

Antimicrobial Activity and Phytochemical Constituents of Leaf and Stem Extracts of *Tinospora Sinensis*

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Currently, the pursuit of plant extracts exhibiting potent antimicrobial properties has heightened. As a result of *Tinospora sinensis* being employed in traditional medicine for combating inflammatory and infectious conditions, this research assessed the in vitro antimicrobial capabilities and phytochemical makeup of extracts derived from its stem and leaves. Phytochemical screening was conducted utilizing established reference methods. The antimicrobial activities of various extracts were assessed through the agar well diffusion method, while the antibiotic susceptibility of selected microorganisms was examined using the disc diffusion method. The preliminary examination of the phytochemical profile reveals that the various extracts in the formulation encompass a wide array of vital phytochemicals, including alkaloids, flavonoids, glycosides, cardiac glycosides, tannins, phenols, steroids, quinones, proteins, carbohydrates, and saponins. The leaf and stem displayed sufficient activity, with the leaf exhibiting the highest level of activity. The chloroform extracts showed the highest activity, followed by aqueous, acetone, ethyl acetate, and methanol extracts. *Staphylococcus aureus* was found to be more susceptible to both leaf and stem extracts, followed by *Bacillus subtilis*, *Pseudomonas fluorescens*, and *Streptococcus mutans*, respectively, at a concentration of 10mg/l. These findings support the exploration of active components that may serve as lead molecules in the creation of novel antimicrobial medications.

1. Introduction

Phytochemical analysis is an essential area of study that focuses on identifying and characterizing the chemical compounds found in medicinal plants. These compounds, known as phytochemicals, play a critical role in the therapeutic properties of plants that have been used for centuries in traditional medicine. By examining these unique biochemical constituents, researchers can uncover how specific plants can support health and treat various ailments. Different methods, such as chromatography and mass spectrometry, are employed to separate and analyze these compounds, providing a clearer understanding of their biological activities. This analysis is vital not only for validating traditional remedies but also for discovering new pharmaceutical drugs derived from natural sources. As the interest in natural products continues to grow, phytochemical analysis holds promise for enhancing our understanding of how plants can be leveraged for better health and well-being. The process of

phytochemical analysis offers exciting opportunities for both scientific advancement and the future of medicine. As we uncover the complex interactions between phytochemicals and biological systems, we gain insights into how these natural compounds can influence health outcomes. This research is particularly important in the context of increasing antibiotic resistance and the need for novel treatments. The antimicrobial activity of medicinal plants is a fascinating and essential area of study that focuses on the natural ability of various plants to combat harmful microorganisms. These microorganisms, which include bacteria, viruses, fungi, and parasites, can lead to a range of health issues, making it crucial to find effective ways to eliminate them. Medicinal plants have been used for centuries in traditional medicine, often being trusted sources of healing. Many of these plants contain bioactive compounds that possess antimicrobial properties, allowing them to inhibit the growth and spread of these pathogens. For example, substances like essential oils, alkaloids, and flavonoids in certain herbs and spices can disrupt the cell walls of bacteria or interfere with their metabolic processes. As antibiotic resistance becomes an increasingly serious global health concern, exploring these natural alternatives is gaining importance.

2. Material and methods

Plant description

Tinospora species are succulent woody climbing shrubs, gracefully extending long aerial roots from host trees, creating a beautiful spread (figure 1). The bark is corky or papery, exfoliating as it dries. Leaves are simple, alternate, broadly ovate, and subcordate. The margin is either entire or dentate, and they are not peltate. They have 3-7 basal veins and 1-3 pairs of lateral veins. The petioles are geniculate and swollen near the base. Inflorescences are situated either axillary or on old leafless branches, appearing as pseudoracemes, pseudopaniculate, or thyrsoid. They are few-flowered and have peduncles. Flowers are unisexual and pedicellate. Male flowers consist of six sepals in two series, with the inner series being the largest and the outer series the smallest. They are free, imbricate, and subelliptic, with an acute apex. The flowers also have three to six free petals that are obovate with a cuneate apex and inrolled lateral edges. Additionally, there are six stamens with free filaments, anthers that are loculed, subextorse, and either obliquely or longitudinally dehiscent. In female flowers, the sepals and petals are similar to those in male flowers. The staminodes are six in number and subulate, while the carpels are three and ellipsoid-curved. The style is short and thick, with the stigma reflexed and peltate. Fruits drupes, subglobose or ellipsoid, with a columnar carpophore. The endocarp is hard, bony, dorsally convex, often verrucose, deeply ruminant, and the ventral side has a central aperture. Seeds are curved or half-moon shaped, endospermic, with flattened cotyledons that are leaflike, and a short radicle.



Figure 1: *Tinospora sinensis* (Lour.) Merr..

1. Phytochemical analysis

Phytochemical analysis is a crucial process used to identify the various chemical compounds present in plant extracts. This procedure typically begins with the collection of plant materials, which should be carefully selected based on the intended study. After harvesting, the plant parts, such as leaves, stems, or roots, should be thoroughly washed to remove any dirt or contaminants, and then dried to reduce moisture content. Once dried, the plant materials are ground into a fine powder to increase the surface area, facilitating the extraction of phytochemicals (figure 2-4). The extraction process often employs solvents, like ethanol, methanol, or water, which dissolve the desired compounds (figure 3). The powdered plant material is mixed with the solvent and allowed to sit for a specified period to enable the phytochemicals to leach into the solution. After the extraction time has elapsed, the mixture is filtered to separate the solid debris from the liquid extract. The resulting extract can then be concentrated further, if necessary, through evaporation of the solvent, ultimately yielding concentrated sample rich in phytochemicals ready for subsequent analysis.



Figure 2: Dried stem and leaves of *Tinospora sinensis* (Lour.) Merr..

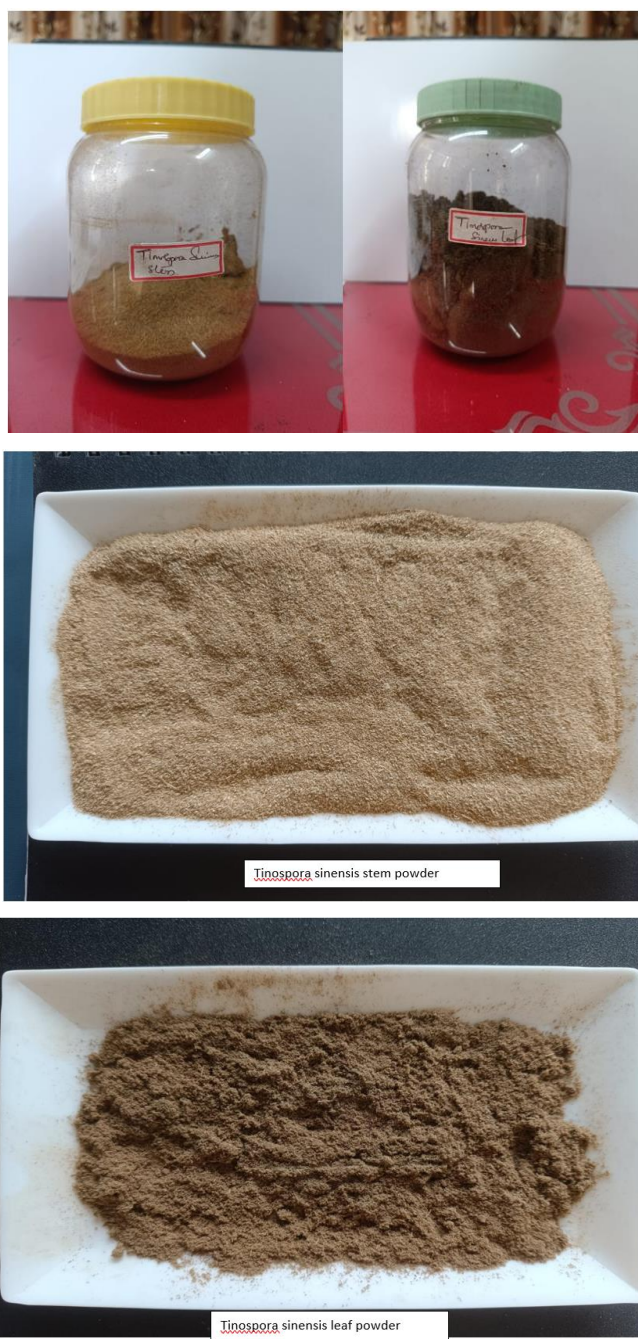


Figure 3: Dried stem and leaves powders of *Tinospora sinensis* (Lour.) Merr..



Figure 4: Stem and leaves extracts of *Tinospora sinensis* (Lour.) Merr..

The next step in phytochemical analysis involves qualitative and quantitative assessments of

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the extracted compounds. Qualitative analysis is primarily focused on identifying the various classes of phytochemicals present in the extract, such as Carbohydrate, Tannin, Saponin, Flavonoid, Alkaloid, Quinone, Glycoside, Cardiac Glycoside, Terpenoid, Phenol and Protein. This is typically conducted using specific chemical tests established by Harborne (1973) and Sofowora (1993), where reagents are added to the extract to observe any reactions or color changes, indicating the presence of certain phytochemical classes (1,2). For example, a color change when a particular reagent is added may suggest the presence of flavonoids. Collectively, these analytical techniques give researchers a comprehensive understanding of the phytochemical profile, laying the groundwork for further studies in pharmacology and toxicology. After the phytochemical analysis is completed, the results can be interpreted to draw meaningful conclusions about the biological significance and potential applications of the plant extract.

2. Antimicrobial activity

Antimicrobial activity is an important area of study, particularly when researching plant extracts that may offer natural solutions to combat various pathogens. To begin the procedure for assessing the antimicrobial activity of a plant extract, the first step is to prepare the plant material. This entails harvesting the plant parts, such as leaves, roots, or flowers, and drying them if necessary. The dried plant material is then ground into a fine powder, which allows for maximum surface area to extract the bioactive compounds. Following this, the powdered material is performed for Soxhlet extraction with different solvents like ethanol or methanol for a specific duration. This process, known as maceration or infusion, helps to dissolve the desired antimicrobial compounds from the plant material. Once the extraction is complete, the mixture is filtered to separate the liquid extract from the solid residues. The collected extract can then be concentrated through evaporation if a more potent solution is desired. This preparation is crucial, as the quality and concentration of the extract will greatly influence the results of the antimicrobial tests that follow. The next step in the antimicrobial activity procedure is to conduct various tests to evaluate the effectiveness of the plant extract against specific microorganisms. In laboratory settings, common methods include disk diffusion and broth micro dilution. Disk diffusion involves placing filter paper disks soaked in the plant extract on an agar plate that has been inoculated with bacteria or fungi. The agar cup procedure is a widely used method to test the effectiveness of plant extracts against various microorganisms like bacteria and fungi. This process begins with preparing an agar plate, which serves as the growth medium for the microorganisms. Small wells, or cups, are then made in the agar using a specialized tool. After this, the plant extract, which may come from leaves, stems, or roots, is introduced into these wells. The agar plate is then inoculated with the target microorganisms, allowing them to spread and grow across the surface. After incubation, if the extract possesses antimicrobial properties, clear zones called inhibition zones will appear around the well, indicating areas where microbial growth has been prevented. On the other hand, broth micro dilution assesses the minimum inhibitory concentration (MIC) of the extract, which is the lowest concentration capable of inhibiting microbial growth. In this method, varying concentrations of the plant extract are added to separate tubes containing a liquid growth medium and inoculated with the target microorganisms. By analyzing the turbidity of the medium after incubation, researchers can determine the extract's efficacy. These tests provide valuable information about the potential of the plant extract to serve as a

natural antimicrobial agent, offering insights into its application in medicine or agriculture. After conducting the antimicrobial tests, the final phase of the procedure involves analyzing and interpreting the data obtained. It is critical to compare the results of the plant extract with those of standard antibiotics to evaluate the relative effectiveness of the extract. Statistical analysis may be employed to establish significance and reproducibility of the findings.

3. Results

In the present study *Tinospora sinensis* leaf and stem extracts i.e., chloroform, aqueous, acetone, Ethyl acetate and methanol was evaluated for phytochemical and antibacterial analysis.

The initial analysis of the phytochemical profile indicates that the different extracts of the formulation contain a diverse mix of essential phytochemicals, such as alkaloids, flavonoids, glycosides, cardiac glycosides, tannins, phenols, steroids, quinones, proteins, carbohydrates, and saponins as shown in Table 1 and figure 5. Medicinal plants and herbs contain essential phytochemicals, a diverse range of primary and secondary plant metabolites. These compounds are behind various biological activities like anti-hyperglycemic, anti-inflammatory, anti-diabetic, and anti-microbial effects [3,4].

Table1: Phytochemical analysis of *Tinospora sinensis* (Lour.) Merr.

S. NO	Phytochemical test	Stem extract Acetone	Stem extract methnol	Stem extract ethyl acetate	Stem extract chloroform	Leaf extract Acetone	Leaf extract methnol	Leaf extract ethyl acetate	Leaf extract chloroform
1.	Alkaloids	Present	Present	Present	Present	Present	Present	Present	Absent
2.	Flavanoids	Absent	Absent	Absent	Absent	Absent	Absent	Absent	Present
3.	Glycosides	Present	Present	Present	Present	Absent	Absent	Absent	Absent
4.	Cardiac glycosides	Present	Present	Present	Present	Absent	Absent	Absent	Absent
5.	Tannins	Absent	Absent	Absent	Absent	Absent	Present	Present	Absent
6.	Phenols	Present	Present	Present	Present	Absent	Present	Present	Present
7.	Steroids	Present	Present	Absent	Absent	Absent	Absent	Absent	Absent
8.	Quniones	Absent	Absent	Absent	Absent	Present	Present	Present	Present
9.	Proteins	Absent	Present	Absent	Absent	Present	Present	Present	Present
10.	Carbohydrates	Present	Present	Present	Present	Present	Present	Present	Present
11.	Saponins	Present	Present	Present	Present	Present	Present	Present	Present

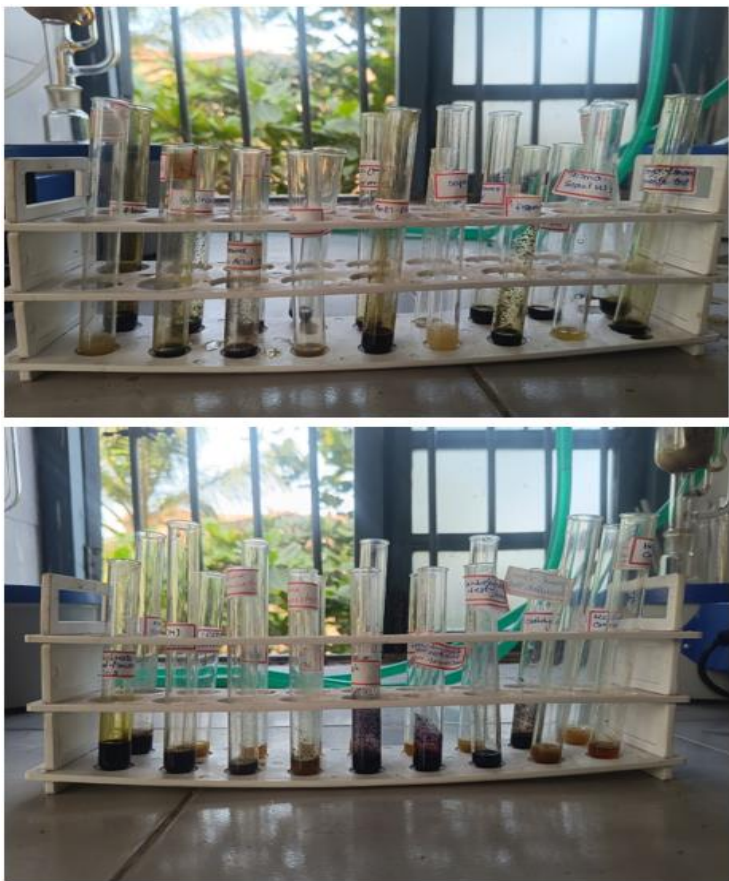


Figure 5: Phytochemical analysis of *Tinospora sinensis* (Lour.) Merr..

All the extracts confirm the presence of alkaloids, except for the leaf chloroform extract. All extracts confirm the lack of flavonoids, except for the leaf chloroform extract. All stem extracts indicate the presence of glycosides and cardiac glycosides, while leaf extracts lack them. All extracts confirm the absence of tannins, with the exception of leaf methanol and ethyl acetate extracts. All extracts have been verified to contain phenols, with the exception of the leaf acetone extract. All extracts confirm the absence of steroids except for the stem methanol and acetone extract. All stem extracts confirm the absence of quinones and proteins and are present in leaf extracts. All extracts confirm the presence of carbohydrates and saponins.

Leaf and stem showed adequate activity among both leaf showed the highest activity. Highest activity was observed chloroform extracts followed by aqueous, acetone, Ethyl acetate and methanol extracts respectively whereas *Staphylococcus aureus* was more susceptible to both leaf and stem extracts followed by *Bacillus subtilis*, *Pseudomonas fluorescens* and *Streptococcus mutans* respectively at 10mg/l concentration given in table 2 (Figure 6-10).

Table 2: Antibacterial analysis of *Tinospora sinensis* (Lour.) Merr.

Sample	Bacillus subtilis			Staphylococcus aureus			Streptococcus mutans			Pseudomonas fluorescens		
	10Mg	5Mg	2.5Mg	10Mg	5Mg	2.5Mg	10Mg	5Mg	2.5Mg	10Mg	5Mg	2.5Mg
ML	19	16	15	20	18	15	17	14	11	17	12	11

MS	18	17	14	19	18	10	18	12	9	15	11	10
CL	26	20	17	22	10	15	15	11	8	19	16	10
CS	18	14	13	20	13	12	19	15	9	14	13	8
EL	19	15	13	19	17	11	17	14	4	19	14	12
ES	20	14	12	26	16	9	15	11	0	18	15	13
Aq. L	23	18	18	24	21	17	19	16	11	21	17	11
Aq. S	23	19	17	19	20	17	18	15	11	18	14	13
AL	21	18	15	20	16	15	14	11	0	16	15	9
AS	16	14	16	22	13	11	17	14	10	20	12	10

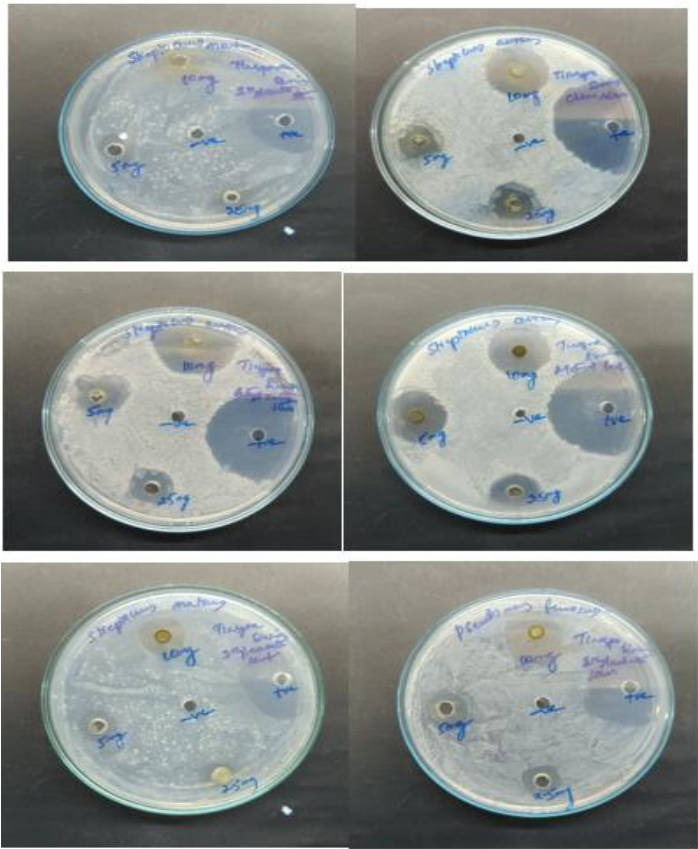


Figure 6: Antibacterial activity of Tinospora sinensis methanol extract

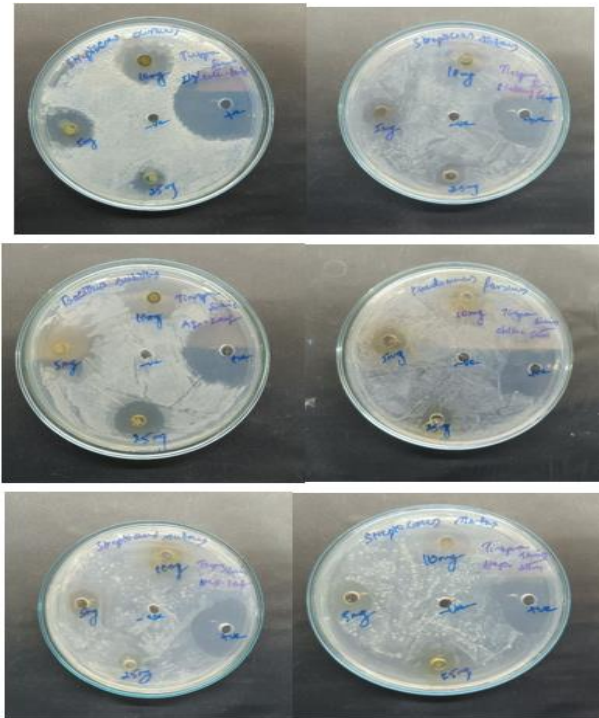


Figure 7: Antibacterial activity of Tinospora sinensis chloroform extract

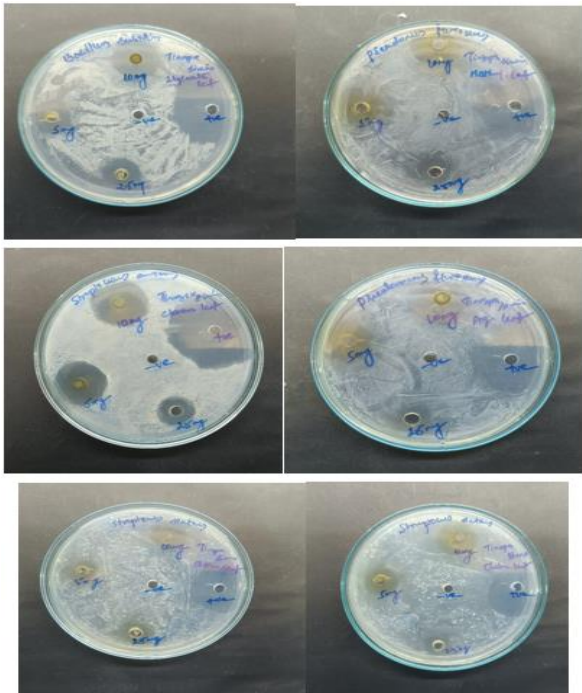


Figure 8: Antibacterial activity of Tinospora sinensis Ethyl acetate extract

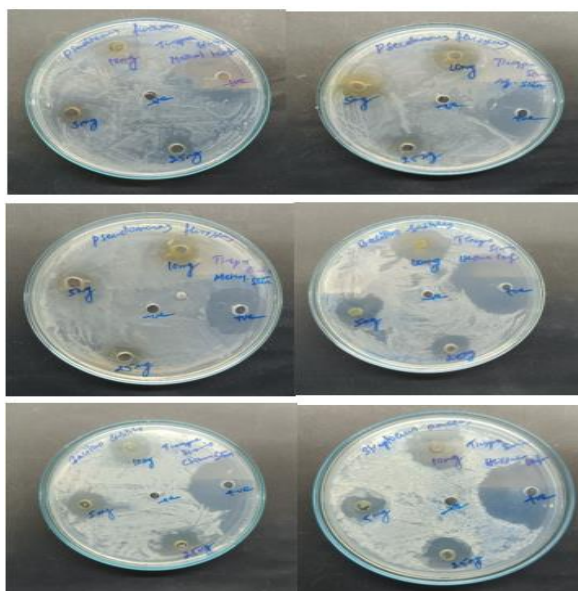


Figure 9: Antibacterial activity of *Tinospora sinensis* Aqueous extract

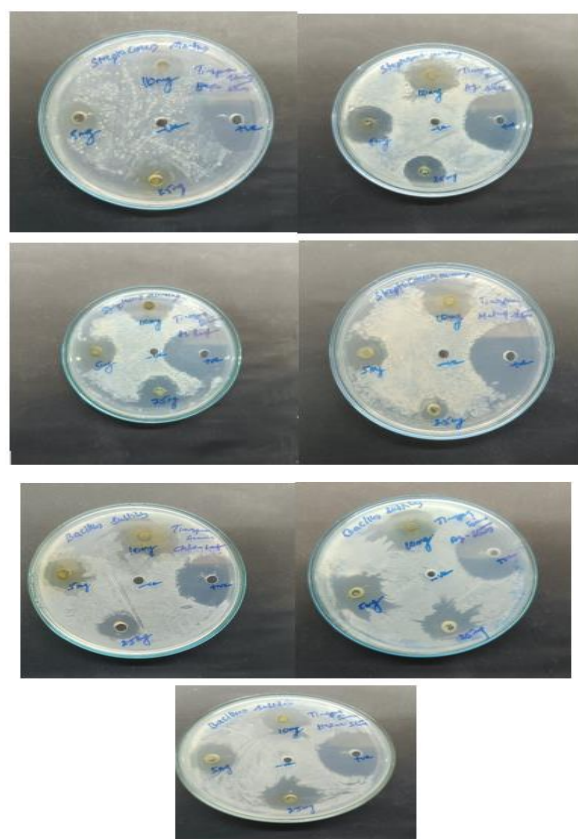


Figure 10: Antibacterial activity of *Tinospora sinensis* Acetone extract

Staphylococcus aureus (20 mm) exhibited the largest zone of inhibition in the methanolic leaf extract of *Tinospora sinensis*, followed by *Bacillus subtilis* (19 mm), *Pseudomonas fluorescens*, and *Streptococcus mutans* (17 mm). The stem extract showed maximum zone of inhibition in *Staphylococcus aureus* (19 mm) followed by *Bacillus subtilis* and *Streptococcus mutans* (19 mm) and *Pseudomonas fluorescens* (15 mm).

Bacillus subtilis (26 mm) exhibited the largest zone of inhibition in the chloroform leaf extract of *Tinospora sinensis*, followed by *Staphylococcus aureus* (22 mm), *Pseudomonas fluorescens* (19 mm), and *Streptococcus mutans* (17 mm). *Bacillus subtilis* (18 mm), *Pseudomonas fluorescens* (14 mm), *Streptococcus mutans* (19 mm), and *Staphylococcus aureus* (20 mm) exhibited the maximum zone of inhibition in the stem extract.

Pseudomonas fluorescens, *Staphylococcus aureus*, and *Bacillus subtilis* demonstrated the largest zone of inhibition in the ethyl acetate leaf extract of *Tinospora sinensis* (19 mm), followed by *Streptococcus mutans* (17 mm). *Staphylococcus aureus* (26 mm) exhibited the largest zone of inhibition in the stem extract, followed by *Bacillus subtilis* (20 mm), *Pseudomonas fluorescens* (18 mm), and *Streptococcus mutans* (15 mm), in that order.

Bacillus subtilis (21 mm) displayed the largest zone of inhibition in the acetone leaf extract of *Tinospora sinensis*, followed by *Staphylococcus aureus* (20 mm), *Pseudomonas fluorescens* (16 mm), and *Streptococcus mutans* (14 mm). The stem extract exhibited the largest zone of inhibition in *Pseudomonas fluorescens* (20 mm), *Streptococcus mutans* (17 mm), *Bacillus subtilis* (16 mm), and *Staphylococcus aureus* (22 mm).

The maximum zone of inhibition in *Staphylococcus aureus* (24 mm) was observed in an aqueous leaf extract of *Tinospora sinensis*. This was followed by *Bacillus subtilis* (23 mm), *Pseudomonas fluorescens* (21 mm), and *Streptococcus mutans* (19 mm). *Pseudomonas fluorescens*, *Streptococcus mutans*, *Bacillus subtilis*, and *Staphylococcus aureus* all had lower zones of inhibition in the stem extract (18 mm), compared to 23 mm for *Bacillus subtilis*.

4. Discussion

Many medicinal plants have a rich history of traditional use, often supported by anecdotal evidence of their efficacy. Phytochemical analysis bridges this gap between traditional knowledge and scientific validation, providing a rigorous framework for understanding how these plant materials can be transformed into effective treatments. Additionally, this research contributes to the conservation of biodiversity, as it highlights the importance of preserving plant species that hold therapeutic potential. Ultimately, by fostering an appreciation for the medicinal properties of plants through phytochemical analysis, we can enrich modern medicine and promote a sustainable approach to health care that respects and utilizes the natural world. The results confirm the presence of constituents which are known to exhibit medicinal as well as physiological activity (2). The findings closely align with the results from the research conducted by Kushwah et al (5). In 2015, researchers analyzed both the quantitative and qualitative aspects of ashwagandha root powder. They have also identified the existence of heavy metals and inorganic substances within the root powder. Velu and Baskaran (2012) delved into the analysis of the antimicrobial properties and phytochemical screening of *Withania somnifera* in various solvents such as ethanol, methanol, ethyl acetate,

acetone, chloroform, petroleum ether, acetone, and hot water (6).

Antimicrobial activity is a fundamental aspect of microbiology and health sciences, focusing on the ability of substances to inhibit the growth of or kill microorganisms, including bacteria, fungi, viruses, and parasites. Understanding antimicrobial activity is crucial, particularly in the context of increasing antibiotic resistance, which poses significant challenges to public health worldwide. Antimicrobials can be derived from various sources, including natural products like plant extracts and synthetic compounds developed through rigorous scientific processes. When evaluating antimicrobial activity, researchers typically assess factors such as the concentration of the antimicrobial agent, the type of microorganism involved, and the environmental conditions under which the interaction occurs. Effective antimicrobial agents show potency against pathogens without adversely affecting human cells, which is a critical consideration in drug development. Furthermore, studying the mechanisms by which these agents exert their effects helps in enhancing their efficacy and discovering new drug targets, ultimately contributing to better therapeutic strategies in combating infectious diseases.

In the realm of healthcare, the discussion around antimicrobial activity extends beyond the laboratory into clinical practice, where the appropriate use of these agents is paramount. Misuse and overuse of antimicrobial medications have led to the emergence of resistant strains of pathogens, complicating treatment protocols and posing risks of prolonged illness or increased mortality. As a result, healthcare professionals emphasize the importance of stewardship programs aimed at optimizing the efficacy of antimicrobial therapies while minimizing the potential for resistance development. This involves careful selection of antimicrobial agents based on susceptibility testing, as well as educating patients about the importance of adhering to prescribed treatments. Additionally, researchers continuously explore alternative therapies and combinations of existing medications to enhance antimicrobial effectiveness. The ongoing investigation into naturally occurring compounds, such as those found in certain foods and herbal remedies, provides exciting prospects for discovering novel antimicrobial agents that may function synergistically with conventional therapies. Through coordinated efforts between laboratories and healthcare settings, significant strides can be made in understanding and improving antimicrobial activity.

The exploration of antimicrobial activity in medicinal plants has garnered significant attention in recent years due to the growing concern over antibiotic resistance. Various studies have shown that numerous plant extracts possess bioactive compounds with the ability to inhibit the growth of pathogenic microorganisms. For instance, plants such as Neem (*Azadirachta indica*) and Turmeric (*Curcuma longa*) have been noted for their antibacterial and antifungal properties. The active ingredients in these plants, including azadirachtin in Neem and curcumin in Turmeric, exhibit mechanisms that disrupt microbial cell walls and metabolic pathways, thus preventing infection. Furthermore, the utilization of these plants can be particularly beneficial in regions where access to pharmaceutical antibiotics is limited, providing an alternative means of addressing infections that pose a threat to public health (7). The incorporation of traditional knowledge surrounding these medicinal plants into modern research may lead to the development of novel antimicrobial agents that could complement existing therapeutic strategies and help counteract the alarming rise in antibiotic-resistant strains.

Research has increasingly focused on the bioactive components of various medicinal plants, which demonstrate promising antimicrobial properties against a broad spectrum of pathogens. For example, extracts from plants such as Oregano (*Origanum vulgare*) and Garlic (*Allium sativum*) have shown activity against both gram-positive and gram-negative bacteria. The primary active ingredient in Oregano, carvacrol, exhibits potent antimicrobial effects through its ability to compromise bacterial membrane integrity. Similarly, Allicin, found in Garlic, has been demonstrated to possess significant antibacterial, antifungal, and antiviral properties. These findings suggest that traditional uses of such plants may hold scientific merit in the development of new antimicrobial therapies. Moreover, the low toxicity profile of many herbal remedies compared to synthetic antibiotics underscores the potential of integrating these natural alternatives into modern medicine. This integration not only enhances the therapeutic options available for treating infections but also aligns with the trend toward more sustainable and environmentally friendly healthcare practices, which increasingly recognize the undeniable value of our natural resources in combating illness (8).

The use of medicinal plants as natural sources of antimicrobial agents has been recognized across various cultures and traditional medicine systems for centuries. Recent scientific investigations have corroborated these age-old practices, revealing that many plants contain phytochemicals effective against harmful microorganisms. For instance, the essential oils derived from plants like Tea Tree (*Melaleuca alternifolia*) and Thyme (*Thymus vulgaris*) exhibit strong antimicrobial activity due to their high concentrations of terpenes and phenolic compounds. These substances can disrupt microbial enzymes and essential metabolic processes, contributing to their efficacy against bacteria and fungi. Moreover, the synergistic effects observed in some plant mixtures highlight the potential for developing multifaceted antimicrobial formulations derived from nature. Comparative studies that evaluate these plant extracts alongside conventional antibiotics also emphasize the other benefits of using medicinal plants, including fewer side effects and the potential to enhance the effect of existing antibiotics through adjuvant therapy (9). Therefore, the integration of traditional knowledge along with scientific validation can pave the way for innovative approaches in addressing the global threat posed by microbial resistance.

The increasing prevalence of antibiotic-resistant infections necessitates the exploration of alternative treatments, such as those derived from medicinal plants with established antimicrobial properties. Many studies have identified a wide range of plants that exhibit significant inhibitory effects against pathogens of clinical importance. For example, extracts from Ginger (*Zingiber officinale*) and Eucalyptus (*Eucalyptus globulus*) have displayed promising antimicrobial characteristics, attributed to their active compounds such as gingerol and eucalyptus oil. Research indicates that these compounds can interfere with the growth and survival of various bacteria and fungi, offering a potential foothold against infections that are resistant to conventional antibiotics. Additionally, the ability of these plants to enhance the immune response illustrates their dual role in both preventing infection and supporting overall health. The shift toward natural products in pharmaceutical applications is spurred by the urgent need for novel antimicrobial agents, while also reducing the reliance on synthetic options which can be financially burdensome and present various health risks (10). The collaboration between ethnobotanists and microbiologists promises to enrich the field of antimicrobial drug discovery, combining modern techniques with traditional plant wisdom.

The search for effective antimicrobial agents from medicinal plants is a promising avenue in the quest to combat multidrug-resistant pathogens. Numerous studies have highlighted the antimicrobial properties of a variety of plants, showcasing their potential in treating infections where conventional antibiotics fail. For instance, research has shown that extracts from plants such as Basil (*Ocimum basilicum*) and Echinacea (*Echinacea purpurea*) possess strong antimicrobial activities attributed to their rich content of flavonoids and essential oils. These components not only inhibit microbial growth but also can modulate the immune system, enhancing the body's ability to fight infections effectively. There has been a growing body of evidence supporting the synergistic effects of different plant extracts, indicating that combining various medicinal plants could lead to more potent antimicrobial solutions. Importantly, the low incidence of adverse effects associated with these natural remedies makes them attractive alternatives to synthetic antibiotics. The ongoing research into the pharmacological potential of these plants is essential as it may lead to innovative treatments that could bolster existing healthcare systems and provide safer, sustainable solutions to combat microbial resistance (11).

Researchers are now diving into the world of ethnobotany to find plants with promising antimicrobial qualities. By studying these plants, scientists aim to unlock their potential and possibly develop new treatments that can support or replace conventional antibiotics.

In recent years, the interest in the antimicrobial effects of medicinal plants has seen a notable resurgence, largely driven by the need to find alternatives to synthetic drugs. Traditional knowledge, passed down through generations, has played a significant role in this exploration. Many cultures around the world have long used specific plants for their healing properties, often citing their effectiveness in treating infections. For instance, plants like garlic, ginger, and various herbs have been investigated for their potential antibacterial and antifungal activities. Research has revealed that these plants not only help fight infections but can also enhance the immune system, providing a holistic approach to health. Additionally, scientists are exploring ways to isolate and purify the active compounds in these plants to formulate effective remedies. This exploration is not without its challenges, as there is a need to ensure safety and efficacy through rigorous testing. However, the promise of harnessing nature's bounty to create new antimicrobial agents opens up exciting avenues for healthcare, especially in an era where drug resistance poses a threat to effective treatment.

The activity of the organic extracts could also be attributed to the presence of higher terpenoids containing carboxylic acid groups. Numerous workers have documented the analgesic qualities of alkaloids (12,13) and the anti-inflammatory as well as anti-bacterial traits of tannins (14). These compound classes are recognized for their curative effects on various bacteria. It is no wonder that herbalists traditionally utilize these plant extracts to treat illnesses caused by bacteria. Tannins, along with their protein content are present in 232 Afr. J. Pure Appl. Chem.

The combined effects of precipitation and vasoconstriction present potential benefits in averting the onset of ulcers (15,16). The diuretic and antibacterial properties of plant extracts rich in flavonoids have been well-documented by various researchers (2, 17-19). The alkaloids found in plants are utilized in medicine as anesthetic agents (20). Saponins found in plants have been identified as the cause of the invigorating and tonic effects seen in Chinese and

Japanese medicinal herbs, as reported by Alinnor in 2008 (21). The findings of this study suggest that the phytochemical compounds identified may be the bioactive constituents responsible for the efficacy of the leaves of the plants studied. The confirmed presence of certain compounds has been linked to antimicrobial activity (22). Therefore, it is plausible to infer that plant extracts could serve as a valuable resource for the production of pharmaceuticals employed in combating various microbial infections (23).

5. Conclusion

Researchers typically create detailed reports outlining the methodology, results, and potential implications of their studies. If the plant extract demonstrates significant antimicrobial activity, further studies may be warranted to identify the specific compounds responsible for this effect. This often involves isolating and characterizing the active substances using various techniques such as chromatography or mass spectrometry. Additionally, researchers may explore the mechanisms of action to understand how the plant extract interacts with the microbial cells. Such findings could lead to the development of new antimicrobial agents derived from natural sources, addressing the urgent need for alternative treatments in the face of rising antibiotic resistance. Overall, the antimicrobial activity procedure for plant extracts not only highlights the therapeutic potential of nature but also contributes to the broader field of pharmacognosy.

Understanding the chemical composition of a plant allows researchers to evaluate its medicinal properties, helping to validate traditional uses in herbal medicine. The integration of traditional knowledge with modern analytical techniques paves the way for innovative approaches to harness the vast potential of plants in health and nutrition. Ultimately, thorough phytochemical analysis not only advances scientific understanding but also supports the sustainable use of plant resources, fostering a deeper respect for biodiversity and natural remedies.

References

1. Harborne JB (1973). *Phytochemical Methods*. Chapman and Hall Ltd., London pp. 49-188.
2. Sofowora A (1993). *Medicinal Plants and Traditional Medicine in Africa*. Spectrum Books Ltd., Ibadan, Nigeria, pp. 191-289.
3. Saiful Yazan, L.; Armania, N. *Dillenia species: A review of the traditional uses, active constituents and pharmacological properties from pre-clinical studies*. *Pharm. Biol.* 2014, 52, 890– 897, DOI: 10.3109/13880209.2013.872672.
4. Seth, S. D.; Sharma, B. *Medicinal plants in India*. *Indian J. Med. Res.* 2004, 120, 9– 11.
5. Kushwaha S, Betsy A, Chawla P. *Effect of Ashwagandha (Withania somnifera) Root Powder Supplementation in Treatment of Hypertension*. *Journal of Ethnobiology and Ethnomedicine*. 2012; 6(2):111-115
6. Velu S, Baskaran C. *Phytochemical analysis and in-vitro antimicrobial activity of Withania somnifera (Ashwagandha)*. *Journal of Natural Product and Plant Resources*. 2012; 2(6):711-716
7. Ncube B, Finnie JF, Van Staden J. *Quality from the field: The impact of environmental factors as quality determinants in medicinal plants*. *South African Journal of Botany*. 2012 Sep 1;82:11-20.
8. Sharma G, Sharma S, Kumar A, Ala'a H, Naushad M, Ghfar AA, Mola GT, Stadler FJ. *Guar gum and its composites as potential materials for diverse applications: A review*. *Carbohydrate polymers*. 2018 Nov 1;199:534-45.
9. Bakkali F, Averbeck S, Averbeck D, Idaomar M. *Biological effects of essential oils—a review*. *Food Nanotechnology Perceptions* Vol. 21 No. S1 (2025)

- and chemical toxicology. 2008 Feb 1;46(2):446-75.
10. Mishra S, Mindermann S, Sharma M, Whittaker C, Mellan TA, Wilton T, Klapsa D, Mate R, Fritzsche M, Zambon M, Ahuja J. Changing composition of SARS-CoV-2 lineages and rise of Delta variant in England. *EClinicalMedicine*. 2021 Sep 1;39.
 11. Cowan MM. Plant products as antimicrobial agents. *Clinical microbiology reviews*. 1999 Oct 1;12(4):564-82.
 12. Antherden LM (1969). *Textbook of Pharmaceutical Chemistry*, 8th edn. Oxford University Press, London pp. 813-814.
 13. Duguid JP (1989). A guide to the laboratory diagnosis and control of infection. In Collee et al. (eds) MacKie and McCartney *Medical Microbiology*, 13th edn., Vol. 1, Churchill Livingstone, London, p. 163
 14. Aguwa CN, Nwankwo SO (1988). Preliminary studies on the root extract of *Nauclea latifolia* S. for antiulcer properties. *Nig. J. Pharmaceutical Sci.* 4(1): 16-23.
 15. Dahiru D, Onubiyi JA, Umaru HA (2006). Phytochemical screening and antiulcerogenic effect of *Moringa oleifera* aqueous leaf extract. *Afr. J. Trad. Comp. Alt. Med.* 3(3): 70-75
 16. Enwerem NM, Okogun JI, Wambebe CO, Ajoku GA, Okorie DA (2003). Antibacterial principle from the stem bark of *Berlina grandiflora*. *J. Chem. Soc. Niger.* 28(1): 52-54.
 17. Enwerem NM, Wambebe CO, Okogun JI, Akah PA, Gamaniel KS (2001). Anthelmintic screening of the stem bark of *Berlina grandiflora*. *J. Natural Remedies*, 1: 17-23.
 18. Monache GD, Botta B, Vinciguerra V, de Mello J, Chiappeta A (1996). Antimicrobial isoflavanones from *Desmodium canum*. *Phytochemistry*, 41(2): 537-544.
 19. Rao CP, Prashant A, Krupadanam GLD (1996). Two prenylated isoflavans from *Milletia racemosa*, *Phytochemistry*, 41(4): 1223-1224.
 20. Herourat D, Sangwin RS, Finiaux MA, Sangwan-Norrell BS (1988). Variations in the leaf alkaloid content of androgenic diploid plants of *Datura innoxia*, *Planta medical. J. Med. Plant Res.* 54: 14-20.
 21. Alinnor IJ (2008). Preliminary phytochemical and antibacterial activity screening of leaves of *Vernonia amygdalina*, *J. Chem. Soc. Nigeria*, 33(1): 172-177.
 22. Odebiyi A, Sofowora AE (1978). Phytochemical screening of Nigerian medicinal plants. *Lloydia* 41(3): 234-246.
 23. Kubmarawa D, Wase GA, Ayinla OG (2007). Preliminary studies on phytochemical analysis and antimicrobial evaluation of extracts of *Commiphora kerstingii*, *J. Chem. Soc. Nigeria*, 32(1): 38-40.