

Unveiling Nature's Arsenal: Virtual Screening Of Phytoconstituents For Anti-Acne Potentials In Medicinal Plants

Shubhangi B. Sutar (Corresponding author)¹, Pranali R. Pangam², Sachin S. Mali³, Sachinkumar V. Patil⁴, Ravindra Ganpati Gaikwad⁵, Harshawardhan Patil⁶

¹Associate Professor, Department of Pharmaceutical Chemistry

Ashokrao Mane College of Pharmacy, Peth Vadgaon, Maharashtra, India

Email: shubhangi.sutar28@gmail.com. <https://orcid.org/0000-0003-1523-4873>

²Department of Pharmaceutical Quality Assurance Ashokrao Mane College of Pharmacy, Peth Vadgaon, Maharashtra, India Email: pranalipangam27@gmail.com

<https://orcid.org/0009-0001-0534-4414>

³Professor and Head, Department of Pharmaceutics (PG), Ashokrao Mane College of Pharmacy, Peth-Vadgaon, Maharashtra, India Email: sachinmali143@gmail.com

<https://orcid.org/0000-0002-8104-5854>

⁴Principal, Dr. Ashok Gujar Institute of Pharmacy, Maharashtra, India Email: sachinpatil.krd@gmail.com.

⁵Associate professor, Womens College of Pharmacy Peth-Vadgaon, Maharashtra, India Email: rvgaikwad92@gmail.com <https://orcid.org/0000-0002-7424-4150>

⁶Ph.D scholar, Dept. of Pharmaceutics, Government College of Pharmacy, Karad, Maharashtra, India

Email: harshawardhan87@gmail.com

<https://orcid.org/0009-0009-8222-7172>

This study utilizes in-silico screening techniques to identify potential anti-acne phytoconstituents from six medicinal plants. Through comprehensive virtual screening against acne-related protein targets, ten promising compounds were prioritized based on their favorable biological activity, drug likeliness, bioavailability, and non-toxic profiles. These findings highlight the potential of these phytoconstituents for developing safe and effective anti-acne formulations, although further validation through clinical studies is warranted. Overall, this in-silico screening approach offers valuable insights for efficiently identifying potential anti-acne compounds and accelerating the drug discovery process.

Keywords: In-silico screening, Phytoconstituents, Anti-acne activity, Medicinal plants, Drug discovery.

1 Introduction:

Acne is a common skin condition that affects millions of people worldwide, leading to significant physical and psychological distress [1], [2]. Despite various treatment options available, including topical creams, antibiotics, and oral medications, the increasing prevalence of antibiotic resistance and adverse effects has sparked interest in natural alternatives [3], [4]. Medicinal plants have long been recognized for their therapeutic potential due to the presence of diverse bioactive compounds [5] known as phytoconstituents [6], [7], [8].

In recent years, in-silico screening techniques have emerged as powerful tools in drug discovery, enabling the identification of potential lead compounds in a cost-effective and time-efficient manner [6], [9], [10]. Virtual screening allows researchers to computationally evaluate large datasets of phytoconstituents and predict their interactions with specific protein targets related to acne pathogenesis [7], [8].

In this context, we aimed to exploit the vast potential of in-silico screening to identify novel phytoconstituents from commonly used medicinal plants with promising anti-acne activity [1], [2], [11], [12], [13]. We focused on six medicinal plants, including orange peel, lemon peel, green tea, turmeric, neem, and pomegranate peel, known for their therapeutic properties against various skin ailments [14], [15]. TNF-alpha (Tumor Necrosis Factor-alpha) and GehA (Glycerol-ester hydrolase A) are two potential anti-acne targets that have been studied for their roles in the pathogenesis of acne and as potential points of intervention for acne treatment [16].

TNF-alpha as an anti-acne target:

TNF-alpha is a pro-inflammatory cytokine that plays a crucial role in the inflammatory response, including skin inflammation. In the context of acne, increased levels of TNF-alpha have been observed in acne lesions, indicating its involvement in the inflammatory process associated with acne development. Elevated levels of TNF-alpha contribute to the recruitment of immune cells, formation of inflammatory lesions, and exacerbation of acne symptoms [17]. Targeting TNF-alpha as an anti-acne strategy involves inhibiting its activity or blocking its receptor. Drugs that can neutralize or reduce TNF-alpha activity have been used in various inflammatory conditions and may have potential for treating acne by reducing inflammation and suppressing the inflammatory response in the skin [7], [12], [18].

GehA as an anti-acne target:

GehA, also known as lipase or glycerol-ester hydrolase A, is an enzyme produced by certain bacteria, including *Propionibacterium acnes*, which is a key bacterium involved in the development of acne. GehA is responsible for breaking down triglycerides in sebum into free fatty acids, contributing to the formation of comedones and inflammation in acne lesions [19]. Inhibiting the activity of GehA could be a promising approach for acne treatment, as it may reduce the release of pro-inflammatory free fatty acids and mitigate the inflammatory response associated with acne. By targeting GehA, the overproduction of free fatty acids can be controlled, potentially leading to a decrease in the severity of acne lesions.

Both TNF-alpha and GehA represent potential targets for the development of novel anti-acne treatments. Targeting these molecules could help modulate the inflammatory response and lipase activity, respectively, contributing to the management of acne symptoms.

In this study, we conducted a comprehensive analysis of 50 phytoconstituents [6], [7] sourced from the IMPPAT database, which is a curated repository of phytochemicals from Indian medicinal plants. Our primary objective was to identify potential lead compounds with favorable drug-like properties that could be utilized for anti-acne applications. To ensure that the selected phytoconstituents had a higher likelihood of being orally bioavailable and drug-like, we applied Lipinski's rule of five. This well-established rule helps filter out compounds with poor pharmacokinetic properties, such as high molecular weight, excessive hydrogen bond donors, hydrogen bond acceptors, and lipophilicity. By adhering to Lipinski's rule, we narrowed down the candidate pool to compounds with improved prospects for further development [7], [8].

Subsequently, we performed molecular docking studies to predict the binding affinity of the selected phytoconstituents with specific target proteins (GehA and TNF-alpha), involved in acne pathogenesis [6]. This process allowed us to identify compounds that potentially interacted favorably with the target proteins, suggesting their potential efficacy against acne. To further assess the safety and pharmacokinetic characteristics of the shortlisted compounds, we employed ADMET (Absorption, Distribution, Metabolism, Excretion, and Toxicity) property analysis. This step is crucial in evaluating the compounds' potential for absorption, distribution within the body, metabolism, excretion, and potential toxicity risks. By considering these ADME and toxicity parameters, we aimed to identify compounds that have a higher likelihood of successful development as anti-acne agents. Moreover, we also conducted PASS (Prediction of Activity Spectra for Substances) evaluation, which is a computational tool used to predict the probability of biological activity for the compounds [20], [21], [22]. This analysis enabled us to identify compounds with potential anti-acne activity based on their structural similarity to known active compounds in the PASS database. Overall, the combined use of Lipinski's rule of five, molecular docking, ADMET properties, and PASS evaluation allowed us to prioritize and identify promising phytoconstituents with optimal drug-like properties and potential biological activity against acne [6], [7], [23], [24], [25]. The compounds selected through this rigorous in-silico screening process hold great promise for further investigation and development as potential components in safe and effective anti-acne formulations [26]. However, it is important to note that further experimental validation, such as in vitro and in vivo studies, and potentially clinical trials, will be necessary to confirm their actual anti-acne efficacy and safety before practical application [10].

2 Methodology:

2.1 Selection of Phytoconstituents:

A total of 50 phytoconstituents were chosen from the IMPPAT database, a curated repository of phytochemicals derived from Indian medicinal plants, based on their reported anti-acne potential in the existing literature. These compounds were derived from six different medicinal plants, including orange peel, lemon peel, green tea, turmeric, neem, and pomegranate peel [27], [28], [29], [30], [31], [32], [33], [34].

2.2 Lipinski's Rule of Five Screening:

The selected 50 phytoconstituents were subjected to Lipinski's Rule of Five analysis to assess their drug-like properties. Parameters such as molecular weight (MW), lipophilicity (LogP), hydrogen bond donors (HBD), and hydrogen bond acceptors (HBA) were evaluated. Compounds that satisfied all of Lipinski's criteria ($MW \leq 500$, $LogP \leq 5$, $HBD \leq 5$, $HBA \leq 10$) were retained for further analysis, as they are more likely to have favorable pharmacokinetic properties [35], [36].

2.3 Protein Targets Selection:

Two potential protein targets, GehA (Glycerol-ester hydrolase A) and TNF-alpha (Tumor Necrosis Factor-alpha), were chosen based on their known involvement in acne pathogenesis. GehA is associated with the hydrolysis of lipids, while TNF-alpha is a pro-inflammatory cytokine [8], [37], [38], [39], [40], [41].

2.4 Homology Modeling (GehA):

The amino acid sequence of GehA was retrieved from the UniProtKB database (UniProt ID: Q6A601) for homology modeling. A suitable template with a known three-dimensional structure and high sequence similarity to GehA was identified from the Protein Data Bank (PDB) using the Swiss Model automated homology modeling server. The homology model of GehA was generated using the target sequence and the chosen template through alignment-based comparative modeling [8], [41].

2.5 Receptor Preparation (TNF-alpha):

The X-ray crystal structure of TNF-alpha (PDB ID: 2AZ5, resolution: 2.10Å) was downloaded from the RCSB-PDB database in PDB format. The protein structure was prepared by removing water molecules, heteroatoms, and co-crystallized ligands using BIOVIA Visualizer software, leaving only the TNF-alpha protein for further analysis [42], [43].

2.6 Molecular Docking-Based Virtual Screening:

The homology model of GehA and the prepared structure of TNF-alpha were used as receptors in molecular docking simulations. The library of the 50 selected phytoconstituents was prepared by converting their chemical structures into 3D molecular structures. Molecular docking simulations were performed using specialized docking software (PyRx) to predict the binding interactions between the phytoconstituents and the GehA and TNF-alpha proteins. Discovery Studio Visualizer. Docking scores, as well as binding modes and interactions, were analyzed to identify phytoconstituents with potential high affinity to the target proteins [7], [15], [26].

2.7 ADMET Property Analysis:

The phytoconstituents with high docking scores were subjected to ADMET property prediction and evaluation using two computational tools, pkCSM and SwissADME. The ADMET prediction aimed to assess the compounds' drug-like physicochemical and pharmacokinetic properties, such as solubility, absorption, metabolism, distribution, and potential toxicity. ADMET analysis assessed the compounds' pharmacokinetic properties, including solubility, permeability, metabolism, and potential toxicity, to ensure their suitability for further development as drug candidates [7], [8].

2.8 PASS Evaluation:

The selected phytoconstituents were evaluated using the PASS (Prediction of Activity Spectra for Substances) online tool. PASS predicts the probability of biological activity for the compounds based on their structural similarity to known active compounds in the PASS database. Compounds with high PASS prediction scores for anti-acne activity were prioritized as potential lead candidates[6].

2.9 Interaction Analysis

The interaction analysis aimed to investigate the binding interactions between the docked protein-ligand complexes. Using Discovery Studio Visualizer, the binding poses and all possible interactions were explored. Discovery Studio Visualizer was used to explore the type of interactions, residual participation, and atomic coordinates. Compounds with specific interactions towards critical residues of GehA and TNF α , including the active site and binding site, were chosen for further analysis. This analysis identified lead compounds with potential anti-acne activity and provided valuable insights for further investigation and formulation development [16], [44], [45], [46], [47].

2.10 Data Analysis and Lead Molecule Selection:

The data obtained from molecular docking, ADMET analysis, and PASS evaluation were collectively analyzed to identify the most promising phytoconstituents with high binding affinity, favorable drug-like properties, and potential anti-acne activity. Based on the comprehensive analysis, a set of lead molecules that exhibited the most desirable characteristics were selected as potential candidates for anti-acne drug development [48], [49], [50], [51], [52], [53], [54], [55].

3 Results and Discussion

3.1 Molecular Docking-Based Virtual Screening

A virtual screening process was conducted using molecular docking to identify phytoconstituents from the IMPPAT database that could strongly bind to GehA and TNF α , two target proteins. The output provided the affinities and docked poses for each compound. Based on their binding affinity towards GehA and TNF α , the compounds were filtered, and the top 10 hits out of 50 were selected. [Table 1] These chosen compounds demonstrated significant binding affinity with the binding pocket of GehA and TNF α , suggesting their potential as high-affinity partners. This outcome highlights the promising therapeutic potential of these selected phytoconstituents in the drug development process.

3.1.1 Table 1

The top 10 hits and their binding affinities

Sr .	Compo und ID	Phytochemical Name	Source	Tar get	Bindi ng	Interac ting	Dista nce	Interact ions
------	--------------	--------------------	--------	---------	----------	--------------	-----------	---------------

N o.					Affin ity	Residu e		
1.	179442 7	o- Caffeoylquinic acid	<i>Camelli a sinensis</i>	Geh A	-7.8	HIS A:168	3.00	Conventi onal H- bond
						THR A:103	3.15	
						GLY A:97	2.83	
						SER A:58	3.14	
						ARG A:62	2.90	
							3.73	
								C-H bond
						LEU A:331	5.00	Pi-alkyl
						VAL A:330	3.74	Pi- Sigma
				2AZ 5	-8	ARG B:82	3.00	Conventi onal H- bond
						ASN D:92	2.92, 1.94, 2.50	
						LEU D:93	5.10	
								Pi-alkyl
2.	65064	Epigallocatechin gallate	<i>Camelli a sinensis</i>	Geh A	-8.4	SER A:169	2.35	Conventi onal H- bond
						GLN A:237	3.09	
						THR A:96	3.07	
						GLY A:298	3.20	
						THR A:103	3.12	
						SER A:58	2.48, 1.99, 2.89	
						SER A:59	2.71	
						ALA A:102	2.51	
						ARG A:62	2.89	

3.	73330	strictinin	<i>Punica granatum</i>	Geh A	-9.5	HIS A:297	3.62	C-H bond
							4.76	Pi-Pi T-shaped
						THR A:269	3.79	Pi-Sigma
						VAL A:3330	4.26	Pi-alkyl
						2AZ 5	-9	
						GLN B:125	3.26	Conventional H-bond
						GLY C:121	2.45, 1.97	
						LEU C:120	2.06	
						SER C:60	2.99	
						TYR C:151	2.97	
						LEU B:55	5.34	Pi-alkyl
						TYR C:119	4.95	Pi-Pi stacked
						THR A:103	1.71	Conventional H-bond
						THR A:269	3.04	
						THR A:103	2.81	
						THR A:269	2.82	
						HIS A:297	3.05	
						HIS A:297	3.19	
						SER A:58	3.22	
						ARG A:62	2.97	
						VAL A:330	5.37	Pi-alkyl
						ALA A:102	5.10	

						ARG A:62	3.31	C-H bond
						GLY A:98	3.61	
						GLY A:298	3.11	
4.	101518 74	Valoneic_acid_b ilactone	<i>Punica granatu m</i>	Geh A	-10 5	LYS B90	2.79	Conventi onal H- bond
						ARG B:82	3.07	
						GLN B:125	2.64	
						ARG D:82	2.94	
						ASN B:92	3.18	
						ASN B:92	3.39	C-H bond
						HIS A:297	2.96	Conventi onal H- bond
						GLY A:298	3.12	
						ARG A:62	3.33	
						SER A:58	2.95	
						ASN A:99	3.27	
							3.05	
							3.27	
						THR A:103	2.16	
					-9.2 5	VAL A:330	5.33	Pi-alkyl
						LEU A:331	5.33	
						ARG A:62	5.33	
						ALA A:102	4.85	
						GLY C:121	2.90	

						TYR D:151	2.21	Conventi onal H- bond
						TYR C:151	3.04	
						TYR C:59	4.40 4.70 3.70	Pi-Pi stacked
						LEU C:120	4.67	Amide- Pi- Stacked
5.	9064	Catechin	<i>Camelli a sinensis & Punica granatu m</i>	Geh A	-7	ALA A:102	2.04	Conventi onal H- bond
						THR A:103	2.06	
						THR A:96	3.97	Pi- Sigma
				2AZ 5	-8.6	LEU D:93	2.42	Conventi onal H- bond
						LEU B:93	2.56	
6.	72276	Epicatechin	<i>Camelli a sinensis & Punica granatu m</i>	Geh A	-6.7	THR A:103	2.10	Conventi onal H- bond
						THR A:269	2.85	
						SER A:58	2.87	
						ALA A:102	5.10	Pi-alkyl
						VAL A:330	4.50	
						ARG A:62	5.42	
				2AZ 5	-8.4	GLN D:125	2.06	Conventi onal H- bond
						ARG D:82	2.95	
						ASN B:92	2.97	
						GLN B:125	2.18 2.38	

7.	12313376	Nimbolide	<i>Azadirachta indica</i>	GehA	-8.4	TYR A:234	3.12	Conventional H-bond
						TYR A:234	3.14	
						TYR A:234	3.11	
						THR A:103	2.98	
						TYR A:234	3.76	
						GLU A:132	3.68	Pi-donor H bond
						GLY C:121	3.27	Conventional H-bond
						TYR D:151	2.75	
						TYR C:59	3.71	Pi-Pi stacked
						GLY C:121	3.62	C-H bond
						ALA A:204	4.39	Pi-alkyl
						ALA A:204	4.64	
						LEU A:207	5.09	
8.	11119228	Nimbiol	<i>Azadirachta indica</i>	GehA	-7.3	PRO A:215	4.45	
						TYR A:234	4.59	Pi-Pi stacked
						TYR C:59	4.46	Pi-alkyl
						LEU C:57	3.71	
						GLY C:121	5.51	Amide-Pi stacked
						SER A:58	3.00	Conventional H-bond
						HIS A:297	3.30	
						GLY A:298	2.80	

						LEU A:31	5.15	Pi-alkyl
						ARG A:329	4.54	
						MET A:268	5.54	Pi-sulfur
						2AZ 5	-11	Pi-alkyl
						VAL B:123	5.11	
						LEU D:57	4.54	
						TYR C:119	4.09	Pi-donor H bond
						TYR C:119	4.60	C-H bond
10.	529518 92	17- hydroxyazadirad ione	Azadira chta indica	Geh A	-8.1	GLY A:298	2.96	Conventi onal H- bond
						HIS A:297	3.16	
						SER A:58	3.03	
						2AZ 5	-9.1	-

3.2 Drug likeliness

The pharmacokinetic properties (Absorption, Distribution, Metabolism, Excretion; ADME), drug-likeness, and toxicity profiles of selected ligands were evaluated using computational tools, namely SwissADME, Pro Tox II, and pkCSM. This investigation aimed to assess the suitability of these bioactive phytochemicals as potential drug candidates. Through in-silico methodologies, essential physicochemical characteristics governing the efficacy, safety, and metabolic fate of the molecules within the biological system were predicted. Notably, Lipinski's rule of five was employed as a pivotal criterion in rational drug design, facilitating the identification of compounds with drug-like properties. As per Lipinski's rule, optimal drug candidates typically possess molecular weights below 500 Daltons, LogP values less than 3, and fewer than 10 hydrogen bond acceptors, indicative of favorable oral absorption and permeability. The application of computational tools enabled the expedited assessment of prospective drug candidates, streamlining the process of drug discovery and development.

3.2.1 Table 2

Drug likeliness of the top 10 compounds.

Sr No.	Ligand	MF	Molecule PubChem CID	Lipinski's Rule of 5					Lipinski's Viol.	Lipinski's Rule
				MW (g/mol)	Log P	HB A	HB D	MR		
1	Epicatechin	C15H14O6	72276	290.27	1.5461	6	5	74.33	0	Yes
2	O-Caffeoyl quinic acid	C16H18O9	1794427	354.31	-0.6459	9	6	83.50	1	Yes
3	Strictinin	C27H22O18	73330	634.45	-0.2965	18	11	141.85	3	No
4	Valoneic acid dilactone	C21H10O13	10151874	470.30	2.2145	13	7	112.83	2	No
5	catechin	C15H14O6	9064	290.27	1.5461	6	5	74.33	0	Yes
6	Epigallocatechin Gallate	C22H18O11	65064	458.37	2.2332	11	8	112.06	2	No
7	Nimbiol	C18H24O2	11119228	272.38	4.37102	2	1	82.44	0	Yes
8	Nimboldide	C27H30O7	12313376	466.52	3.7431	7	0	120.99	0	Yes
9	7-Deacetyl-7-benzoyl poxyazadiradione	C33H36O6	52951893	528.64	5.923	6	0	144.86	1	Yes
10	17-Hydroxy azadiradione	C28H34O6	52951892	466.57	4.5219	6	1	126.52	0	Yes

3.3 ADMET features of selected ligands

pharmacokinetic properties of chemical compounds. The compounds selected as hits from the molecular docking study were subjected to an additional screening process to predict their respective ADMET properties, as shown in Table 3. Nine compounds out of the initial ten were chosen for further analysis as they exhibited toxicity levels falling within the acceptable range for drug candidacy. These selected compounds demonstrated predicted LD50 values and toxicity class that met the required criteria, making them suitable for advancing to the next stage of evaluation.

3.3.1 Table 3

ADMET properties of the top 10 compounds.

Ligand	In-silico ADMET								Toxicity
	Absorption		Distribution	Metabolism					
	Water Solubility	Skin Permeability	BBB permeability	2D6	3A4	1A2	2C19	2C9	Predicted LD50 & Toxicity Class:
Epicatechin	-3.117	-2.735	-1.054	No	No	No	No	No	10000 mg/kg (Class:6)
O-Caffeoylquinic acid	-2.449	-2.735	-1.407	No	No	No	No	No	5000 mg/kg (Class: 5)
Strictinin	-2.892	-2.735	-2.612	No	No	No	No	No	2260 mg/kg (Class: 5)
Valonic acid	-2.892	-2.735	-2.735	No	No	No	No	No	1000 mg/kg

dilactone									(Class: 4)
catechin	-3.117	-2.735	-1.054	No	No	No	No	No	10000 mg/kg (Class: 6)
Epigallocatechin Gallate	-2.894	-2.735	-2.184	No	No	No	No	No	1000 mg/kg (Class: 4)
Nimbiol	-4.077	-2.69	0.106	Yes	No	No	Yes	Yes	5000 mg/kg (Class: 5)
Nimbolide	-5.166	-3.599	-0.675	No	No	No	No	No	1000 mg/kg (Class: 4)
7-Deacetyl-7-benzoyl-7-oxepoxy-7-azadiene	-4.798	-2.809	-0.472	No	No	No	No	Yes	274 mg/kg (Class: 3)
17-Hydroxyazadiradione	-4.186	-3.149	-0.302	No	Yes	No	No	No	555 mg/kg (Class: 4)

3.4 PASS Evaluation:

3.4.1

Natural compounds often exhibit diverse chemico-biological properties, potentially leading to synergistic or antagonistic effects. To discern compounds that manifest both safety and efficacy with desirable attributes, it is imperative to scrutinize the biological properties of identified hit molecules. In this study, the biological properties of hit molecules were investigated utilizing Prediction of Activity Spectra for Substances (PASS) analysis. This analysis furnished insights into the potential properties of the compounds, with summarized results, including confidence levels, presented in Table 4. The findings revealed that the four selected compounds—Valoneic acid dilactone, Strictinin, Epigallocatechin Gallate, and Nimbiol—possess substantial potential for acne treatment, offering antioxidant, anti-inflammatory, astringent, dermatologic, and antiseborrheic benefits. PASS analysis corroborated these observations, affirming the considerable promise of these compounds for utilization in anti-acne therapeutic interventions.

3.4.2 Table 4

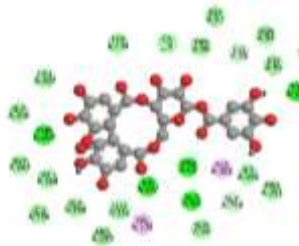
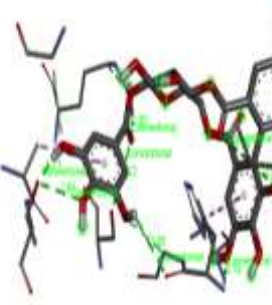
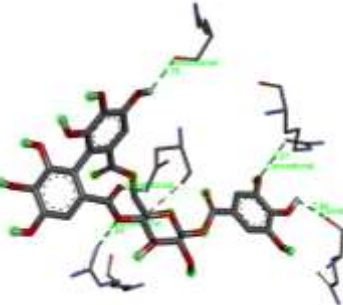
Biological properties of the elucidated phytoconstituents predicted through the PASS webserver.

Compound	Pa	Pi	Biological Activity
Epicatechin	0,737	0,031	Antiseborrheic
	0,548	0,044	Antiinflammatory
	0,810	0,003	Antioxidant
O-Caffeoylquinic acid	0,785	0,004	Antioxidant
	0,598	0,032	Antiinflammatory
	0,537	0,013	Antibacterial
	0,123	0,042	Antiacne
	0,479	0,036	Dermatologic
	0,840	0,003	Antioxidant
Strictinin	0,809	0,006	Antiinflammatory
	0,591	0,009	Antibacterial
	0,434	0,045	Dermatologic
	0,935	0,001	Astringent
Valoneic acid dilactone	0,809	0,006	Antiinflammatory
	0,707	0,004	Antioxidant
	0,449	0,022	Antibacterial
	0,426	0,087	Antiseborrheic
	0,548	0,044	Antiinflammatory
catechin	0,810	0,003	Antioxidant
	0,737	0,031	Antiseborrheic
	0,623	0,027	Antiinflammatory
Epigallocatechin Gallate	0,814	0,003	Antioxidant
	0,505	0,072	Antiseborrheic

Nimbiol	0,814	0,017	Antiseborrheic
	0,373	0,012	Antiacne
	0,644	0,024	Antiinflammatory
	0,636	0,012	Dermatologic
Nimbolide	0,371	0,111	Antiinflammatory
17-	0,118	0,044	Antiacne
Hydroxyazadiradione	0,532	0,048	Antiinflammatory
	0,417	0,089	Antiseborrheic
	0,575	0,019	Dermatologic

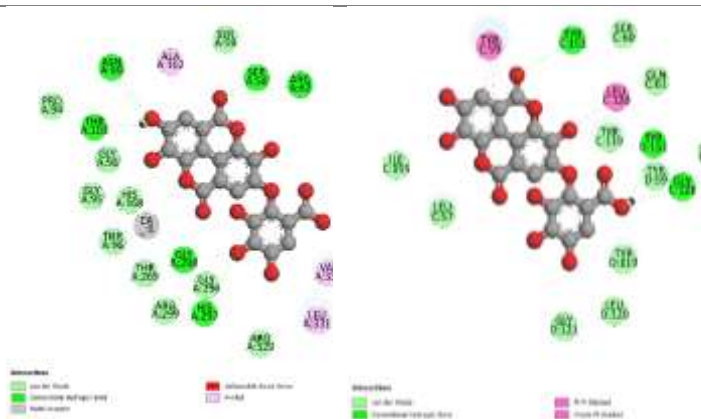
3.5 Interaction Analysis of Selected Compounds

The compounds' interactions with GehA and TNF α were analyzed using PyRx and Discovery Studio visualizer. The focus was on understanding how the compounds bind to the target proteins and identifying the specific residues they interact with. Through this analysis, hydrogen bonding and other types of interactions between the compounds and the proteins were visualized and studied. Discovery Studio and PyRx were utilized for this purpose, helping to gain insights into the binding modes and interaction patterns of the selected compounds with GehA and TNF α based on the interacting residues. [Table 5]

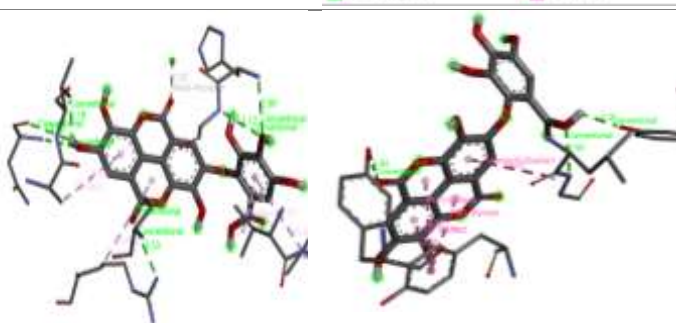
Ligand	Structure	Docked with	
		Geh-A	2AZ5
Strictinin	2D		
	3D		

**Valoneic
acid
dilactone**

2D

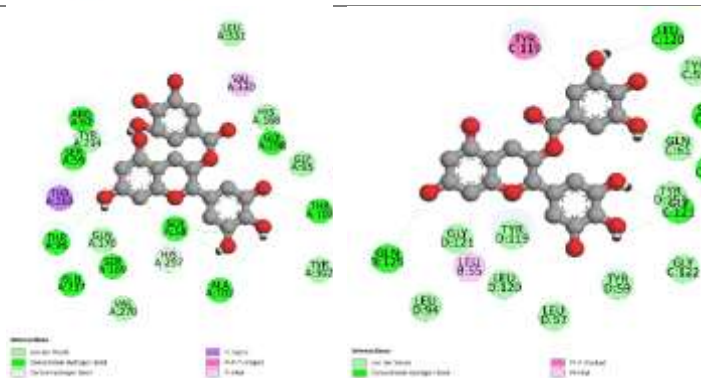


3D

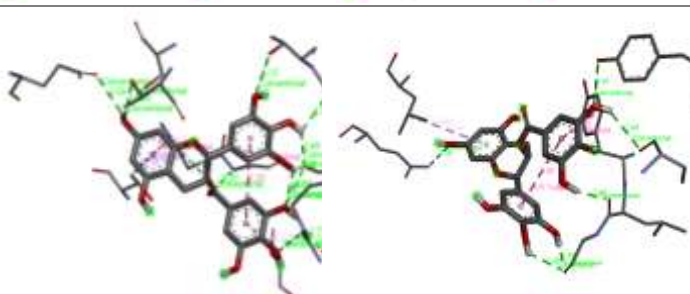


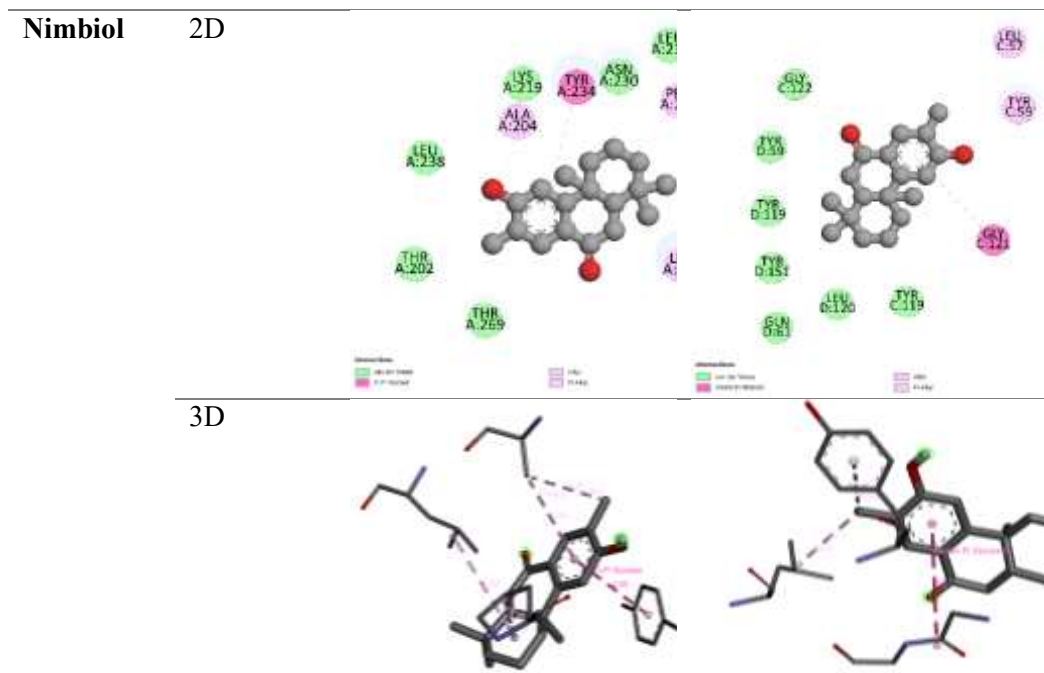
**Epigalloca
techin
Gallate**

2D



3D





4. Conclusion

This research highlights the promising potential of natural compounds derived from medicinal plants as therapeutic agents for acne management. By targeting key proteins *GehA* and *TNF-alpha* implicated in acne pathogenesis, our computational approach identified ten phytoconstituents with notable anti-acne activity. Among these, Valoneic acid dilactone, Strictinin, Epigallocatechin Gallate, and Nimbiol demonstrated significant biological activities encompassing anti-inflammatory, antioxidant, antimicrobial, and dermatological properties, making them attractive candidates for further development as anti-acne therapeutics. The systematic virtual screening approach employed in this study underscores the utility of computational methods in accelerating the discovery and development of novel acne treatments from natural sources. These findings pave the way for future investigations and potential translation of these natural compounds into effective clinical interventions for acne and related dermatological conditions.

CRedit authorship contribution statement

- Credit authorship contribution statement Pranali R. Pangam: Investigation, Conceptualization, Methodology, Visualization, Writing – original draft.
- Shubhangi B. Sutar: Conceptualization, Methodology, Writing – review & editing.
- Sachinkumar V. Patil: Supervision, Writing – review & editing.
- Sachin S. Mali: Supervision, Writing – review & editing.

Data availability

Data will be made available on request.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

5. References:

- [1] Y. F. Zhang et al., “Curcumin-mediated photodynamic therapy for mild to moderate Acne: A self-controlled split-face randomized study,” *Photodiagnosis Photodyn Ther*, vol. 45, Feb. 2024, doi: 10.1016/j.pdpdt.2023.103887.
- [2] L. Stein Gold et al., “Clindamycin phosphate 1.2%/adapalene 0.15%/benzoyl peroxide 3.1% gel for moderate-to-severe acne: Efficacy and safety results from two randomized phase 3 trials,” *J Am Acad Dermatol*, vol. 89, no. 5, pp. 927–935, Nov. 2023, doi: 10.1016/j.jaad.2022.08.069.
- [3] S. H. Cheng et al., “Multimodal imaging distribution assessment of a liposomal antibiotic in an infectious disease model,” *Journal of Controlled Release*, vol. 352, pp. 199–210, Dec. 2022, doi: 10.1016/j.jconrel.2022.08.061.
- [4] V. Durán et al., “Fucosylated lipid nanocarriers loaded with antibiotics efficiently inhibit mycobacterial propagation in human myeloid cells,” *Journal of Controlled Release*, vol. 334, pp. 201–212, Jun. 2021, doi: 10.1016/j.jconrel.2021.04.012.
- [5] D. Alimi, A. Hajri, S. Jallouli, and H. Sebai, “Toxicity, repellency, and anti-cholinesterase activities of bioactive molecules from clove buds *Syzygium aromaticum* L. as an ecological alternative in the search for control *Hyalomma scupense* (Acari: Ixodidae),” *Heliyon*, vol. 9, no. 8, Aug. 2023, doi: 10.1016/j.heliyon.2023.e18899.
- [6] D. Das, M. Nanda, P. Banjare, and S. Lanjhiyana, “Exploration of multitargeted antialzheimer’s activity of safflower leaves phytoconstituents: In silico molecular docking approach,” *European Journal of Medicinal Chemistry Reports*, vol. 10, Apr. 2024, doi: 10.1016/j.ejmcr.2023.100119.
- [7] J. Wambui, R. I. O. Ikedi, R. W. Macharia, F. Kama-Kama, and E. N. Nyaboga, “Phytoconstituents of Kenyan stinging nettle (*Urtica* species) and their molecular docking interactions revealed anti-inflammatory potential as cyclooxygenase-2 inhibitors,” *Sci Afr*, vol. 23, Mar. 2024, doi: 10.1016/j.sciaf.2024.e02088.
- [8] S. K. Mandal et al., “In silico anti-viral assessment of phytoconstituents in a traditional (Siddha Medicine) polyherbal formulation – Targeting Mpro and pan-coronavirus post-fusion Spike protein,” *J Tradit Complement Med*, vol. 14, no. 1, pp. 55–69, Jan. 2024, doi: 10.1016/j.jtcme.2023.07.004.
- [9] T. Casalini, “Not only in silico drug discovery: Molecular modeling towards in silico drug delivery formulations,” *Journal of Controlled Release*, vol. 332. Elsevier B.V., pp. 390–417, Apr. 10, 2021. doi: 10.1016/j.jconrel.2021.03.005.
- [10] D. Nieoczym et al., “A comprehensive assessment of palmitine as anticonvulsant agent – In vivo and in silico studies,” *Biomedicine and Pharmacotherapy*, vol. 172, Mar. 2024, doi: 10.1016/j.biopha.2024.116234.
- [11] N. Zheng et al., “Utilizing the photodynamic properties of curcumin to disrupt biofilms in *Cutibacterium acnes*: A promising approach for treating acne,” *Photodiagnosis Photodyn Ther*, vol. 45, Feb. 2024, doi: 10.1016/j.pdpdt.2023.103928.
- [12] Y. Wang et al., “The Anti-Inflammatory Activities of *Propionibacterium acnes* CAMP Factor-Targeted Acne Vaccines,” *Journal of Investigative Dermatology*, vol. 138, no. 11, pp. 2355–2364, Nov. 2018, doi: 10.1016/j.jid.2018.05.032.
- [13] R. Han, H. M. Blencke, H. Cheng, and C. Li, “The antimicrobial effect of CEN1HC-Br against *Propionibacterium acnes* and its therapeutic and anti-inflammatory effects on acne vulgaris,” *Peptides (N.Y.)*, vol. 99, pp. 36–43, Jan. 2018, doi: 10.1016/j.peptides.2017.11.001.

- [14] K. Ngamsou Abdel et al., “Ethnobotanical study and vulnerability of medicinal plants used against the symptoms of COVID-19 in the Lomié subdivision, East Region of Cameroon,” *Heliyon*, vol. 10, no. 7, p. e28247, Apr. 2024, doi: 10.1016/j.heliyon.2024.e28247.
- [15] Y. T. M. Alharbi et al., “Investigation of phytochemicals isolated from selected Saudi medicinal plants as natural inhibitors of SARS CoV-2 main protease: In vitro, molecular docking and simulation analysis,” *Saudi Pharmaceutical Journal*, p. 102023, Mar. 2024, doi: 10.1016/j.jsps.2024.102023.
- [16] X. Yang et al., “High-throughput RNA sequencing reveals the anti-inflammatory mechanism of baicalin on *Propionibacterium acnes*-induced acne in rabbits,” *Journal of Traditional Chinese Medical Sciences*, vol. 6, no. 3, pp. 201–210, Jul. 2019, doi: 10.1016/j.jtcms.2019.09.001.
- [17] G. P. Khumalo, T. Nguyen, B. E. Van Wyk, Y. Feng, and I. E. Cock, “Inhibition of pro-inflammatory cytokines by selected southern African medicinal plants in LPS-stimulated RAW 264.7 macrophages,” *J Ethnopharmacol*, vol. 319, Jan. 2024, doi: 10.1016/j.jep.2023.117268.
- [18] A. N. Tavanappanavar et al., “Phytochemical analysis, GC–MS profile and determination of antibacterial, antifungal, anti-inflammatory, antioxidant activities of peel and seeds extracts (chloroform and ethyl acetate) of *Tamarindus indica* L,” *Saudi J Biol Sci*, vol. 31, no. 1, Jan. 2024, doi: 10.1016/j.sjbs.2023.103878.
- [19] Y. G. Kim, J. H. Lee, and J. Lee, “Antibiofilm activities of fatty acids including myristoleic acid against *Cutibacterium acnes* via reduced cell hydrophobicity,” *Phytomedicine*, vol. 91, Oct. 2021, doi: 10.1016/j.phymed.2021.153710.
- [20] M. Bharathithasan, V. Kotra, S. Atif Abbas, and A. Mathews, “Review on biologically active natural insecticides from Malaysian tropical plants against *Aedes aegypti* and *Aedes albopictus*,” *Arabian Journal of Chemistry*, vol. 17, no. 1. Elsevier B.V., Jan. 01, 2024. doi: 10.1016/j.arabjc.2023.105345.
- [21] E. Ramsay et al., “Selective drug delivery to the retinal cells: Biological barriers and avenues,” *Journal of Controlled Release*, vol. 361, pp. 1–19, Sep. 2023, doi: 10.1016/j.jconrel.2023.07.028.
- [22] I. Domingues et al., “Exploiting the biological effect exerted by lipid nanocapsules in non-alcoholic fatty liver disease,” *Journal of Controlled Release*, vol. 356, pp. 542–553, Apr. 2023, doi: 10.1016/j.jconrel.2023.03.012.
- [23] A. Mahal et al., “Molecular docking, drug-likeness and DFT study of some modified tetrahydrocurcumins as potential anticancer agents,” *Saudi Pharmaceutical Journal*, vol. 32, no. 1, Jan. 2024, doi: 10.1016/j.jsps.2023.101889.
- [24] M. Mandal and S. Mandal, “Discovery of multitarget-directed small molecule inhibitors from *Andrographis paniculata* for Nipah virus disease therapy: Molecular docking, molecular dynamics simulation and ADME-Tox profiling,” *Chemical Physics Impact*, vol. 8, Jun. 2024, doi: 10.1016/j.chphi.2024.100493.
- [25] S. Kanwal et al., “Chemical Profiling, in-vitro biological evaluation and molecular docking studies of *Ruellia tweediana*: An unexplored plant,” *Saudi Pharmaceutical Journal*, vol. 32, no. 2, Feb. 2024, doi: 10.1016/j.jsps.2023.101939.
- [26] R. Y. Halayal, Z. K. Bagewadi, N. A. Aldabaan, I. A. Shaikh, and A. A. Khan, “Exploring the therapeutic mechanism of potential phytochemicals from *Kalanchoe pinnata* in the treatment of diabetes mellitus by integrating network pharmacology, molecular docking and simulation approach,” *Saudi Pharmaceutical Journal*, p. 102026, Mar. 2024, doi: 10.1016/j.jsps.2024.102026.
- [27] M. K. Zubko, “Towards sustainable antimicrobials from plants: Some ways to abridge current methodological approaches,” *Sustainable Materials and Technologies*, vol. 39, Apr. 2024, doi: 10.1016/j.susmat.2023.e00801.

- [28] S. Rehman et al., "Ethno-Dentistry of Medicinal Plants Used in North Waziristan, Pakistan," *Int Dent J*, Apr. 2023, doi: 10.1016/j.identj.2023.10.001.
- [29] L. L. Denga, B. Diedericks, A. M. Kok, and N. Lall, "The antithrombotic potential of selected South African plants for venous thromboembolism," *South African Journal of Botany*, vol. 167, pp. 209–216, Apr. 2024, doi: 10.1016/j.sajb.2024.02.028.
- [30] A. Arrout, Y. El Ghallab, M. Yafout, M. R. Lefriyekh, and A. A. H. Said, "Medicinal plants for gallstones: A cross-sectional survey of Moroccan patients," *Phytomedicine Plus*, vol. 4, no. 1, Feb. 2024, doi: 10.1016/j.phyplu.2024.100524.
- [31] M. Shabbir, M. Singh, S. Maiti, and S. K. Saha, "Organophosphate pesticide (Chlorpyrifos): Environmental menace; study reveals genotoxicity on plant and animal cells," *Environmental Challenges*, vol. 5, Dec. 2021, doi: 10.1016/j.envc.2021.100313.
- [32] A. Wendimu and W. Tekalign, "Field efficacy of ethnomedicinal plant smoke repellency against *Anopheles arabiensis* and *Aedes aegypti*," *Heliyon*, vol. 7, no. 7, Jul. 2021, doi: 10.1016/j.heliyon.2021.e07373.
- [33] J. Cao et al., "Graphene enhances artemisinin production in the traditional medicinal plant *Artemisia annua* via dynamic physiological processes and miRNA regulation," *Plant Commun*, vol. 5, no. 3, Mar. 2024, doi: 10.1016/j.xplc.2023.100742.
- [34] A. Wendimu, W. Tekalign, E. Bojago, and Y. Abrham, "Traditional ethnobotanical knowledge and ethnomedicinal use of plants in the Tropical Rift Valley of Ethiopia," *Heliyon*, vol. 10, no. 6, Mar. 2024, doi: 10.1016/j.heliyon.2024.e27528.
- [35] S. Zhao et al., "Screening and identifying natural products with SARS-CoV-2 infection inhibitory activity from medicinal fungi," *Biosaf Health*, vol. 6, no. 1, pp. 12–20, Feb. 2024, doi: 10.1016/j.bsheal.2023.12.006.
- [36] Y. L. Ji et al., "Chemical characterization of different parts of *Forsythia suspensa* and α -glucosidase and pancreatic lipase inhibitors screening based on UPLC-QTOF-MS/MS and plant metabolomics analysis: Chemical characterization of different parts of *Forsythia suspensa* and α -glucosidase and pancreatic lipase inhibitors screening based on UPLC-QTOF-MS/MS," *Arabian Journal of Chemistry*, vol. 17, no. 5, May 2024, doi: 10.1016/j.arabjc.2024.105723.
- [37] N. F. Ilahibaks et al., "TOP-EVs: Technology of Protein delivery through Extracellular Vesicles is a versatile platform for intracellular protein delivery," *Journal of Controlled Release*, vol. 355, pp. 579–592, Mar. 2023, doi: 10.1016/j.jconrel.2023.02.003.
- [38] M. Danielsen, C. Hempel, T. L. Andresen, and A. J. Urquhart, "Biopharmaceutical nanoclusters: Towards the self-delivery of protein and peptide therapeutics," *Journal of Controlled Release*, vol. 347. Elsevier B.V., pp. 282–307, Jul. 01, 2022. doi: 10.1016/j.jconrel.2022.04.050.
- [39] D. Witzigmann et al., "Non-viral gene delivery of the oncotoxic protein NS1 for treatment of hepatocellular carcinoma," *Journal of Controlled Release*, vol. 334, pp. 138–152, Jun. 2021, doi: 10.1016/j.jconrel.2021.04.023.
- [40] T. W. Liu, S. J. Hsu, Y. S. Y. Hsieh, H. K. Liu, and C. K. Lee, "Polymethoxyflavone from *Citrus depressa* as an inhibitor against various variants of SARS-CoV-2 spike protein," *J Ethnopharmacol*, vol. 320, Feb. 2024, doi: 10.1016/j.jep.2023.117412.
- [41] C. T. Namba-Nzanguim et al., "Investigation of some plant stilbenoids and their fragments for the identification of inhibitors of SARS-CoV-2 viral spike/ACE2 protein binding," *The Microbe*, vol. 3, p. 100059, Jun. 2024, doi: 10.1016/j.microb.2024.100059.
- [42] Y.-T. Zhang, H.-W. Liu, C. Song, B. Li, L. Yang, and G.-R. Wang, "Identification of transient receptor potential channel genes and functional characterization of TRPA1 in *Spodoptera frugiperda*." [Online]. Available: <http://transdecoder.sourceforge.net/>

- [43] S. Hirsch et al., “Hepatic targeting of the centrally active cannabinoid 1 receptor (CB1R) blocker rimonabant via PLGA nanoparticles for treating fatty liver disease and diabetes,” *Journal of Controlled Release*, vol. 353, pp. 254–269, Jan. 2023, doi: 10.1016/j.jconrel.2022.11.040.
- [44] K. Lyman, T. Kelley, J. Walthall, S. D. Lang, B. B. Gilmer, and D. Guttman, “Refractory shoulder injury related to vaccine administration: correlation with culture presence of *Cutibacterium acnes*,” *JSES Reviews, Reports, and Techniques*, vol. 3, no. 3, pp. 350–355, Aug. 2023, doi: 10.1016/j.xrrt.2023.02.011.
- [45] A. M. O'Neill et al., “Genetic and Functional Analyses of *Cutibacterium Acnes* Isolates Reveal the Association of a Linear Plasmid with Skin Inflammation,” in *Journal of Investigative Dermatology*, Elsevier B.V., Jan. 2024, pp. 116–124.e4. doi: 10.1016/j.jid.2023.05.029.
- [46] C. Folle et al., “Colloidal hydrogel systems of thymol-loaded PLGA nanoparticles designed for acne treatment,” *Colloids Surf B Biointerfaces*, vol. 234, Feb. 2024, doi: 10.1016/j.colsurfb.2023.113678.
- [47] P. Ju et al., “Analysis on the treatment of acne by Sanhuang with network pharmacology and experimental research,” 2022. [Online]. Available: <https://www>.
- [48] M. ping Wei, H. Yu, Y. hui Guo, Y. liang Cheng, Y. fei Xie, and W. rong Yao, “Potent in vitro synergistic antibacterial activity of natural amphiphilic Sapindoside A and B against *Cutibacterium acnes* with destructive effect on bacterial membrane,” *Biochim Biophys Acta Biomembr*, vol. 1863, no. 11, Nov. 2021, doi: 10.1016/j.bbamem.2021.183699.
- [49] Z. Li, S. Zhang, and Y. Wang, “Inhibition of *Propionibacterium acnes* by refined bamboo vinegar and preparation of the slow-release system with bamboo charcoal as the carrier,” *Journal of Dermatologic Science and Cosmetic Technology*, p. 100016, Feb. 2024, doi: 10.1016/j.jdsct.2024.100016.
- [50] I. Batubara, I. Julita, L. K. Darusman, A. M. Muddathir, and T. Mitsunaga, “Flower Bracts of Temulawak (*Curcuma Xanthorrhiza*) for Skin Care: Anti-acne and Whitening Agents,” *Procedia Chem*, vol. 14, pp. 216–224, 2015, doi: 10.1016/j.proche.2015.03.031.
- [51] M. Rihova et al., “Biopolymeric fibers prepared by centrifugal spinning blended with ZnO nanoparticles for the treatment of *Acne vulgaris*,” *Appl Mater Today*, vol. 37, Apr. 2024, doi: 10.1016/j.apmt.2024.102151.
- [52] T. Falla, K. Rodan, K. Fields, D. Ong, and C. Skobowiat, “Safety and efficacy of a novel three-step anti-acne regimen formulated specifically for women,” *Int J Womens Dermatol*, vol. 6, no. 5, pp. 419–423, Dec. 2020, doi: 10.1016/j.ijwd.2020.07.013.
- [53] X. Ren et al., “Integrative network pharmacology and transcriptomics analysis reveal the mechanism of Tanreqing in the treatment of *acne vulgaris*,” *Phytomedicine Plus*, vol. 4, no. 1, Feb. 2024, doi: 10.1016/j.phyplu.2023.100505.
- [54] S. F. Ruan et al., “Potential role of mTORC1 and the PI3K-Akt pathway in anti-acne properties of licorice flavonoids,” *J Funct Foods*, vol. 70, Jul. 2020, doi: 10.1016/j.jff.2020.103968.
- [55] J. Zhang et al., “A novel natural polysaccharide dissolving microneedle capable of adsorbing pus to load EGCG for the treatment of *acne vulgaris*,” *Mater Des*, vol. 238, Feb. 2024, doi: 10.1016/j.matdes.2024.112639.