

Photobiogenic Mediated Silver Nanoparticles Synthesis and Its Antimicrobial Activity

Durgadevi S¹, Divya Tejaswini D¹, Gracy Sattu², Ancy Jenifer A³,
Rajendran.G^{3*}

¹M.Sc., Students, Department of Biotechnology, Faculty of Science and Humanities, SRM Institute of Science and Technology, Ramapuram, Chennai-600089, Tamil Nadu, India.

²Students, Department of Biotechnology, Faculty of Science and Humanities, SRM Institute of Science and Technology, Ramapuram, Chennai-600089, Tamil Nadu, India.

³Department of Biotechnology, Faculty of Science and Humanities, SRM Institute of Science and Technology, Ramapuram, Chennai-600089, Tamil Nadu, India.

Silver nanoparticles were synthesized by photobiogenic method using *Euphorbia hirta* aqueous extract under the bright sunlight in 25 min at pH 7.5. *Euphorbia hirta* extract acts as a capping agent of silver nanoparticle synthesis. The synthesized Eh-AgNPs were characterized by UV-vis spectroscopy, HR-TEM, and zeta potential. The SPR peak of Eh-AgNPs was produced at 424nm. The Eh-AgNPs were found to be spherical shape with average size of 12nm. The synthesized Eh-AgNPs were highly stable in the liquid medium. The antibacterial activity of different concentration of Eh-AgNPs against both Gram positive and Gram negative pathogenic bacteria were analyzed by the well diffusion assay method. Eh-AgNPs inhibited the growth of both groups of bacteria in the concentration manner. The maximum zone of inhibition 15mm, 22mm, 19mm and 20mm were obtained at 100µg/ml concentration for *Enterococcus faecalis*, *Staphylococcus aureus*, *Escherichia coli*, *Pseudomonas aeruginosa* respectively. The synthesized Eh-AgNPs have well antibacterial activity against both groups of bacteria particularly it has maximum activity against pathogenic bacteria such as *Pseudomonas aeruginosa*, and *Staphylococcus aureus*.

Keywords: Silver nanoparticles, *Euphorbia hirta*, Sunlight irradiation, Green synthesis, Physicochemical characterization, Antibacterial activity.

1. Introduction

Nanoparticles have special attraction due to its size, shape, adaptable surface modification,

bioconjugation, optical, electrical and thermal properties¹. Among the different metal nanoparticles, silver nanoparticles have huge potential due to having high surface area to volume ratio and it has vast applications from antimicrobial to photo catalyst^{2,3}. The silver nanoparticles can be synthesized by physical, chemical and biological methods. The physical and chemical method of nanoparticles synthesis is required high energy, pressure, sophisticated instruments, and involves toxic chemicals. Biological synthesis of nanoparticles has gained attention compare to physical and chemical methods. The bioinspired syntheses of nanoparticles using plants are the non toxic, easily available, low cost, environmentally friendly methods⁴. Previously, several research works have been published on biogenic synthesis of metal nanoparticles using plant extract. Recently, Shereen et al., 2024 reported that plant extract preparation and green synthesis of silver nanoparticles using *Swertia chirata*⁵. Ejaz et al., 2024 explained the characterization, synthesis, and biological activities of silver nanoparticles produced via green synthesis method using *Thymus vulgaris* aqueous extract⁶. Silver ions and silver-based compounds are highly toxic to microorganisms including 16 major species of bacteria. This makes silver an excellent choice for multiple roles in the medical field. Silver is generally used in the nitrate form to induce antimicrobial effect, and when nanoparticles are used, there is a huge increase in the surface area to be in contact with microbial cells⁷.

The present research work describes the biogenic synthesis of silver nanoparticles using *Euphorbia hirta* aqueous extract under the sunlight in biological pH. *Euphorbia hirta* is belongs to the family of Euphorbiaceae, which is traditionally using medicinal plant for treatment of various disorders and diseases⁸. *Euphorbia hirta* has several potential pharmacological activities including antioxidant, anti-inflammatory and anticancer⁹, wound healing potential¹⁰, anthelmintic, antidipsogenic, antiarthritis¹¹, antibacterial¹², antianaphylactic¹³, anxiolytic¹⁴, antidiabetic¹⁵ and hepatoprotective¹⁶. The synthesized silver nanoparticles were characterized by UV-vis spectroscopy, HR-TEM, zeta potential. Further, Antibacterial activity of the synthesized Eh-AgNPs was analyzed using the well diffusion assay method. The antibacterial activity of different concentration of Eh-AgNPs was analyzed against both Gram positive and Gram negative bacteria. Eh-AgNPs inhibited the growth of bacteria in the concentration manner. Particularly, the synthesized Eh-AgNPs has excellent antibacterial activity against pathogenic bacteria such as *Pseudomonas aeruginosa*, *Staphylococcus aureus*. Previously several authors explained the antimicrobial activity of silver nanoparticles synthesized by plants extract. Crisan et al., 2024 reported that In vitro antimicrobial activity of silver nanoparticles against selected Gram-negative and Gram-positive pathogens¹⁷. Holubnycha et al., 2024 explained that the antimicrobial activity of two different types of silver nanoparticles against wide range of pathogenic bacteria¹⁸.

2. Materials and Method

Materials

Silver nitrate was obtained from Sigma Aldrich, USA. Muller Hinton Agar medium was purchased from Himedia, India. Sodium hydroxide was obtained from SRL India.

Preparation of Euphorbia hirta Aqueous Extract

Euphorbia hirta was collected from the herbal garden, University of Madras, Chennai. 35% of aqueous extract of Euphorbia hirta was prepared using sterile distilled water, clean mortar and pestle. The extract was filtered with whatman No:1 filter paper and stored at 4°C for silver nanoparticles synthesis.

Synthesis of Silver Nanoparticles

50ml of Silver nitrate solution (1mM) was taken in a clean conical flask, in which 2.5ml of Euphorbia hirta aqueous extract was added and pH of the solution was adjusted to 7.5 using sodium hydroxide solution. The solution mixture was stirred in magnetic stirrer and kept under the bright sun light; the colorless Silver nitrate solution was changed to yellow colour in 25 minutes that indicates the synthesis of Silver nanoparticles. It is stored at 4°C for further characterization and applications studies.

Characterization of Eh-AgNPs

UV-visible Spectrophotometer Analysis

The synthesized Eh-AgNPs 3ml was taken in a clean quartz cuvette, the surface plasmon resonance (SPR) peak of Eh-AgNPs was analyzed using Shimadzu UV 1601 spectrometer, Japan and the SPR peak reading was recorded and graph was plotted using origin software.

High Resolution - Transmission Electron Microscopy (HR-TEM) Analysis

The size and shape of Eh-AgNPs was analyzed using HR-TEM (FEI TECHNAI G2 MODEL T-30-S-TWIN, USA). 5µl of Eh-AgNPs was placed in a copper grid and allowed it to dry in room temperature. Then the grid was examined in HR-TEM operated at 200kV to measure the size of the synthesized nanoparticles.

Zeta Potential Analysis

The net charge of synthesized Eh-AgNPs was estimated using Nano Series (Malvern Instruments, United Kingdom). Eh-AgNPs solution was taken in an electrode cuvette without air bubbles and placed in the sample holder and the instrument was run at 100VA at 25°C and 232 kcps with the duration of 60 seconds.

In-vitro Hemolytic Assay

The hemolytic activity of silver nanoparticles was analyzed using human red blood cells by In-vitro hemolytic assay¹⁹. The blood sample was collected and appropriate anticoagulant was added into it. The red cells suspension (5%) was prepared from the collected blood sample using saline. The different concentration (20, 40, 60, 80 and 100µg/ml) of silver nanoparticles were allowed into the RBCs suspension and incubate at 37°C for 1h. Then, the samples were centrifuged and the supernatant was used for the hemolysis estimation at 570nm in UV-visible spectroscopy.

In-vitro Antibacterial Activity

Antibacterial activity of synthesized Eh-AgNPs was analyzed by Muller Hinton Agar well diffusion method²⁰ against Gram positive (Enterococcus faecalis, Staphylococcus aureus) and Gram negative (Escherichia coli, Pseudomonas aeruginosa) pathogenic bacteria. The bacteria

were grown in Nutrient broth for 24 hours. Muller Hinton Agar (MHA) (3.8gms/100ml) was prepared by the sterile distilled water and transferred to sterile petriplates. About 1.5×10^6 CFU/mL suspensions of each test bacteria were inoculated in surface of MHA by sterile swab. Then the following concentration of Eh-AgNPs (40, 60, 80 and 100 μ g/mL) was added into the well of agar plates. Plates were incubated for 24h at 37°C. The bactericidal activity of Eh-AuNPs was detected by measuring the zone of inhibition in millimeter (mm). Streptomycin and Ampicillin (30 μ g) were used as a control for Gram positive and Gram negative bacteria respectively.

Statistical Analysis

All the data were evaluated using the statistical software SPSS/16. Hypothesis-testing methods included one-way analysis of variance (ANOVA) followed by least significant difference test. P-values less than 0.05 were considered statistically significant. All the results were expressed as mean \pm standard deviation (n = 3).

3. Results and Discussion

Synthesis of Eh-AgNPs

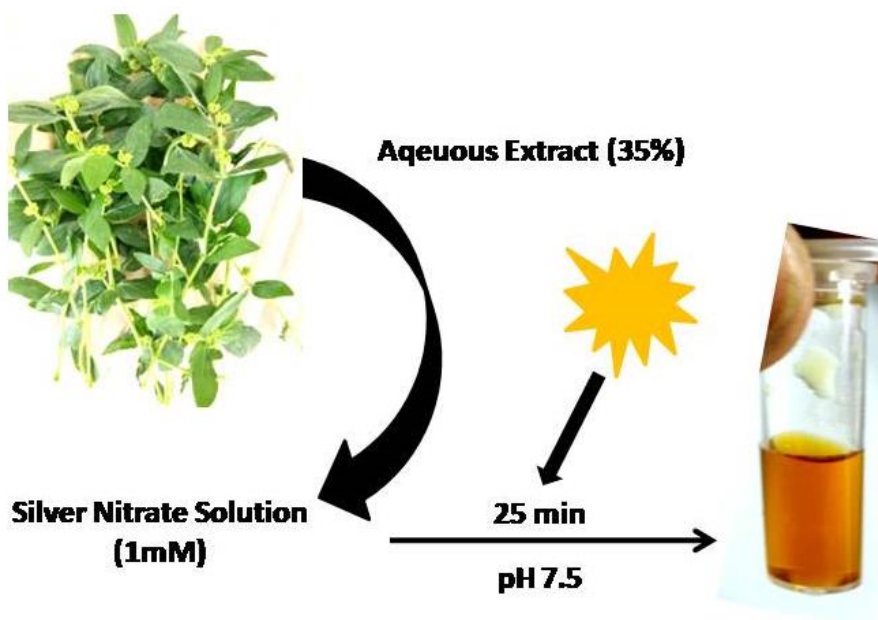


Figure 1: Schematic representation of synthesis of Eh-AgNPs by photobiogenic method under the bright sunlight irradiation.

The present work describes the *Euphorbia hirta* aqueous extract capped biogenic synthesis of silver nanoparticles under the bright sunlight. The colourless silver nitrate solution was changed to yellow colour after adding the *Euphorbia hirta* aqueous extract in 25 minutes at pH:7.5 (Figure 1). *Euphorbia hirta* aqueous extract acts as a capping agent and the sunlight act as a catalyst in the silver nanoparticles synthesis. This biogenic and sunlight mediated synthesis

of silver nanoparticles is a cost effective, facile, and non toxic eco-friendly method. The physical and chemical methods are costly, require sophisticated instruments, toxic chemical were used for nanoparticles synthesis^{21, 22}.

Physico-chemical Characterization of Eh-AgNPs

UV-Visible Spectroscopy

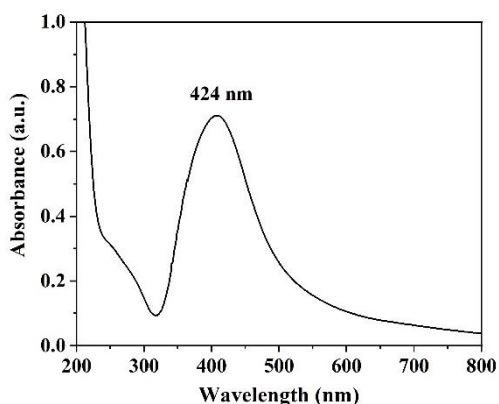


Figure 2: UV-visible spectroscopy analysis of Eh-AgNPs

The UV-visible spectroscopy is an early characterization technique to validate the nanoparticles synthesis. Generally, The AgNPs are showed SPR peak between 400 to 450nm²³. The synthesized Eh-AgNPs showed narrow, single SPR peak at 424nm that confirmed the synthesis of silver nanoparticles (Figure 2). The obtained SPR peak exhibited in narrow, single peak that means the particles are in small size and homogenous. The morphology of metal nanoparticles are depending on the extract concentration, composition, pH, temperature and reaction time^{24,25}.

Zeta Potential Analysis

The surface net charge of Eh-AgNPs was estimated by zeta potential and it was found to be -20.7mV (Figure 3). The zeta potential is determined from the electrophoretic mobility of silver nanoparticles by its degree of repulsion between adjacent, similarities charged particles ²⁶. This strong negative charge of Eh-AgNPs is prevented the aggregation of particles, and maintain the stability in the liquid medium for long duration²⁷.

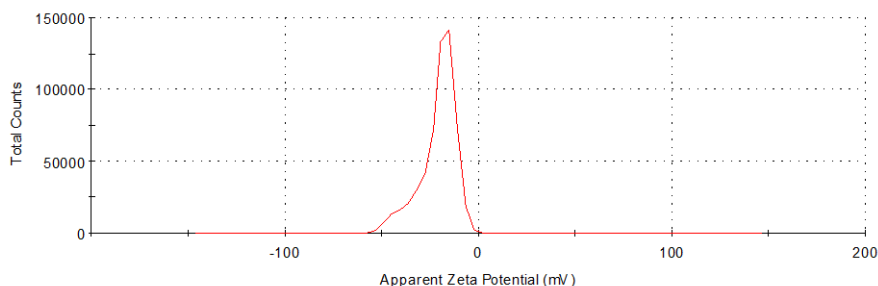


Figure 3: Zeta potential of Eh-AgNPs was found to be -20.7mV.

HR-TEM Analysis of Eh-AgNPs

The size and shape of Eh-AgNPs was analyzed through the HR-TEM. The Eh-AgNPs were spherical in shape with monodispersity. The particle size was between 9 to 21nm, and the average particles size was found to be 12nm (Figure 4). The detection of morphology of nanoparticles is very important in biological applications. A small size nanomaterials is easily penetrates the cell membrane and it can be used for various biomedical applications.

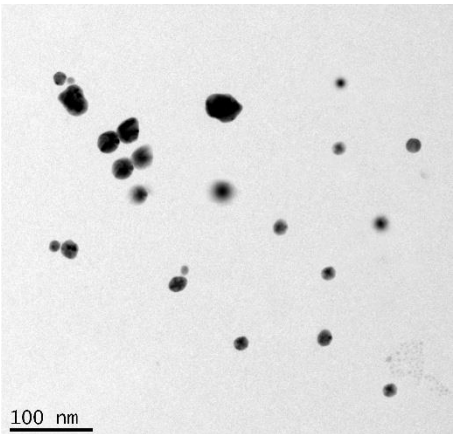


Figure 4: The morphology of Eh-AgNPs were analyzed by HR-TEM.

Hemolytic assay

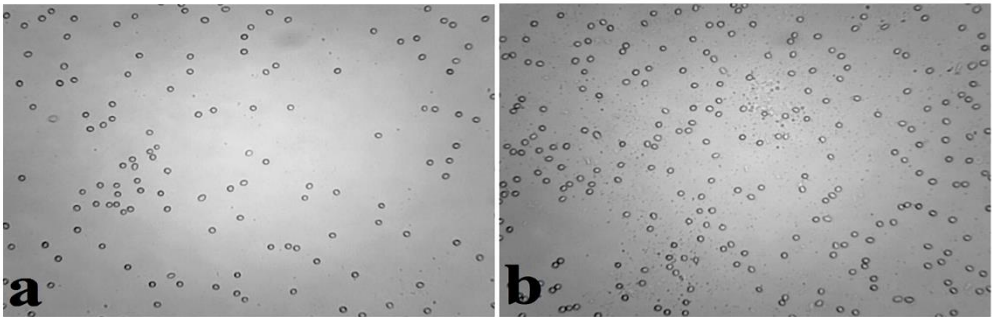
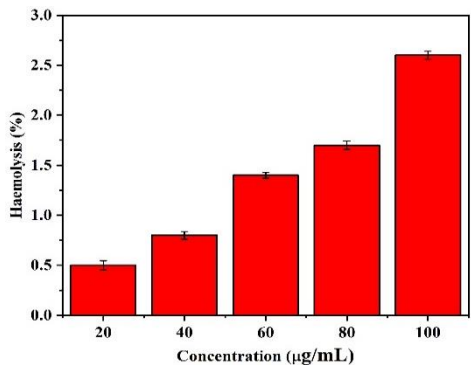


Figure 5: The hemolytic assay of Eh-AgNPs and its corresponding morphology of RBCs.

The hemocompatibility of Eh-AuNPs was analyzed in human RBCs. The different concentration of Eh-AuNPs was incubated with human RBCs and hemolysis (%) was estimated by UV-visible spectroscopy (Figure 5). About 0.5%, 0.8%, 1.45%, 1.7%, and 2.6% of hemolysis was observed for 20, 40, 60 80 and 100 μ g/mL concentration of Eh-AuNPs. The hemolysis (%) was depended on the concentration of the Eh-AuNPs, but the maximum tested concentration of Eh-AuNPs (100 μ g/mL) induced normal acceptable level of hemolysis (<5%). Therefore, Eh-AuNPs is having fine hemocompatibility property upto 100 μ g/mL. According to the ASTM (American standard for testing materials) upto 5% of hemolysis is considered as hemocompatible. Previously Muthukumarasamyvel et al., 2017 reported hemocompatibility of cholic acid capped AuNPs (DCaC- AuNPs, DCaDC- AuNPs, DCaLC- AuNPs) in human RBCs. AuNPs treated human blood sample was produced 1.90, 1.97, and 2.09%, hemolysis in the respective order at 120 μ g/mL concentration²⁸. Ramadurai et al.,2010 explained that the CuNCs treated RBCs, 0.5% of haemolysis was observed for a minimum concentration of CuNCs (4 μ g/mL), 4.7% of haemolysis was observed for 16 μ g/mL and 7.5% of haemolysis was obtained for maximum concentration of 22 μ g/mL respectively²⁹.

The hemocompatibility of Eh-AuNPs was further confirmed through the morphological analysis of RBCs. The normal RBCs is oval or biconcave shape³⁰. There was no significant morphological change observed in Eh-AuNPs treated RBCs. Eh-AuNPs treated RBCs exhibited normal morphology like untreated control RBCs morphology and RBCs was appeared in disperse manner without any aggregation (Figure 5).

In-vitro Antibacterial Activity

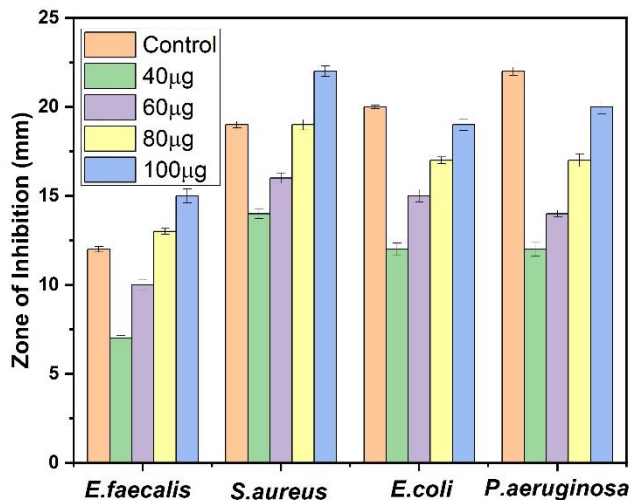


Figure 6: Antibacterial activity of Eh-AgNPs pathogenic bacteria (Zone of inhibition in millimeter)

Antibacterial activity of the Eh-AgNPs was tested using a well diffusion assay method. The antibacterial activity of different concentration of Eh-AgNPs against both Gram positive and Gram negative bacteria are shown in the figure 6. Eh-AgNPs inhibited the growth of bacteria in the concentration manner. The maximum zone of inhibition 15mm, 22mm, 19mm and 20mm were obtained at 100 μ g/ml concentration for Enterococcus faecalis, Staphylococcus aureus,

Escherichia coli, *Pseudomonas aeruginosa* respectively. The synthesized Eh-AgNPs has excellent antibacterial activity against pathogenic bacteria such as *Pseudomonas aeruginosa*, *Staphylococcus aureus*. AgNPs will lead to high antimicrobial activity as compared with bulk silver metal³¹. The mode of action of AgNPs against bacteria is not completely understood yet. However, several hypotheses are explaining the antibacterial activity of silver nanoparticle by generation of reactive oxygen species, release of Ag⁺ ions from AgNPs denaturize proteins by bonding with sulfhydryl groups and attachment of AgNPs on bacteria and subsequent damage to bacteria^{32,33,34}.

4. Conclusion

The AgNPs was synthesized in a cost effective, eco-friendly, photobiogenic method using *Euphorbia hirta* aqueous extract under the bright sunlight irradiation. The synthesized Eh-AgNPs was exhibited in spherical shape with average size of 12nm. Eh-AgNPs has high stability with strong negative surface net charge (-20.7mV). Eh-AgNPs inhibited the growth of both Gram positive and negative pathogenic bacteria in the concentration manner. However, Eh-AgNPs has excellent antibacterial activity against pathogenic bacteria such as *Pseudomonas aeruginosa* and *Staphylococcus aureus*. Hence, this synthesized Eh-AgNPs can be used in pharmaceutical and biomedical industries for preparation of bactericidal cream for treating these above listed microorganisms.

Acknowledgement

The authors would like to acknowledge the GNR Instrumentation Centre, University of Madras, Guindy Campus, Chennai, Tamil Nadu for providing analytical facilities.

References

1. Petros, R. A., & DeSimone, J. M. (2010). Strategies in the design of nanoparticles for therapeutic applications. *Nature reviews Drug discovery*, 9(8), 615-627
2. Ahmed, S., Ahmad, M., Swami, B. L., & Ikram, S. (2016). A review on plants extract mediated synthesis of silver nanoparticles for antimicrobial applications: a green expertise. *Journal of advanced research*, 7(1), 17-28.
3. Shankar, T., Karthiga, P., Swarnalatha, K., & Rajkumar, K. (2017). Green synthesis of silver nanoparticles using *Capsicum frutescens* and its intensified activity against *E. coli*. *Resource-Efficient Technologies*, 3(3), 303-308.
4. Renuka, R., Devi, K. R., Sivakami, M., Thilagavathi, T., Uthrakumar, R., & Kaviyarasu, K. (2020). Biosynthesis of silver nanoparticles using *Phyllanthus emblica* fruit extract for antimicrobial application. *Biocatalysis and Agricultural Biotechnology*, 24, 101567.
5. Shereen, M. A., Ahmad, A., Khan, H., Satti, S. M., Kazmi, A., Bashir, N., ... & Zouidi, F. (2024). Plant extract preparation and green synthesis of silver nanoparticles using *Swertia chirata*: Characterization and antimicrobial activity against selected human pathogens. *Heliyon*, 10(6).
6. Ejaz, U., Afzal, M., Mazhar, M., Riaz, M., Ahmed, N., Rizg, W. Y., ... & Yean, C. Y. (2024). Characterization, synthesis, and biological activities of silver nanoparticles produced via green synthesis method using *Thymus vulgaris* aqueous extract. *International Journal of Nanomedicine*, 453-469.
7. Salem, W. M., Haridy, M., Sayed, W. F., & Hassan, N. H. (2014). Antibacterial activity of silver nanoparticles synthesized from latex and leaf extract of *Ficus sycomorus*. *Industrial Crops and*

- products, 62, 228-234.
8. Ali, M. Z., Mehmood, M. H., Saleem, M., & Gilani, A. H. (2020). The use of *Euphorbia hirta* L.(Euphorbiaceae) in diarrhea and constipation involves calcium antagonism and cholinergic mechanisms. *BMC complementary medicine and therapies*, 20, 1-16.
 9. Sharma, N., Samarakoon, K. W., Gyawali, R., Park, Y. H., Lee, S. J., Oh, S. J., ... & Jeong, D. K. (2014). Evaluation of the antioxidant, anti-inflammatory, and anticancer activities of *Euphorbia hirta* ethanolic extract. *Molecules*, 19(9), 14567-14581.
 10. Tuhin, R. H., Begum, M. M., Rahman, M. S., Karim, R., Begum, T., Ahmed, S. U., ... & Begum, R. (2017). Wound healing effect of *Euphorbia hirta* linn.(Euphorbiaceae) in alloxan induced diabetic rats. *BMC complementary and alternative medicine*, 17, 1-14.
 11. Al-Snafi, A. E. (2017). Pharmacology and therapeutic potential of *Euphorbia hirta* (Syn: *Euphorbia pilulifera*)-A review. *IOSR Journal of Pharmacy*, 7(3), 7-20.
 12. Kader, J., Noor, H. M., Radzi, S. M., & Wahab, N. A. A. (2013). Antibacterial activities and phytochemical screening of the acetone extract from *Euphorbia hirta*. *International Journal of Medicinal Plant Research*, 2(4), 209-214.
 13. Youssouf, M. S., Kaiser, P., Tahir, M., Singh, G. D., Singh, S., Sharma, V. K., ... & Johri, R. K. (2007). Anti-anaphylactic effect of *Euphorbia hirta*. *Fitoterapia*, 78(7-8), 535-539.
 14. Xia, M., Liu, L., Qiu, R., Li, M., Huang, W., Ren, G., & Zhang, J. (2018). Anti-inflammatory and anxiolytic activities of *Euphorbia hirta* extract in neonatal asthmatic rats. *AMB Express*, 8, 1-11.
 15. Subramanian, S. P., Bhuvaneshwari, S., & Prasath, G. S. (2011). Antidiabetic and antioxidant potentials of *Euphorbia hirta* leaves extract studied in streptozotocin-induced experimental diabetes in rats. *General physiology and biophysics*, 30(3), 278-285.
 16. Tiwari, N., Mishra, A., Bhatt, G., & Chaudhary, A. (2015). Anti-stress activity of A bioflavanoid: Quercetin from *euphorbia hirta*. *British journal of pharmaceutical research*, 6(2), 68-75.
 17. Crisan, M. C., Pandrea, S. L., Matros, L., Mocan, T., & Mocan, L. (2024). In vitro antimicrobial activity of silver nanoparticles against selected Gram-negative and Gram-positive pathogens. *Medicine and Pharmacy Reports*, 97(3), 280.
 18. Holubnycha, V., Husak, Y., Korniienko, V., Bolshanina, S., Tveresovska, O., Myronov, P., & Pogorielov, M. (2024). Antimicrobial activity of two different types of silver nanoparticles against wide range of pathogenic bacteria. *Nanomaterials*, 14(2), 137.
 19. Rajendran, G., Rajamuthuramalingam, T., Jesse, D. M. I., & Kathiravan, K. (2019). Synthesis and characterization of biocompatible acetaminophen stabilized gold nanoparticles. *Materials Research Express*, 6(9), 095043.
 20. Sanchooli, N., Saeidi, S., Barani, H. K., & Sanchooli, E. (2018). In vitro antibacterial effects of silver nanoparticles synthesized using *Verbena officinalis* leaf extract on *Yersinia ruckeri*, *Vibrio cholera* and *Listeria monocytogenes*. *Iranian journal of microbiology*, 10(6), 400.
 21. Dozol, H., Méridet, G., Ancian, B., Cabuil, V., Xu, H., Wang, D., & Abou-Hassan, A. (2013). On the synthesis of Au nanoparticles using EDTA as a reducing agent. *The Journal of Physical Chemistry C*, 117(40), 20958-20966.
 22. Wan, J., Wang, J. H., Liu, T., Xie, Z., Yu, X. F., & Li, W. (2015). Surface chemistry but not aspect ratio mediates the biological toxicity of gold nanorods in vitro and in vivo. *Scientific reports*, 5(1), 11398.
 23. Bhaumik, J., Thakur, N. S., Aili, P. K., Ghanghoriya, A., Mittal, A. K., & Banerjee, U. C. (2015). Bioinspired nanotheranostic agents: synthesis, surface functionalization, and antioxidant potential. *ACS Biomaterials Science & Engineering*, 1(6), 382-392.
 24. Dwivedi, A. D., & Gopal, K. (2010). Biosynthesis of silver and gold nanoparticles using *Chenopodium album* leaf extract. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 369(1-3), 27-33.
 25. Gericke, M., & Pinches, A. (2006). Biological synthesis of metal nanoparticles.

- Hydrometallurgy, 83(1-4), 132-140.
26. Gautam, A., & van Veggel, F. C. (2013). Synthesis of nanoparticles, their biocompatibility, and toxicity behavior for biomedical applications. *Journal of Materials Chemistry B*, 1(39), 5186-5200.
27. Paul, P., Chatterjee, S., Pramanik, A., Karmakar, P., Chandra Bhattacharyya, S., & Kumar, G. S. (2018). Thionine conjugated gold nanoparticles trigger apoptotic activity toward HepG2 cancer cell line. *ACS Biomaterials Science & Engineering*, 4(2), 635-646
28. Muthukumarasamyvel, T., Rajendran, G., Santhana Panneer, D., Kasthuri, J., Kathiravan, K., & Rajendiran, N. (2017). Role of surface hydrophobicity of dicationic amphiphile-stabilized gold nanoparticles on A549 lung cancer cells. *ACS omega*, 2(7), 3527-3538.
29. Ramadurai, M., Rajendran, G., Bama, T. S., Prabhu, P., & Kathiravan, K. (2020). Biocompatible thiolate protected copper nanoclusters for an efficient imaging of lung cancer cells. *Journal of Photochemistry and Photobiology B: Biology*, 205, 111845.
30. Rajendran, K., Rajendran, G., Kasthuri, J., Kathiravan, K., & Rajendiran, N. (2019). Sweet corn (*Zea mays* L. var. *rugosa*) derived fluorescent carbon quantum dots for selective detection of hydrogen sulfide and bioimaging applications. *ChemistrySelect*, 4(46), 13668-13676.
31. Shahverdi, A. R., Fakhimi, A., Shahverdi, H. R., & Minaian, S. (2007). Synthesis and effect of silver nanoparticles on the antibacterial activity of different antibiotics against *Staphylococcus aureus* and *Escherichia coli*. *Nanomedicine: Nanotechnology, Biology and Medicine*, 3(2), 168-171.
32. Jyoti, K., Baunthiyal, M., & Singh, A. (2016). Characterization of silver nanoparticles synthesized using *Urtica dioica* Linn. leaves and their synergistic effects with antibiotics. *Journal of Radiation Research and Applied Sciences*, 9(3), 217-227.
33. Siddiqi, K. S., Husen, A., & Rao, R. A. (2018). A review on biosynthesis of silver nanoparticles and their biocidal properties. *Journal of nanobiotechnology*, 16, 1-28.
34. Banala, R. R., Nagati, V. B., & Karnati, P. R. (2015). Green synthesis and characterization of *Carica papaya* leaf extract coated silver nanoparticles through X-ray diffraction, electron microscopy and evaluation of bactericidal properties. *Saudi journal of biological sciences*, 22(5), 637-644.