (2nd ed.). Hoboken, NJ: Wiley-Interscience. ISBN 978-0-471-75401-5.
- 9. ^ Hanai, Toshihiko; Hanai, T. (1999). HPLC: a practical guide. RSC chromatography monographs. Royal Society of Chemistry. Cambridge: Royal Society of Chemistry. ISBN 978-0-85404-515-0.
- 10. ^ Jump up to: ^{a b} Karger, Barry L. (1997). "HPLC: Early and Recent Perspectives". Journal of Chemical Education. **74** (1): 45. Bibcode:1997JChEd..74...45K. doi:10.1021/ed074p45.
- 11. ^ Jump up to: a b c d e f Henry, Richard A. (1 February 2009) "The Early Days of HPLC at Dupont". Chromatography Online. Avanstar Communications Inc.
- 12. ^ Giddings, Calvin (1965). Dynamics of Chromatography: Principles and Theory. Marcel Dekker.
- 13. ^ Jump up to: ^{a b} Levin, Shulamit (2017). Grinberg, Nelu; Carr, Peter W. (eds.). Solid-Core or Fully Porous Columns in Ultra High-Performance Liquid Chromatography—Which Way to Go for Better Efficiency of the Separation?. Advances in Chromatography. Vol. 55 (1 ed.). Boca Raton: CRC Press. pp. 185–203. ISBN 9781315158075.
- 14. ^ Iler, R.K. (1979) The Chemistry of Silica. John Wiley & Sons. New York.
- 15. ^ Karger, B. L.; Berry, L. V. (1971). "Rapid liquid-chromatographic separation of steroids on columns heavily loaded with stationary phase". Clin. Chem. **17** (8): 757–64. doi:10.1093/clinchem/17.8.757. PMID 4254537.
- 16. ^ Neue, Uwe D. (1997). HPLC columns: theory, technology, and practice. New York, NY: Wiley VCH. ISBN 978-0-471-19037-0.
- 17. ^ Giddings, J. Calvin (1965) Dynamics of Chromatography, Part I. Principles and Theory. Marcel Dekker, Inc., New York. p. 281.
- 18. ^ Ettre, C. (2001). "Milestones in Chromatography: The Birth of Partition Chromatography" (PDF). LCGC. **19** (5): 506–512. Retrieved 2016-02-26.
- 19. ^ Martin, A J P; Synge, R L M (1941). "Separation of the higher monoamino-acids by counter-current liquid-liquid extraction: the amino-acid composition of wool". Biochemical Journal. **35** (1–2): 91–121. doi:10.1042/bj0350091. PMC 1265473. PMID 16747393.
- Lindsay, S.; Kealey, D. (1987). High performance liquid chromatography. Wiley. OSTI 7013902. from review Hung, L. B.; Parcher, J. F.; Shores, J. C.; Ward, E. H. (1988). "Theoretical and experimental foundation for surface-coverage programming in gas-solid chromatography with an adsorbable carrier gas". J. Am. Chem. Soc. 110 (11): 1090–1096. doi:10.1021/ac00162a003.
- 21. ^ Displacement Chromatography. Sacheminc.com. Retrieved 2011-06-07. Archived September 15, 2008, at the Wayback Machine

- 22. ^ LoBrutto, Rosario; Kazakevich, Yuri (2007-01-22), Kazakevich, Yuri; LoBrutto, Rosario (eds.), "Reversed-Phase HPLC", HPLC for Pharmaceutical Scientists (1 ed.), Wiley, pp. 139–239, doi:10.1002/9780470087954.ch4, ISBN 978-0-471-68162-5, retrieved 2023-10-10
- 23. ^ Kazakevich, Yuri; LoBrutto, Rosario (2007-01-22), Kazakevich, Yuri; LoBrutto, Rosario (eds.), "Size-Exclusion Chromatography", HPLC for Pharmaceutical Scientists (1 ed.), Wiley, pp. 263–279, doi:10.1002/9780470087954.ch6, ISBN 978-0-471-68162-5, retrieved 2023-10-10
- 24. ^ Mulloy, Barbara; Heath, Alan; Shriver, Zachary; Jameison, Fabian; Al Hakim, Ali; Morris, Tina S.; Szajek, Anita Y. (2014-08-01). "USP compendial methods for analysis of heparin: chromatographic determination of molecular weight distributions for heparin sodium". Analytical and Bioanalytical Chemistry. 406 (20): 4815–4823. doi:10.1007/s00216-014-7940-3. hdl:1721.1/104914. ISSN 1618-2650. PMID 24958344. S2CID 492085.
- 25. ^ Fritz, James S.; Gjerde, Douglas T. (2000-04-25). Ion Chromatography (1 ed.). Wiley. doi:10.1002/9783527613243. ISBN 978-3-527-29914-0.
- A Zhang, Chenhua; Rodriguez, Elliott; Bi, Cong; Zheng, Xiwei; Suresh, Doddavenkatana; Suh, Kyungah; Li, Zhao; Elsebaei, Fawzi; Hage, David S. (2018). "High performance affinity chromatography and related separation methods for the analysis of biological and pharmaceutical agents". Analyst. 143 (2): 374—391. Bibcode:2018Ana...143..374Z. doi:10.1039/C7AN01469D. ISSN 1364-5528. PMC 5768458. PMID 29200216.
- 27. ^ Jump up to:^{a b} McCalley, David V. (2017-11-10). "Understanding and manipulating the separation in hydrophilic interaction liquid chromatography". Journal of Chromatography A. **1523**: 49–71. doi:10.1016/j.chroma.2017.06.026. ISSN 1873-3778. PMID 28668366.
- 28. ^ Jump up to:^{a b} Buszewski, Bogusław; Noga, Sylwia (2012). "Hydrophilic interaction liquid chromatography (HILIC)—a powerful separation technique". Analytical and Bioanalytical Chemistry. **402** (1): 231–247. doi:10.1007/s00216-011-5308-5. ISSN 1618-2650. PMC 3249561. PMID 21879300.
- 29. ^ Schellinger, Adam P.; Carr, Peter W. (2006). "Isocratic and gradient elution chromatography: A comparison in terms of speed, retention reproducibility and quantitation". Journal of Chromatography A. **1109** (2): 253–266. doi:10.1016/j.chroma.2006.01.047. PMID 16460742. S2CID 26072994.
- 30. ^ Ettre, L. S.; Zlatkis, A., eds. (1979), "Csaba Horváth", Journal of Chromatography Library, 75 years of Chromatography a Historical Dialogue, vol. 17, Elsevier, pp. 151–158, doi:10.1016/s0301-4770(08)60645-4, ISBN 9780444417541, retrieved 2023-10-15
- 31. ^ Snyder, Lloyd R.; Dolan, John W. (2006). High-Performance Gradient Elution: The Practical Application of the Linear-Solvent-Strength Model. Wiley Interscience. ISBN 978-0470055519.
- 32. ^ Schellinger, Adam P.; Carr, Peter W. (2006). "Isocratic and gradient elution chromatography: A comparison in terms of speed, retention reproducibility and quantitation". Journal of Chromatography A. 19th International Symposium on MicroScale Bioseparations. **1109** (2): 253–266. doi:10.1016/j.chroma.2006.01.047. ISSN 0021-9673. PMID 16460742. S2CID 26072994.
- 33. ^ Dolan, John W. (2014). "LC Method Scaling, Part II: Gradient Separations". LCGC North America. 32 (3): 188–193.
- 34. ^ Martin, A. J. P.; Synge, R. L. M. (1941-12-01). "A new form of chromatogram employing two liquid phases". Biochemical Journal. **35** (12): 1358–1368. doi:10.1042/bj0351358. ISSN 0306-3283. PMC 1265645. PMID 16747422.
- 35. ^ https://www.usp.org/sites/default/files/usp/document/harmonization/gen-chapter/harmonization-november-2021-m99380.pdf [bare URL PDF], CHROMATOGRAPH, Stage 4 Harmonization (December 1, 2022)

- 36. *Wren, Stephen A. C. (2005-06-15). "Peak capacity in gradient ultra performance liquid chromatography (UPLC)". Journal of Pharmaceutical and Biomedical Analysis. **38** (2): 337–343. doi:10.1016/j.jpba.2004.12.028. ISSN 0731-7085. PMID 15925228.
- 37. ^ Zelenyánszki, Dóra; Felinger, Attila (2020-10-01). "The Impact of Column Hardware on Efficiency in Liquid Chromatography (LC)". LCGC Europe. LCGC Europe-10-01-2020. **33** (10): 498–504.
- 38. ^ Dolan, John (2014). "LC Method Scaling, Part II: Gradient Separations". LCGC North America. LCGC North America-03-01-2014. **32** (3): 188–193.
- 39. ^ Jensen, Ole Elvang; Kidal, Steffen (2006-03-01). "Using Volumetric Flow to Scaleup Chromatographic Processes". BioPharm International. BioPharm International-03-01-2006. **19** (3).
- 40. ^ Jump up to: ^{a b} Walter, Thomas H.; Andrews, Richard W. (2014). "Recent innovations in UHPLC columns and instrumentation". Trends in Analytical Chemistry. **63**: 14–20. doi:10.1016/j.trac.2014.07.016. ISSN 0165-9936.
- 41. ^ Majors, Ronald E.. (2010-09-07) Fast and Ultrafast HPLC on sub-2 μm Porous Particles Where Do We Go From Here? LC-GC Europe. Lcgceurope.com. Retrieved 2011-06-07.
- 42. ^ Xiang, Y.; Liu Y.; Lee M.L. (2006). "Ultrahigh pressure liquid chromatography using elevated temperature". Journal of Chromatography A. **1104** (1–2): 198–202. doi:10.1016/j.chroma.2005.11.118. PMID 16376355.
- 43. ^ Horváth, Cs.; Preiss B.A.; Lipsky S.R. (1967). "Fast liquid chromatography. Investigation of operating parameters and the separation of nucleotides on pellicular ion exchangers". Analytical Chemistry. **39** (12): 1422–1428. doi:10.1021/ac60256a003. PMID 6073805.
- 44. ^ Nguyen, Dao T.-T.; Guillarme, Davy; Rudaz, Serge; Veuthey, Jean-Luc (2006). "Fast analysis in liquid chromatography using small particle size and high pressure". Journal of Separation Science. **29** (12): 1836–1848. doi:10.1002/jssc.200600189. ISSN 1615-9306. PMID 16970187.
- 45. ^ Gritti, Fabrice; Guiochon, Georges (2013). "The van Deemter equation: Assumptions, limits, and adjustment to modern high performance liquid chromatography". Journal of Chromatography A. **1302**: 1–13. doi:10.1016/j.chroma.2013.06.032. PMID 23838304.
- 46. ^ Xiang, Yanqiao; Liu, Yansheng; Lee, Milton L. (2006). "Ultrahigh pressure liquid chromatography using elevated temperature". Journal of Chromatography A. **1104** (1–2): 198–202. doi:10.1016/j.chroma.2005.11.118. PMID 16376355.
- 47. ^ 1290 Infinity Quaternary Pump Archived 2015-11-20 at the Wayback Machine. Agilent
- 48. ^ waters. "Trademarks: Waters". www.waters.com.
- K., Robards (1994). Principles and practice of modern chromatographic methods. Haddad, P. R., Jackson, P. E. Amsterdam: Elsevier/Academic Press. ISBN 9780080571782. OCLC 815471219.
- 50. ^ "HPLC-ECD (Electrochemical Detection) Fundamentals".
- 51. ^ Markovitch, Omer; Ottelé, Jim; Veldman, Obe; Otto, Sijbren (2020). "Automated device for continuous stirring while sampling in liquid chromatography systems". Communications Chemistry. **3** (1): 180. doi:10.1038/s42004-020-00427-5. PMC 9814086. PMID 36703458.
- 52. ^ Gerber, Frederic (May 2004). "Practical aspects of fast reversed-phase high-performance liquid chromatography using 3 μm particle packed columns and monolithic columns in pharmaceutical development and production working under current good manufacturing practice". Journal of Chromatography. **1036** (2): 127–33. doi:10.1016/j.chroma.2004.02.056. PMID 15146913.
- 53. ^ Siddiqui, Masoom Raza; AlOthman, Zeid A.; Rahman, Nafisur (2013). "Analytical techniques in pharmaceutical analysis: A review". Arabian Journal of Chemistry. **10**: S1409—S1421. doi:10.1016/j.arabjc.2013.04.016.
- 54. ^ The European Pharmacopoeia, 2002. fourth ed., Council of Europe, Strasbourg.
- 55. ^ United States Pharmacopoeia, 2004. 27th ed. The USP Convention Inc., Rockville, MD.

- 56. ^ Merone, Giuseppe M.; Tartaglia, Angela; Rossi, Sandra; Santavenere, Francesco; Bassotti, Elisa; D'Ovidio, Cristian; Bonelli, Martina; Rosato, Enrica; de Grazia, Ugo; Locatelli, Marcello; Savini, Fabio (2021). "Fast Quantitative LC-MS/MS Determination of Illicit Substances in Solid and Liquid Unknown Seized Samples". Analytical Chemistry. 93 (49): 16308–16313. doi:10.1021/acs.analchem.1c03310. ISSN 0003-2700. PMC 8674870. PMID 34843645.
- 57. ^ Pesce, Amadeo; Rosenthal, Murray; West, Robert; West, Cameron; Crews, Bridgit; Mikel, Charles; Almazan, Perla; Latyshev, Sergey (2010-06-01). "An evaluation of the diagnostic accuracy of liquid chromatography-tandem mass spectrometry versus immunoassay drug testing in pain patients". Pain Physician. 13 (3): 273–281. PMID 20495592.
- 58. ^ Tsai, I.-Lin; Weng, Te-I.; Tseng, Yufeng J.; Tan, Happy Kuy-Lok; Sun, Hsiao-Ju; Kuo, Ching-Hua (2013-12-01). "Screening and confirmation of 62 drugs of abuse and metabolites in urine by ultra-high-performance liquid chromatography-quadrupole time-of-flight mass spectrometry". Journal of Analytical Toxicology. 37 (9): 642–651. doi:10.1093/jat/bkt083. PMID 24084874.
- 59. ^ Weinmann, W.; Renz, M.; Vogt, S.; Pollak, S. (2000-01-01). "Automated solid-phase extraction and two-step derivatisation for simultaneous analysis of basic illicit drugs in serum by GC/MS". International Journal of Legal Medicine. **113** (4): 229–235. doi:10.1007/s004149900098. PMID 10929239. S2CID 20451772.
- 60. ^ Kolmonen, Marjo; Leinonen, Antti; Pelander, Anna; Ojanperä, Ilkka (2007-02-28). "A general screening method for doping agents in human urine by solid phase extraction and liquid chromatography/time-of-flight mass spectrometry". Analytica Chimica Acta. **585** (1): 94–102. Bibcode:2007AcAC..585...94K. doi:10.1016/j.aca.2006.12.028. PMID 17386652.
- 61. ^ Pelander, Anna; Ojanperä, Ilkka; Laks, Suvi; Rasanen, Ilpo; Vuori, Erkki (2003-11-01). "Toxicological screening with formula-based metabolite identification by liquid chromatography/time-of-flight mass spectrometry". Analytical Chemistry. **75** (21): 5710–5718. doi:10.1021/ac0301620. PMID 14588010.
- 62. ^ Nobilis, Milan; Pour, Milan; Senel, Petr; Pavlík, Jan; Kunes, Jirí; Voprsalová, Marie; Kolárová, Lenka; Holcapek, Michal (2007-06-15). "Metabolic profiling of a potential antifungal drug, 3-(4-bromophenyl)-5-acetoxymethyl-2,5-dihydrofuran-2-one, in mouse urine using high-performance liquid chromatography with UV photodiode-array and mass spectrometric detection". Journal of Chromatography B. **853** (1–2): 10–19. doi:10.1016/j.jchromb.2007.02.045. PMID 17400036.
- 63. ^ Gu, Jatin; Patel, Kumar; Shah, Dhiren (2016). "APPLICATION OF LC-MS". PharmaTutor.
- 64. ^ Tallam, Anil Kumar; Alapati, Sahithi; Nuli, Mohana Vamsi (2023). "A review on bioanalytical method development and validation of anticancer drugs by using lc/ms/ms and its applications on routine analysis". Journal of Integral Sciences: 4–19. doi:10.37022/jis.v6i1.51. ISSN 2581-5679. S2CID 257295079.
- 65. ^ O'Driscoll, Aimee (2021). "HPLC in Pharmaceutical Applications". Lab Manager.
- A Beccaria, Marco; Cabooter, Deirdre (2020). "Current developments in LC-MS for pharmaceutical analysis". Analyst. 145 (4): 1129–1157. Bibcode:2020Ana...145.1129B. doi:10.1039/C9AN02145K. hdl:11392/2479221. ISSN 1364-5528. PMID 31971527. S2CID 210866236.
- 67. ^ Gu, Chunang (Christine); Russell, David; Yehl, Peter (2016). "Application of LCMS in small-molecule drug development". European Pharmaceutical Review. **21** (4): 54–57.
- 68. ^ D'Ovidio, Cristian; Locatelli, Marcello; Perrucci, Miryam; Ciriolo, Luigi; Furton, Kenneth G.; Gazioglu, Isil; Kabir, Abuzar; Merone, Giuseppe Maria; de Grazia, Ugo; Ali, Imran; Catena, Antonio Maria; Treglia, Michele; Marsella, Luigi T.; Savini, Fabio (2023). "LC-MS/MS Application in Pharmacotoxicological Field: Current State and New

- Applications". Molecules. **28** (5): 2127. doi:10.3390/molecules28052127. ISSN 1420-3049. PMC 10004468. PMID 36903374.
- 69. ^ Zhou, Juntuo; Zhong, Lijun (2022). "Applications of liquid chromatography-mass spectrometry based metabolomics in predictive and personalized medicine". Frontiers in Molecular Biosciences. 9. doi:10.3389/fmolb.2022.1049016. ISSN 2296-889X. PMC 9669074. PMID 36406271.
- 70. ^ Mathias, Patricia I.; Connor, Thomas H.; B'Hymer, Clayton (2017). "A review of high performance liquid chromatographic-mass spectrometric urinary methods for anticancer drug exposure of health care workers". Journal of Chromatography B. **1060**: 316–324. doi:10.1016/j.jchromb.2017.06.028. ISSN 1570-0232. PMC 5585056. PMID 28654869.
- 71. ^ Hernández, Félix; Sancho, Juan V.; Ibáñez, María; Guerrero, Carlos (2007). "Antibiotic residue determination in environmental waters by LC-MS". TrAC Trends in Analytical Chemistry. Pharmaceutical-residue analysis. **26** (6): 466–485. doi:10.1016/j.trac.2007.01.012. ISSN 0165-9936.
- Seger, Christoph; Salzmann, Linda (2020). "After another decade: LC–MS/MS became routine in clinical diagnostics". Clinical Biochemistry. Advancement and Applications of Mass Spectrometry in Laboratory Medicine. 82: 2–11. doi:10.1016/j.clinbiochem.2020.03.004. ISSN 0009-9120. PMID 32188572. S2CID 213186669.
- 73. ^ Sundström, Mira; Pelander, Anna; Angerer, Verena; Hutter, Melanie; Kneisel, Stefan; Ojanperä, Ilkka (2013-10-01). "A high-sensitivity ultra-high performance liquid chromatography/high-resolution time-of-flight mass spectrometry (UHPLC-HR-TOFMS) method for screening synthetic cannabinoids and other drugs of abuse in urine". Analytical and Bioanalytical Chemistry. 405 (26): 8463–8474. doi:10.1007/s00216-013-7272-8. PMID 23954996. S2CID 25743579.
- 74. ^ Gelb, Michael H.; Basheeruddin, Khaja; Burlina, Alberto; Chen, Hsiao-Jan; Chien, Yin-Hsiu; Dizikes, George; Dorley, Christine; Giugliani, Roberto; Hietala, Amy; Hong, Xinying; Kao, Shu-Min; Khaledi, Hamid; Klug, Tracy; Kubaski, Francyne; Liao, Hsuan-Chieh (2022). "Liquid Chromatography—Tandem Mass Spectrometry in Newborn Screening Laboratories". International Journal of Neonatal Screening. 8 (4): 62. doi:10.3390/ijns8040062. ISSN 2409-515X. PMC 9781967. PMID 36547379.
- 75. *Wang, Yanyun; Sun, Yun; Jiang, Tao (2019). "Clinical Application of LC–MS/MS in the Follow-Up for Treatment of Children with Methylmalonic Aciduria". Advances in Therapy. **36** (6): 1304–1313. doi:10.1007/s12325-019-00955-0. ISSN 1865-8652. PMID 31049874. S2CID 143432183.
- 76. ^ Arunkumar, Nivethitha; Langan, Thomas J.; Stapleton, Molly; Kubaski, Francyne; Mason, Robert W.; Singh, Rajendra; Kobayashi, Hironori; Yamaguchi, Seiji; Suzuki, Yasuyuki; Orii, Kenji; Orii, Tadao; Fukao, Toshiyuki; Tomatsu, Shunji (2020). "Newborn screening of mucopolysaccharidoses: past, present, and future". Journal of Human Genetics. 65 (7): 557–567. doi:10.1038/s10038-020-0744-8. ISSN 1435-232X. PMID 32277174. S2CID 92042115.
- 77. ^ Skogvold, Hanne Bendiksen; Rootwelt, Helge; Reubsaet, Léon; Elgstøen, Katja Benedikte Prestø; Wilson, Steven Ray (2023). "Dried blood spot analysis with liquid chromatography and mass spectrometry: Trends in clinical chemistry". Journal of Separation Science. **46** (15): e2300210. doi:10.1002/jssc.202300210. hdl:10852/105845. ISSN 1615-9306. PMID 37269205. S2CID 259047202.
- 78. ^ Zahedi Rad, Maliheh; Neyestani, Tirang Reza; Nikooyeh, Bahareh; Shariatzadeh, Nastaran; Kalayi, Ali; Khalaji, Niloufar; Gharavi, Azam (2015-01-01). "Competitive Protein-binding assay-based Enzyme-immunoassay Method, Compared to High-pressure Liquid Chromatography, Has a Very Lower Diagnostic Value to Detect Vitamin D Deficiency in 9–12 Years

Children". International Journal of Preventive Medicine. **6**: 67. doi:10.4103/2008-7802.161069. PMC 4542329. PMID 26330983.